Package 'lefser'

May 26, 2026

Type Package

Title R implementation of the LEfSE method for microbiome biomarker discovery

Description lefser is the R implementation of the popular microbiome biomarker discovery too, LEfSe. It uses the Kruskal-Wallis test, Wilcoxon-Rank Sum test, and Linear Discriminant Analysis to find biomarkers from two-level classes (and optional sub-classes).

Version 1.23.0

Date 2026-04-22

License Artistic-2.0

Depends SummarizedExperiment, R (>= 4.5.0)

Imports coin, MASS, ggplot2 (>= 3.4.0), S4Vectors, stats, methods, utils, dplyr, testthat, tibble, tidyr, forcats, stringr, ggtree, BiocGenerics, ape, ggrepel, mia, purrr, tidyselect, treeio

Suggests knitr, rmarkdown, curatedMetagenomicData, BiocStyle, phyloseq, pkgdown, covr, withr

Encoding UTF-8

BugReports https://github.com/waldronlab/lefser/issues

URL https://github.com/waldronlab/lefser

VignetteBuilder knitr

biocViews Software, Sequencing, DifferentialExpression, Microbiome, StatisticalMethod, Classification

RoxygenNote 7.3.3

Roxygen list(markdown = TRUE)

git_url https://git.bioconductor.org/packages/lefser

git_branch devel

git_last_commit a0a8ddd

git_last_commit_date 2026-05-14

Repository Bioconductor 3.24

Date/Publication 2026-05-25

Author Sehyun Oh [cre, ctb] (ORCID: <https://orcid.org/0000-0002-9490-3061>),
Asya Khleborodova [aut],
Samuel Gamboa-Tuz [ctb],
Marcel Ramos [ctb] (ORCID: <https://orcid.org/0000-0002-3242-0582>),
Ludwig Geistlinger [ctb] (ORCID:
<https://orcid.org/0000-0002-2495-5464>),
Levi Waldron [ctb] (ORCID: <https://orcid.org/0000-0003-2725-0694>),
NCI [fnd] (GrantNo.: R01CA230551)

Maintainer Sehyun Oh <shbrief@gmail.com>

Contents

`get_terminal_nodes` *Identify which elements of a string are terminal nodes*

Description

A terminal node in a taxonomy does not have any child nodes. For example, a species is a terminal node if there are no subspecies or strains that belong to that species. This function identifies which elements of a vector are terminal nodes simply by checking whether that element appears as a substring in any other element of the vector.

Usage

```
get_terminal_nodes(string)
```

Arguments

`string`	A character vector of strings to check for terminal nodes

Value

A logical vector indicating which elements of the string are terminal nodes

Examples

```
# What does it do?
data("zeller14")
rownames(zeller14)[988:989]
get_terminal_nodes(rownames(zeller14)[988:989])
# How do I use it to keep only terminal nodes for a lefser analysis?
terminal_nodes <- get_terminal_nodes(rownames(zeller14))
zeller14sub <- zeller14[terminal_nodes, ]
# Then continue with your analysis!
```

`lefser` *R implementation of the LEfSe method*

Description

Perform a LEfSe analysis: the function carries out differential analysis between two sample classes for multiple features and uses linear discriminant analysis to establish their effect sizes. Subclass information for each class can be incorporated into the analysis (see examples). Features with large differences between two sample classes are identified as biomarkers.

Usage

```
lefser(
  relab,
  kruskal.threshold = 0.05,
  wilcox.threshold = 0.05,
  lda.threshold = 2,
  classCol = "CLASS",
  subclassCol = NULL,
  assay = 1L,
  trim.names = FALSE,
  checkAbundances = TRUE,
  method = "none",
  ...
)
```

Arguments

`relab`	A SummarizedExperiment-class with relative abundances in the assay
`kruskal.threshold`	numeric(1) The p-value for the Kruskal-Wallis Rank Sum Test (default 0.05). If multiple hypothesis testing is performed, this threshold is applied to corrected p-values.
`wilcox.threshold`	numeric(1) The p-value for the Wilcoxon Rank-Sum Test when 'subclassCol' is present (default 0.05). If multiple hypothesis testing is performed, this threshold is applied to corrected p-values.
`lda.threshold`	numeric(1) The effect size threshold (default 2.0).
`classCol`	character(1) Column name in `colData(relab)` indicating class, usually a factor with two levels (e.g., `c("cases", "controls")`; default "CLASS").

subclassCol — character(1) Optional column name in colData(relab) indicating the subclasses, usually a factor with two levels (e.g., c("adult", "senior"); default NULL), but can be more than two levels.

assay — The i-th assay matrix in the SummarizedExperiment ('relab'; #' default 1).

trim.names — Default is FALSE. If TRUE, this function extracts the most specific taxonomic rank of organism.

checkAbundances — logical(1) Whether to check if the assay data in the relab input are relative abundances or counts. If counts are found, a warning will be emitted (default TRUE).

method — Default is "none" as in the original LEfSe implementation. Character string of length one, passed on to p.adjust to set option for multiple testing. For multiple pairwise comparisons, each comparison is adjusted separately. Options are "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr" (synonym for "BH"), and "none".

... — Additional inputs to lower level functions (not used).

Details

The LEfSe method expects relative abundances in the expr input. A warning will be emitted if the column sums do not result in 1. Use the relativeAb helper function to convert the data in the SummarizedExperiment to relative abundances. The checkAbundances argument enables checking the data for presence of relative abundances and can be turned off by setting the argument to FALSE.

Value

The function returns a data.frame with two columns, which are names of features and their LDA scores.

Examples

```
data(zeller14)
zeller14 <- zeller14[, zeller14$study_condition != "adenoma"]
tn <- get_terminal_nodes(rownames(zeller14))
zeller14tn <- zeller14[tn,]
zeller14tn_ra <- relativeAb(zeller14tn)

# (1) Using classes only
res_class <- lefser(relab = zeller14tn_ra, classCol = "study_condition")

# (2) Using classes and sub-classes
res_subclass <- lefser(
    relab = zeller14tn_ra,
    classCol = "study_condition",
    subclassCol = "age_category"
)
```

lefserClades *Run lefser at different clades*

Description

`lefesrCaldes` Agglomerates the features abundance at different taxonomic ranks using mia::splitByRanks and performs lefser at each rank. The analysis is run at the species, genus, family, order, class, and phylum levels.

Usage

```
lefserClades(relab, ...)
```

Arguments

`relab`	A (Tree) SummarizedExperiment with full taxonomy in the rowData @param ... Arguments passed to the `lefser` function.
`...`	Additional arguments passed to `lefser`

Details

When running `lefserClades`, all features with NAs in the rowData will be dropped. This is to avoid creating artificial clades with NAs.

Value

An object of class 'lefser_df_clades', "lefser_df", and 'data.frame'.

Examples

```
data("zeller14")
z14 <- zeller14[, zeller14$study_condition != "adenoma"]
tn <- get_terminal_nodes(rownames(z14))
z14tn <- z14[tn, ]
z14tn_ra <- relativeAb(z14tn)
z14_input <- rowNames2RowData(z14tn_ra)

resCl <- lefserClades(relab = z14_input, classCol = "study_condition")
```

lefserPlot *Plots results from* `lefser` *function*

Description

This function plots the biomarkers found by LEfSe, that are ranked according to their effect sizes and linked to their abundance in each class.

Usage

```
lefserPlot(
  df,
  colors = c("c", "l", "g"),
  trim.names = TRUE,
  title = "",
  label.font.size = 3,
  label.font.color = "black",
  label.font.face = c("plain", "italic", "bold", "bold.italic"),
  ...
)
```

Arguments

df	Data frame produced by `lefser`. This data frame contains two columns labeled as `c("features", "scores")`.
colors	Colors corresponding to class 0 and 1. Options: "c" (colorblind), "l" (lefse), "g" (greyscale). Defaults to "c". This argument also accepts a character(2) with two color names.
trim.names	Under the default (`TRUE`), this function extracts the most specific taxonomic rank of organism.
title	A character(1). The title of the plot.
label.font.size	A numeric(1). The font size of the feature labels. The default is `3`.
label.font.color	A character(1). The font color of the feature labels. The default is `"black"`.
label.font.face	A character(1). The font face of the feature labels. Options are "plain", "italic", "bold", or "bold.italic". The default is `"plain"`.
...	Additional arguments passed to `geom_text()`

Value

Function returns plot of effect size scores produced by `lefser`. Positive scores represent the biomarker is more abundant in class '1'. Negative scores represent the biomarker is more abundant in class '0'.

Examples

```
example("lefser")
lefserPlot(res_class)

# Plot with italicized feature labels
lefserPlot(res_class, label.font.face = "italic", label.font.color = "red")
```

`lefserPlotClad` *LEfSer plot cladogram*

Description

`lefserPlotClad` plots a cladogram from the results of `lefserClades`.

Usage

```
lefserPlotClad(
  df,
  colors = c("c", "l", "g"),
  showTipLabels = FALSE,
  showNodeLabels = "p"
)
```

Arguments

`df`	An object of class "lefesr_df_clades".
`colors`	Colors corresponding to class 0 and 1. Options: "c" (colorblind), "l" (lefse), "g" (greyscale). Defaults to "c". This argument also accepts a character(2) with two color names.
`showTipLabels`	Logical. If TRUE, show tip labels. Default is FALSE.
`showNodeLabels`	Label's to be shown in the tree. Options: "p" = phylum, "c" = class, "o" = order, "f" = family, "g" = genus, "s" = species, "t" = strain. It can accept several options, e.g., c("p", "c").

Value

A ggtree object.

Examples

```
data("zeller14")
z14 <- zeller14[, zeller14$study_condition != "adenoma"]
tn <- get_terminal_nodes(rownames(z14))
z14tn <- z14[tn, ]
z14tn_ra <- relativeAb(z14tn)
z14_input <- rowNames2RowData(z14tn_ra)

resCl <- lefserClades(relab = z14_input, classCol = "study_condition")
ggt <- lefserPlotClad(df = resCl)
```

`lefserPlotFeat` *Plot Feature*

Description

`lefserPlotFeat` plots the abundance data of a DA feature across all samples.

Usage

```
lefserPlotFeat(res, fName, colors = c("c", "l", "g"))
```

Arguments

`res`	An object of class lefser_df, output of the `lefser` function.
`fName`	A character string. The name of a feature in the lefser_df object.
`colors`	Colors corresponding to class 0 and 1. Options: "c" (colorblind), "l" (lefse), "g" (greyscale). Defaults to "c". This argument also accepts a character(2) with two color names.

Details

The solid lines represent the mean by class or by class+subclass (if the subclass variable is present). The dashed lines represent the median by class or by class+subclass (if the subclass variable is present).

Value

A ggplot object.

Examples

```
data(zeller14)
zeller14 <- zeller14[, zeller14$study_condition != "adenoma"]
tn <- get_terminal_nodes(rownames(zeller14))
zeller14tn <- zeller14[tn,]
zeller14tn_ra <- relativeAb(zeller14tn)

# (1) Using classes only
res_class <- lefser(zeller14tn_ra,
                    classCol = "study_condition")
# (2) Using classes and sub-classes
res_subclass <- lefser(zeller14tn_ra,
                    classCol = "study_condition",
                    subclassCol = "age_category")
plot_class <- lefserPlotFeat(res_class, res_class$features[[1]])
plot_subclass <- lefserPlotFeat(res_subclass, res_subclass$features[[2]])
```

`relativeAb` *Utility function to calculate relative abundances*

Description

The function calculates the column totals and divides each value within the column by the respective column total.

This function calculates the relative abundance of each feature in the SummarizedExperiment object containing count data, expressed as counts per million (CPM)

Usage

```
relativeAb(se, assay = 1L)
```

Arguments

`se`	A SummarizedExperiment object with counts
`assay`	The i-th assay matrix in the `SummarizedExperiment` ('relab'; #' default 1).

Value

returns a new SummarizedExperiment object with counts per million calculated and added as a new assay named rel_abs.

Examples

```
se <- SummarizedExperiment(
    assays = list(
        counts = matrix(
            rep(1, 4), ncol = 1, dimnames = list(LETTERS[1:4], "SAMP")
        )
    )
)
assay(se)
assay(relativeAb(se))
```

`rowNames2RowData` *RowNames to RowData*

Description

`rowNames2RowData` transforms the taxonomy stored in the row names to the rowData in a SummarizedExperiment.

Usage

```
rowNames2RowData(x)
```

Arguments

x	A SummarizedExperiment with the features taxonomy in the rownames.

Value

The same SummarizedExperiment with the taxonomy now in the rowData.

Examples

```
data("zeller14")

## Keep only "CRC" and "control" (dichotomous variable)
z14 <- zeller14[, zeller14$study_condition %in% c("control", "CRC")]

## Get terminal nodes
tn <- get_terminal_nodes(rownames(z14))
z14_tn <- z14[tn, ]

## Normalize to relative abundance (also known as Total Sum Scaling)
z14_tn_ra <- relativeAb(z14_tn)

## Add the taxonomy to the rowData
input_se <- rowNames2RowData(z14_tn_ra)
```

---

zeller14 *Example dataset for lefser*

Description

The ZellerG_2014 dataset contains microbiome count data for CRC patients and controls. It was for curatedMetagenomicData using the script in the package directory "data-raw".

Usage

```
data("zeller14")
```

Format

A SummarizedExperiment with 1585 features, 199 samples

study_condition adenoma, control, CRC

age_category adult, senior

Source

https://pubmed.ncbi.nlm.nih.gov/25432777/

Index