

# Package ‘iClusterPlus’

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**Description** Integrative clustering of multiple genomic data using a joint latent variable model.

**LazyData** yes

**LazyDataCompression** bzip2

**License** GPL (>= 2)

**biocViews** Multi-omics, Clustering

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breast.chr17	<i>Breast cancer data set DNA copy number and mRNA expression measure on chromosome 17</i>
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## Description

This is a subset of the breast cancer data from Pollack et al. (2002).

## Usage

```
data(breast.chr17)
```

## Format

A list object containing two data matrices: DNA and mRNA. They consist chromosome 17 data in 41 samples (4 cell lines and 37 primary tumors).

## Source

This data can be downloaded at <http://www.pnas.org/content/99/20/12963/suppl/DC1>

## References

Pollack, J.R. et al. (2002) Microarray analysis reveals a major direct role of DNA copy number alteration in the transcriptional program of human breast tumors. Proc. Natl Acad. Sci. USA, 99, 12963-12968.

---

CNregions

*A function to remove redundant copy number regions*

---

### Description

This function is used to reduce copy number regions.

### Usage

```
CNregions(seg, epsilon=0.005, adaptive=FALSE, rmCNV=FALSE, cnv=NULL,  
          frac.overlap=0.5, rmSmallseg=TRUE, nProbes=5)
```

### Arguments

seg	DNAcopy CBS segmentation output.
epsilon	the maximum Euclidean distance between adjacent probes tolerated for denying a nonredundant region. epsilon=0 is equivalent to taking the union of all unique break points across the n samples.
adaptive	Vector of length-m lasso penalty terms.
rmCNV	If TRUE, remove germline CNV.
cnv	A data frame containing germline CNV data.
frac.overlap	Fraction of overlap between 2 segments. If rmCNV=TRUE, overlapped segments will be removed if the overlapped fraction $\geq$ fra.overlap.
rmSmallseg	If TRUE, remove small segment.
nProbes	The segment length threshold below which the segment will be removed if rmSmallseg = TRUE.

### Value

A matrix with reduced copy number regions.

### Author(s)

Ronglai Shen <shenr@mskcc.org>

### References

Qianxing Mo, Sijian Wang, Venkatraman E. Seshan, Adam B. Olshen, Nikolaus Schultz, Chris Sander, R. Scott Powers, Marc Ladanyi, and Ronglai Shen. (2013). Pattern discovery and cancer gene identification in integrated cancer genomic data. Proc. Natl. Acad. Sci. USA.

### See Also

[breast.chr17,plotiCluster](#), [compute.pod,iCluster,iClusterPlus](#)

## Examples

```
#data(gbm)
#library(GenomicRanges)
#library(cluster)
#reducedM=CNregions(seg,epsilon=0,adaptive=FALSE,rmCNV=TRUE,cnv=NULL,
# frac.overlap=0.5, rmSmallseg=TRUE,nProbes=5)
```

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compute.pod	<i>A function to compute the proportion of deviation from perfect block diagonal matrix</i>
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## Description

A function to compute the proportion of deviation from perfect block diagonal matrix.

## Usage

```
compute.pod(fit)
```

## Arguments

fit	A iCluster object
-----	-------------------

## Value

pod	proportion of deviation from perfect block diagonal matrix
-----	--

## Author(s)

Ronglai Shen <shenr@mskcc.org>

## References

Ronglai Shen, Adam Olshen, Marc Ladanyi. (2009). Integrative clustering of multiple genomic data types using a joint latent variable model with application to breast and lung cancer subtype analysis. *Bioinformatics* 25, 2906-2912.

## See Also

[iCluster](#), [iCluster2](#), [plotiCluster](#)

## Examples

```
# library(iCluster)
# data(breast.chr17)
# fit=iCluster(breast.chr17, k=4, lambda=c(0.2,0.2))
# plotiCluster(fit=fit, label=rownames(breast.chr17[[2]]))
# compute.pod(fit)
```

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coord	<i>genomic coordinates</i>
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**Description**

genomic coordinates for the copy number data in gbm

**Usage**

data(coord)

**Format**

A data matrix consists of chr number, start and end position for the genes included in the gbm copy number data.

**References**

Ronglai Shen, Qianxing Mo, Nikolaus Schultz, Venkatraman E. Seshan, Adam B. Olshen, Jason Huse, Marc Ladanyi, Chris Sander. (2012). Integrative Subtype Discovery in Glioblastoma Using iCluster. *PLoS ONE* 7, e35236

---

gbm	<i>GBM data</i>
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**Description**

This is a subset of the glioblastoma dataset from the cancer genome atlas (TCGA) GBM study (2009) used in Shen et al. (2012).

**Usage**

data(gbm)

**Format**

A list object containing three data matrices: copy number, methylation and mRNA expression in 84 samples.

**Value**

gbm.seg	GBM copy number segmentation results generated by DNACopy package.
gbm.exp	GBM gene expression data.
gbm.mut	GBM mutation data.

**References**

Ronglai Shen, Qianxing Mo, Nikolaus Schultz, Venkatraman E. Seshan, Adam B. Olshen, Jason Huse, Marc Ladanyi, Chris Sander. (2012). Integrative Subtype Discovery in Glioblastoma Using iCluster. *PLoS ONE* 7, e35236

---

glp *good lattice points using the uniform design*

---

### Description

good lattice points using the uniform design (Fang and Wang 1995)

### Usage

```
data(glp)
```

### Format

A list object containing sampling design for  $s=2-5$  where  $s$  is the number of tuning parameters.

### References

Ronglai Shen, Qianxing Mo, Nikolaus Schultz, Venkatraman E. Seshan, Adam B. Olshen, Jason Huse, Marc Ladanyi, Chris Sander. (2012). Integrative Subtype Discovery in Glioblastoma Using iCluster. *PLoS ONE* 7, e35236

Fang K, Wang Y (1994) Number theoretic methods in statistics. London, UK: Chapman and Hall.

---

iCluster *Integrative clustering of multiple genomic data types*

---

### Description

Given multiple genomic data types (e.g., copy number, gene expression, DNA methylation) measured in the same set of samples, iCluster fits a regularized latent variable model based clustering that generates an integrated cluster assignment based on joint inference across data types

### Usage

```
iCluster(datasets, k, lambda, scalar=FALSE, max.iter=50, epsilon=1e-3)
```

### Arguments

datasets	A list object containing $m$ data matrices representing $m$ different genomic data types measured in a set of $n$ samples. For each matrix, the rows represent samples, and the columns represent genomic features.
k	Number of subtypes.
lambda	Vector of length- $m$ lasso penalty terms.
scalar	If TRUE, assumes scalar covariance matrix $\Psi$ . Default is FALSE.
max.iter	Maximum iteration for the EM algorithm.
epsilon	EM algorithm convergence criterion.

**Value**

A list with the following elements.

meanZ	Relaxed cluster indicator matrix.
beta	Coefficient matrix.
clusters	Cluster assignment.
conv.rate	Convergence history.

**Author(s)**

Ronglai Shen <shenr@mskcc.org>

**References**

Ronglai Shen, Adam Olshen, Marc Ladanyi. (2009). Integrative clustering of multiple genomic data types using a joint latent variable model with application to breast and lung cancer subtype analysis. *Bioinformatics* 25, 2906-2912.

**See Also**

[breast.chr17,plotiCluster](#), [compute.pod](#)

**Examples**

```
data(breast.chr17)
fit=iCluster(breast.chr17, k=4, lambda=c(0.2,0.2))
plotiCluster(fit=fit, label=rownames(breast.chr17[[2]]))
compute.pod(fit)

#library(gplots)
#library(lattice)
#col.scheme = alist()
#col.scheme[[1]] = bluered(256)
#col.scheme[[2]] = greenred(256)
#cn.image=breast.chr17[[2]]
#cn.image[cn.image>1.5]=1.5
#cn.image[cn.image< -1.5]= -1.5
#exp.image=breast.chr17[[1]]
#exp.image[exp.image>3]=3
#exp.image[exp.image< -3]=3
#plotHeatmap(fit, datasets=list(cn.image,exp.image), type=c("gaussian","gaussian"),
# row.order=c(FALSE,FALSE), width=5, col.scheme=col.scheme)
```

**Description**

Given multiple genomic data types (e.g., copy number, gene expression, DNA methylation) measured in the same set of samples, iCluster fits a regularized latent variable model based clustering that generates an integrated cluster assignment based on joint inference across data types

**Usage**

```
iCluster2(x, K, lambda, method=c("lasso","enet","flasso","glasso","gflasso"),
  chr=NULL, maxiter=50, eps=1e-4, eps2=1e-8)
```

**Arguments**

x	A list object containing m data matrices representing m different genomic data types measured in a set of n samples. For each matrix, the rows represent samples, and the columns represent genomic features.
K	Number of subtypes.
lambda	A list with m elements; each element is a vector with one or two elements depending on the methods used.
method	Method used for clustering and variable selection.
chr	Chromosome labels
maxiter	Maximum iteration for the EM algorithm.
eps	EM algorithm convergence criterion 1.
eps2	EM algorithm convergence criterion 2.

**Value**

A list with the following elements.

cluster	Cluster assignment.
centers	cluster centers.
Phivec	parameter phi; a vector.
beta	parameter B; a matrix.
meanZ	meanZ
EZZt	EZZt
dif	difference
iter	iteration

**Author(s)**

Qianxing Mo <qianxing.mo@moffitt.org>, Ronglai Shen, Sijian Wang

**References**

Ronglai Shen, Sijian Wang, Qianxing Mo. (2013). Sparse Integrative Clustering of Multiple Omics Data Sets. *Annals of Applied Statistics*. 7(1):269-294

**See Also**

[plotiCluster](#), [compute.pod](#), [iClusterPlus](#)

**Examples**

```

## clustering
n1 = 20
n2 = 20
n3 = 20
n = n1+n2+n3
p = 5
q = 100

x = NULL
x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[1]] = cbind(xa,xb)

x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[2]] = cbind(xa,xb)

x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[3]] = cbind(xa,xb)

x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[4]] = cbind(xa,xb)

x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[5]] = cbind(xa,xb)

method = c('lasso', 'enet', 'flasso', 'glasso', 'gflasso')
lambda=alist()
lambda[[1]] = 30
lambda[[2]] = c(20,1)
lambda[[3]] = c(20,20)
lambda[[4]] = 30
lambda[[5]] = c(30,20)

chr=c(rep(1,10),rep(2,(p+q)-10))
date()

```

```

fit2 = iCluster2(x, K=3, lambda, method=method, chr=chr, maxiter=20,eps=1e-4, eps2=1e-8)
date()

par(mfrow=c(5,1),mar=c(4,4,1,1))
for(i in 1:5){
  barplot(fit2$beta[[i]][,1])
}

#library(gplots)
#library(lattice)

#plotHeatmap(fit2, datasets=x, type=rep("gaussian",length(x)),
  #row.order=c(TRUE,TRUE,FALSE,TRUE,FALSE),
  #sparse=rep(FALSE,length(x)), scale=rep("row",5), width=5,
  #col.scheme=rep(list(bluered(256)),length(x)))

```

---

iCluster2b

*Integrative clustering of multi-omics data*


---

## Description

Given multi-omics data (e.g., copy number, gene expression, DNA methylation) measured in the same set of samples, iCluster2b fits a regularized latent factor(variable) model that generates an latent factor matrix that can be used for integrative clustering of samples. In addition, the driver features of sample clustering can be identified by sparse coefficient matrices.

## Usage

```

iCluster2b(xList,K=3, lambda, method=c("lasso","enet","flasso","glasso","gflasso"),
  chr=NULL, EM.iter=25, eps=1e-4, eps2=1e-6)

```

## Arguments

xList	A list object containing m data matrices representing m multi-omics data measured in a set of n samples. For each matrix, the rows represent samples, and the columns represent genomic features.
K	An integer representing the number of the latent factors used for modeling.
lambda	A list with m elements corresponding to the method argument. For lasso and glasso, each element is a positive value. For enet, flasso and gflasso, each element is a vector with two positive values.
method	Method used for variable selection.
chr	Chromosome labels
EM.iter	Maximum iteration for the EM algorithm.
eps	EM algorithm convergence criterion 1.
eps2	EM algorithm convergence criterion 2.

**Value**

A list with the following elements.

meanZ	A $n \times k$ matrix; the rows represent samples and the columns represent the $K$ factors.
beta	A list object containing $m$ coefficient matrices corresponding to the $m$ data matrices.
iter	EM iteration.

**Author(s)**

Qianxing Mo <qianxing.mo@moffitt.org>, Ronglai Shen, Sijian Wang

**References**

Ronglai Shen, Sijian Wang, Qianxing Mo. (2013). Sparse Integrative Clustering of Multiple Omics Data Sets. *Annals of Applied Statistics*. 7(1):269-294

**See Also**

[tune.iCluster2b](#), [iClusterPlus2](#), [iClusterBayes](#)

**Examples**

```
## clustering
n1 = 20
n2 = 20
n3 = 20
n = n1+n2+n3
p = 5
q = 100

x = NULL
x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[1]] = cbind(xa,xb)

x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[2]] = cbind(xa,xb)

x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[3]] = cbind(xa,xb)
```

```

x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[4]] = cbind(xa,xb)

x1a = matrix(rnorm(n1*p), ncol=p)
x2a = matrix(rnorm(n1*p, -1.5,1), ncol=p)
x3a = matrix(rnorm(n1*p, 1.5, 1), ncol=p)
xa = rbind(x1a,x2a,x3a)
xb = matrix(rnorm(n*q), ncol=q)
x[[5]] = cbind(xa,xb)

method = c('lasso', 'enet', 'flasso', 'glasso', 'gflasso')
lambda=alist()
lambda[[1]] = 30
lambda[[2]] = c(20,1)
lambda[[3]] = c(20,20)
lambda[[4]] = 30
lambda[[5]] = c(30,20)

chr=c(rep(1,10),rep(2,(p+q)-10))
date()
fit2 = iCluster2b(x, K=3, lambda, method=method, chr=chr, EM.iter=20,eps=1e-4, eps2=1e-6)
date()

par(mfrow=c(5,1),mar=c(4,4,1,1))
for(i in 1:5){
  barplot(fit2$beta[[i]][,1])
}

```

---

iClusterBayes

*Integrative clustering of multiple genomic data types*


---

## Description

Given multiple genomic data types (e.g., copy number, gene expression, DNA methylation) measured in the same set of samples, iClusterBayes fits a Bayesian latent variable model that generates an integrated cluster assignment based on joint inference across data types and identifies genomic features that contribute to the clusters.

## Usage

```

iClusterBayes(dt1,dt2=NULL,dt3=NULL,dt4=NULL,dt5=NULL,dt6=NULL,
  type = c("gaussian","binomial","poisson"),K=2,n.burnin=1000,n.draw=1200,
  prior.gamma=rep(0.1,6),sdev=0.5,beta.var.scale=1,thin=1,pp.cutoff=0.5)

```

## Arguments

dt1                      Data set 1 - a matrix with rows and columns representing samples and genomic features, respectively.

dt2	Data set 2 - a matrix with rows and columns representing samples and genomic features, respectively.
dt3	Data set 3 - a matrix with rows and columns representing samples and genomic features, respectively.
dt4	Data set 4 - a matrix with rows and columns representing samples and genomic features, respectively.
dt5	Data set 5 - a matrix with rows and columns representing samples and genomic features, respectively.
dt6	Data set 6 - a matrix with rows and columns representing samples and genomic features, respectively.
type	Data type corresponding to dt1-6, which can be gaussian, binomial, or poisson.
K	The number of eigen features. Given K, the number of cluster is K+1.
n.burnin	Number of MCMC burnin.
n.draw	Number of MCMC draw.
prior.gamma	Prior probability for the indicator variable gamma of each data set.
sdev	Standard deviation of random walk proposal for the latent variable.
beta.var.scale	A positive value to control the scale of covariance matrix of the proposed beta.
thin	A parameter to thin the MCMC chain in order to reduce autocorrelation. Discard all but every 'thin'th sampling values. When thin=1, all sampling values are kept.
pp.cutoff	Posterior probability cutoff for the indicator variable gamma. The BIC and deviance ratio will be calculated by setting parameter beta to zero when the posterior probability of gamma <= cutoff.

### Value

A list with the following elements.

alpha	Intercept parameter.
beta	Information parameter.
beta.pp	Posterior probability of beta. The higher the beta.pp, the more likely the beta should be included in the model.
gamma.ar	Acceptance ratio for the parameter gamma.
beta.ar	Acceptance ratio for the parameter beta.
Z.ar	Acceptance ratio for the latent variable.
clusters	Cluster assignment.
centers	Cluster center.
meanZ	The latent variable.
BIC	Bayesian information criterion.
dev.ratio	see dev.ratio defined in glmnet package.

### Author(s)

Qianxing Mo <qianxing.mo@moffitt.org>

## References

Mo Q, Shen R, Guo C, Vannucci M, Chan KS, Hilsenbeck SG. (2018). A fully Bayesian latent variable model for integrative clustering analysis of multi-type omics data. *Biostatistics* 19(1):71-86.

## See Also

[tune.iClusterBayes](#), [plotHMBayes](#), [iClusterPlus](#), [tune.iClusterPlus](#), [plotHeatmap](#)

## Examples

```
# see iManual.pdf
```

---

iClusterBayes2	<i>Integrative clustering of multi-omics data</i>
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---

## Description

Given multi-omics data (e.g., Somatic mutation, copy number, gene expression, DNA methylation) measured in the same set of samples, iClusterBayes2 fits a Bayesian latent factor model that generates an latent factor matrix that can be used for integrative clustering of samples. In addition, the driver features of sample clustering can be identified by the posterior probability of the model parameters.

## Usage

```
iClusterBayes2(xList, type = c("gaussian", "binomial", "poisson"), K=3, n.burnin=1000, n.draw=1200,
  prior.gamma=rep(0.1, 6), sdev=0.5, beta.var.scale=1, thin=1, pp.cutoff=0.5)
```

## Arguments

xList	A list object containing m data matrices representing m multi-omics data measured in a set of n samples. For each matrix, the rows represent samples, and the columns represent genomic features. The allowed maximum number of data matrices is 6 ( $m < 7$ ).
type	Data type corresponding to the data matrices in xList, which can be gaussian, binomial, or poisson.
K	An integer representing the number of the latent factors used for modeling.
n.burnin	Number of MCMC burnin.
n.draw	Number of MCMC draw.
prior.gamma	Prior probability for the indicator variable gamma of each data set.
sdev	Standard deviation of random walk proposal for the latent variable.
beta.var.scale	A positive value to control the scale of covariance matrix of the proposed beta.
thin	A parameter to thin the MCMC chain in order to reduce autocorrelation. Discard all but every 'thin'th sampling values. When thin=1, all sampling values are kept.
pp.cutoff	Posterior probability cutoff for the indicator variable gamma. The BIC and deviance ratio will be calculated by setting parameter beta to zero when the posterior probability of gamma $\leq$ cutoff.

**Value**

A list with the following elements.

alpha	A list of object for the intercept parameters corresponding to the m data matrices.
beta	A list object containing m coefficient matrices corresponding to the m data matrices.
meanZ	A n x k matrix; the rows represent samples and the columns represent the K factors.
beta.pp	A list of posterior probability for the parameter beta. The higher the beta.pp, the more likely the beta should be included in the model.
gamma.ar	A list of acceptance ratio for the parameter gamma.
beta.ar	A list of acceptance ratio for the parameter beta.
Z.ar	Acceptance ratio for the latent variable.
BIC	Bayesian information criterion.
dev.ratio	see dev.ratio defined in glmnet package.

**Author(s)**

Qianxing Mo <qianxing.mo@moffitt.org>

**References**

Mo Q, Shen R, Guo C, Vannucci M, Chan KS, Hilsenbeck SG. (2018). A fully Bayesian latent variable model for integrative clustering analysis of multi-type omics data. *Biostatistics* 19(1):71-86.

**See Also**

[iClusterPlus2,plotHMBayes,plotHeatmap](#)

**Examples**

```
# see iManual.pdf
```

---

iClusterPlus

*Integrative clustering of multiple genomic data types*

---

**Description**

Given multiple genomic data types (e.g., copy number, gene expression, DNA methylation) measured in the same set of samples, iClusterPlus fits a regularized latent variable model based clustering that generates an integrated cluster assignment based on joint inference across data types

**Usage**

```
iClusterPlus(dt1,dt2=NULL,dt3=NULL,dt4=NULL,
  type=c("gaussian","binomial","poisson","multinomial"),
  K=2,alpha=c(1,1,1,1),lambda=c(0.03,0.03,0.03,0.03),
  n.burnin=100,n.draw=200,maxiter=20,sdev=0.05,eps=1.0e-4)
```

**Arguments**

dt1	A data matrix. The rows represent samples, and the columns represent genomic features.
dt2	A data matrix. The rows represent samples, and the columns represent genomic features.
dt3	A data matrix. The rows represent samples, and the columns represent genomic features.
dt4	A data matrix. The rows represent samples, and the columns represent genomic features.
type	Data type, which can be gaussian, binomial, poisson, multinomial.
K	The number of eigen features. Given K, the number of cluster is K+1.
alpha	Vector of elasticnet penalty terms. At this version of iClusterPlus, elasticnet is not used. Therefore, all the elements of alpha are set to 1.
lambda	Vector of lasso penalty terms.
n.burnin	Number of MCMC burnin.
n.draw	Number of MCMC draw.
maxiter	Maximum iteration for the EM algorithm.
sdev	standard deviation of random walk proposal.
eps	Algorithm convergence criterion.

**Value**

A list with the following elements.

alpha	Intercept parameter.
beta	Information parameter.
clusters	Cluster assignment.
centers	Cluster center.
meanZ	Latent variable.
BIC	Bayesian information criterion.
dev.ratio	see dev.ratio defined in glmnet package.
dif	absolute difference for the parameters in the last and next-to-last iterations.

**Author(s)**

Qianxing Mo <qianxing.mo@moffitt.org>, Ronglai Shen, Sijian Wang

**References**

Qianxing Mo, Sijian Wang, Venkatraman E. Seshan, Adam B. Olshen, Nikolaus Schultz, Chris Sander, R. Scott Powers, Marc Ladanyi, and Ronglai Shen. (2013). Pattern discovery and cancer gene identification in integrated cancer genomic data. *Proc. Natl. Acad. Sci. USA*. 110(11):4245-50.

**See Also**

[plotiCluster](#), [iCluster](#), [compute.pod](#)

**Examples**

```
# see iManual.pdf
```

---

iClusterPlus2	<i>Integrative clustering of multi-omics data</i>
---------------	---

---

**Description**

Given multi-omics data (e.g., somatic mutation, copy number, gene expression, DNA methylation) measured in the same set of samples, iClusterPlus2 fits a regularized latent factor(variable) model that generates an latent factor matrix that can be used for integrative clustering of samples. In addition, the driver features of sample clustering can be identified by sparse coefficient matrices.

**Usage**

```
iClusterPlus2(xList, type=c("gaussian", "binomial", "poisson", "multinomial"), K=3,
              n.burnin=100, n.draw=200, maxiter=25, sdev=0.05, lambda.scale=1/3,
              BICrate.cutoff=rep(0.01, 4), min.shrinkage.rate=rep(0.05, 4))
```

**Arguments**

<code>xList</code>	A list object containing $m$ data matrices representing $m$ multi-omics data measured in a set of $n$ samples. For each matrix, the rows represent samples, and the columns represent genomic features.
<code>type</code>	Data type, which can be gaussian, binomial, poisson, multinomial.
<code>K</code>	An integer representing the number of the latent factors used for modeling.
<code>n.burnin</code>	Number of MCMC burnin.
<code>n.draw</code>	Number of MCMC draw.
<code>maxiter</code>	Maximum iteration for the EM algorithm.
<code>sdev</code>	standard deviation of random walk proposal.
<code>lambda.scale</code>	scaling factor for the lasso regularization parameter when the data type is binomial. Empirically, <code>lambda.scale</code> is within (0.1, 1).
<code>BICrate.cutoff</code>	BIC rate between iteration $i+1$ and $i$ . If BIC rate $<$ BICrate.cutoff, the search for optimal lambda will stop.
<code>min.shrinkage.rate</code>	The minimum lasso shrinkage rates for multi-omics features.

**Value**

A list with the following elements.

<code>alpha</code>	A list of object for the intercept parameters corresponding to the $m$ data matrices.
<code>beta</code>	A list object containing $m$ coefficient matrices corresponding to the $m$ data matrices.
<code>meanZ</code>	A $n \times k$ matrix; the rows represent samples and the columns represent the $K$ factors.
<code>BIC</code>	Bayesian information criterion.
<code>dev.ratio</code>	see <code>dev.ratio</code> defined in <code>glmnet</code> package.
<code>lambda</code>	Final lasso regularization parameters used for iCluster modeling.

**Author(s)**

Qianxing Mo <qianxing.mo@moffitt.org>

**References**

Qianxing Mo, Sijian Wang, Venkatraman E. Seshan, Adam B. Olshen, Nikolaus Schultz, Chris Sander, R. Scott Powers, Marc Ladanyi, and Ronglai Shen. (2013). Pattern discovery and cancer gene identification in integrated cancer genomic data. *Proc. Natl. Acad. Sci. USA*. 110(11):4245-50.

**See Also**

[tune.iCluster2b](#), [iClusterPlus](#), [iClusterBayes](#)

**Examples**

```
# see iManual.pdf
```

---

pcaVarPlot

*A function to make PCA variance plot of the combined matrix of multi-omics data matrices.*

---

**Description**

Multi-omics data matrices are column-combined and then the PCA variance plot of the combined matrix is made.

**Usage**

```
pcaVarPlot(xList,K=10)
```

**Arguments**

**xList** A list object containing m data matrices representing m multi-omics data measured in a set of n samples. For each matrix, the rows represent samples, and the columns represent genomic features.

**K** The number of principle components.

**Value**

no value returned.

**Author(s)**

Qianxing Mo <qianxing.mo@moffitt.org>

**References**

# TBD

**See Also**

[tune.iCluster2b](#), [iClusterPlus2](#), [iClusterBayes](#), [plotHeatmap](#)

**Examples**

```
# see tutorials
```

---

plotHeatmap	<i>A function to generate heatmap panels sorted by integrated cluster assignment.</i>
-------------	---

---

**Description**

A function to generate heatmap panels sorted by integrated cluster assignment.

**Usage**

```
plotHeatmap(fit, xList, type=c("gaussian", "binomial", "poisson", "multinomial"), sample.order=NULL,
            dist.method = "euclidean", hclust.method="ward.D", sparse=NULL, threshold=rep(0.25, length(
            feature.scale=rep(F, length(xList))), col.scheme=rep(list(bluered(256)), length(xList)), wi
            chr=NULL, plot.chr=NULL, cap=rep(0, length(xList)))
```

**Arguments**

fit	A iCluster object.
xList	A list object of data matrices.
type	Types of data in the xList.
sample.order	User supplied cluster assignment.
feature.order	A vector of logical values each specify whether the genomic features in the corresponding data matrix should be reordered by similarity. Default is TRUE.
dist.method	Method used to calculate distance (similarity) between features. Default method is "euclidean". Another choice is "correlation", which is Pearson correlation coefficient.
hclust.method	Method passed to hclust function. See hclust for details.
sparse	A vector of logical values each specify whether to plot the top cluster-discriminant features. Default is FALSE.
threshold	When sparse is TRUE, a vector of threshold values to include the genomic features for which the absolute value of the associated coefficient estimates fall in the top quantile. threshold=c(0.25,0.25) takes the top quartile most discriminant features in data type 1 and data type 2 for plot.
feature.scale	A vector of logical values each specify whether data should be scaled. Default is FALSE.
col.scheme	Color scheme. Can use bluered(n) in gplots R package.
width	Width of the figure in inches.
chr	A vector of chromosome number.
plot.chr	A vector of logical values each specify whether to annotate chromosome number on the left of the panel. Typically used for copy number data type. Default is FALSE.
cap	A numeric vector used to control the heatmap colors. For example, cap=c(0,0.0.95,0.95) indicates that no truncation for the image data used to make heatmap 1 data, and the data used to make heatmaps 2 and 3 are truncated at 95% quantile.

**Details**

The samples are ordered by the cluster assignment using the R code: `order(fit$clusters)`. For each data set, the features are ordered by hierarchical clustering of the features using the `hclust.method` and euclidean (or 1-correlation coefficient) as the distance.

**Value**

`feature.hclust` A list of objects returned by "hclust" that describes the tree produced by the clustering process. For a given data matrix, if `feature.order` is TRUE, the features of the data matrix are ordered by the tree generated by "hclust". Please see "hclust" for details.

`image.data` A list of data matrices used to make the heatmaps.

**Author(s)**

Ronglai Shen <shenr@mskcc.org>; Qianxing Mo <qianxing.mo@moffitt.org>

**References**

Ronglai Shen, Adam Olshen, Marc Ladanyi. (2009). Integrative clustering of multiple genomic data types using a joint latent variable model with application to breast and lung cancer subtype analysis. *Bioinformatics* 25, 2906-2912.

Ronglai Shen, Qianxing Mo, Nikolaus Schultz, Venkatraman E. Seshan, Adam B. Olshen, Jason Huse, Marc Ladanyi, Chris Sander. (2012). Integrative Subtype Discovery in Glioblastoma Using iCluster. *PLoS ONE* 7, e35236

**See Also**

[iCluster](#), [iCluster2b](#), [iClusterPlus2](#)

**Examples**

```
# see iManual.pdf
```

---

plotHMBayes	<i>A function to generate heatmap panels sorted by integrated cluster assignment.</i>
-------------	---

---

**Description**

A function to generate heatmap panels sorted by integrated cluster assignment.

**Usage**

```
plotHMBayes(fit, xList, type = c("gaussian", "binomial", "poisson"),
  sample.order = NULL, feature.order = NULL, dist.method="euclidean", hclust.method="ward.D",
  sparse = NULL, threshold = rep(0.5,length(xList)), feature.scale = rep(F,length(xList)),
  col.scheme = rep(list(bluered(256)),length(xList)), width=5, chr=NULL,
  plot.chr=NULL, cap=rep(0,length(xList)))
```

**Arguments**

<code>fit</code>	A <code>iClusterBayes</code> object.
<code>xList</code>	A list object of data matrices.
<code>type</code>	Types of data in the <code>xList</code> .
<code>sample.order</code>	User supplied cluster assignment.
<code>feature.order</code>	A vector of logical values each specify whether the genomic features in the corresponding data matrix should be reordered by similarity. Default is <code>TRUE</code> .
<code>dist.method</code>	Method used to calculate distance (similarity) between features. Default method is "euclidean". Another choice is "correlation", which is Pearson correlation coefficient.
<code>hclust.method</code>	Method passed to <code>hclust</code> function. See <code>hclust</code> for details.
<code>sparse</code>	A vector of logical values each specify whether to plot the top cluster-discriminant features. Default is <code>FALSE</code> .
<code>threshold</code>	When <code>sparse</code> is <code>TRUE</code> , a vector of threshold values to include the genomic features on the heatmap. Each data set should have a threshold. For each data set, a feature with posterior probability greater than the threshold will be included. Default value is 0.5 for each data set.
<code>feature.scale</code>	A vector of logical values each specify whether data should be scaled. Default is <code>FALSE</code> .
<code>col.scheme</code>	Color scheme. Can use <code>bluered(n)</code> in <code>gplots</code> R package.
<code>width</code>	Width of the figure in inches.
<code>chr</code>	A vector of chromosome number.
<code>plot.chr</code>	A vector of logical values each specify whether to annotate chromosome number on the left of the panel. Typically used for copy number data type. Default is <code>FALSE</code> .
<code>cap</code>	A numeric vector used to control the heatmap colors. For example, <code>cap=c(0,0.0.95,0.95)</code> indicates that no truncation for the image data used to make heatmap 1 data, and the data used to make heatmaps 2 and 3 are truncated at 95% quantile.

**Details**

The samples are ordered by the cluster assignment using the R code: `order(fit$clusters)`. For each data set, the features are ordered by hierarchical clustering of the features using the `hclust.method` and euclidean (or 1-correlation coefficient) as the distance.

**Value**

<code>feature.hclust</code>	A list of objects returned by "hclust" that describes the tree produced by the clustering process. For a given data matrix, if <code>feature.order</code> is <code>TRUE</code> , the features of the data matrix are ordered by the tree generated by "hclust". Please see "hclust" for details.
<code>image.data</code>	A list of data matrices used to make the heatmaps.
	no value returned.

**Author(s)**

Ronglai Shen <shenr@mskcc.org>, Qianxing Mo <qianxing.mo@moffitt.org>

## References

Mo Q, Shen R, Guo C, Vannucci M, Chan KS, Hilsenbeck SG. (2018). A fully Bayesian latent variable model for integrative clustering analysis of multi-type omics data. *Biostatistics* 19(1):71-86.

## See Also

[iClusterBayes](#), [plotHeatmap](#)

## Examples

```
# see iManual.pdf
```

---

plotiCluster	<i>A function to generate cluster separability matrix plot.</i>
--------------	---

---

## Description

A function to generate cluster separability matrix plot.

## Usage

```
plotiCluster(fit, label=NULL)
```

## Arguments

fit	A iCluster object
label	Sample labels

## Value

no value returned.

## Author(s)

Ronglai Shen <shenr@mskcc.org>

## References

Ronglai Shen, Adam Olshen, Marc Ladanyi. (2009). Integrative clustering of multiple genomic data types using a joint latent variable model with application to breast and lung cancer subtype analysis. *Bioinformatics* 25, 2906-2912.

## See Also

[iCluster](#), [compute.pod](#)

## Examples

```
# library(iCluster)
# data(breast.chr17)
# fit=iCluster(datasets=breast.chr17, k=4, lambda=c(0.2,0.2))
# plotiCluster(fit=fit, label=rownames(breast.chr17[[2]]))
# compute.pod(fit)
```

---

plotRI

*A function to generate reproducibility index plot.*

---

### Description

A function to generate reproducibility index plot.

### Usage

```
plotRI(cv.fit)
```

### Arguments

`cv.fit`            A `tune.iCluster2` object

### Value

no value returned.

### Author(s)

Ronglai Shen <shenr@mskcc.org>

### References

Ronglai Shen, Adam Olshen, Marc Ladanyi. (2009). Integrative clustering of multiple genomic data types using a joint latent variable model with application to breast and lung cancer subtype analysis. *Bioinformatics* 25, 2906-2912.

Ronglai Shen, Qianxing Mo, Nikolaus Schultz, Venkatraman E. Seshan, Adam B. Olshen, Jason Huse, Marc Ladanyi, Chris Sander. (2012). Integrative Subtype Discovery in Glioblastoma Using iCluster. *PLoS ONE* 7, e35236

### See Also

[iCluster](#)

### Examples

```
#data(simu.datasets)
#cv.fit=alist()
#for(k in 2:5){
#  cat(paste("K=",k,sep=""),'\n')
#  cv.fit[[k]]=tune.iCluster2(datasets=simu.datasets, k,nrep=2, n.lambda=8)
#}

##Reproducibility index (RI) plot
#plotRI(cv.fit)
```

---

tune.iCluster2      *Integrative clustering of multiple genomic data types*

---

### Description

Given multiple genomic data types (e.g., copy number, gene expression, DNA methylation) measured in the same set of samples, iCluster fits a regularized latent variable model based clustering that generates an integrated cluster assignment based on joint inference across data types

### Usage

```
tune.iCluster2(x, K, method=c("lasso", "enet", "flasso", "glasso", "gflasso"), base=200,
  chr=NULL, true.class=NULL, lambda=NULL, n.lambda=NULL, save.nonsparse=F, nrep=10, eps=1e-4)
```

### Arguments

x	A list object containing m data matrices representing m different genomic data types measured in a set of n samples. For each matrix, the rows represent samples, and the columns represent genomic features.
K	Number of subtypes.
lambda	User supplied matrix of lambda to tune.
method	Method used for clustering and variable selection.
chr	Chromosome labels
n.lambda	Number of lambda to sample using uniform design.
nrep	Fold of cross-validation.
base	Base.
true.class	True class label if available.
save.nonsparse	Logic argument whether to save the nonsparse fit.
eps	EM algorithm convergence criterion

### Value

A list with the following elements.

best.fit	Best fit.
best.lambda	Best lambda.
ps	Rand index
ps.adjusted	Adjusted Rand index.

### Author(s)

Qianxing Mo <qianxing.mo@moffitt.org>, Ronglai Shen, Sijian Wang

### References

Ronglai Shen, Sijian Wang, Qianxing Mo. (2013). Sparse Integrative Clustering of Multiple Omics Data Sets. *Annals of Applied Statistics*. 7(1):269-294

### See Also

[iCluster2](#)

---

tune.iCluster2b      *Integrative clustering of multi-omics data*

---

### Description

This function finds optimal lasso regularization parameters for iCluster2b.

### Usage

```
tune.iCluster2b(xList,K=3,method=c("lasso","enet"),min.lambda=10,max.lambda=500,
  lambda.iter=25,EM.iter=25,min.shrinkage.rate=rep(0.05,length(xList)), eps=1e-4, eps2=1e-6)
```

### Arguments

xList	A list object containing m data matrices representing m multi-omics data measured in a set of n samples. For each matrix, the rows represent samples, and the columns represent genomic features.
K	An positive integer representing the number of the latent factors used for modeling.
method	Method used for the regularization of model parameters.
min.lambda	The minimum value of the lasso regularization parameter.
max.lambda	The maximum value of the lasso regularization parameter.
lambda.iter	the number of iteration to find an optimal lambda.
EM.iter	The number of iteration for the EM algorithm.
min.shrinkage.rate	The minimum lasso shrinkage rates for multi-omics features.
eps	EM algorithm convergence criterion 1.
eps2	EM algorithm convergence criterion 2.

### Value

A list with the following elements.

meanZ	A n x k matrix; the rows represent samples and the columns represent the K factors.
beta	A list object containing m coefficient matrices corresponding to the m data matrices.
lambda	Final lasso regularization parameters used for iCluster modeling.

### Author(s)

Qianxing Mo <qianxing.mo@moffitt.org>

### References

Ronglai Shen, Sijian Wang, Qianxing Mo. (2013). Sparse Integrative Clustering of Multiple Omics Data Sets. *Annals of Applied Statistics*. 7(1):269-294

### See Also

[iCluster2b](#), [iClusterPlus2](#), [iClusterBayes](#)

---

tune.iClusterBayes      *Integrative clustering of multiple genomic data*

---

### Description

In order to determining the appropriate number of clusters, tune.iClusterBayes calls iClusterBayes function and performs parallel computation for  $K=1,2,\dots$

### Usage

```
tune.iClusterBayes(cpus=6,dt1,dt2=NULL,dt3=NULL,dt4=NULL,dt5=NULL,dt6=NULL,
  type=c("gaussian","binomial","poisson"),
  K=1:6,n.burnin=1000,n.draw=1200,prior.gamma=rep(0.1,6),
  sdev=0.5,beta.var.scale=1,thin=1,pp.cutoff=0.5)
```

### Arguments

cpus	Number of CPU used for parallel computation. If possible, let it be equal to the number of Ks.
dt1	Data set 1 - a matrix with rows and columns representing samples and genomic features, respectively.
dt2	Data set 2 - a matrix with rows and columns representing samples and genomic features, respectively.
dt3	Data set 3 - a matrix with rows and columns representing samples and genomic features, respectively.
dt4	Data set 4 - a matrix with rows and columns representing samples and genomic features, respectively.
dt5	Data set 5 - a matrix with rows and columns representing samples and genomic features, respectively.
dt6	Data set 6 - a matrix with rows and columns representing samples and genomic features, respectively.
type	Data type corresponding to dt1-6, which can be gaussian, binomial, poisson.
K	A vector. Each element is the number of eigen features. Given k, the number of cluster is k+1.
n.burnin	Number of MCMC burnin.
n.draw	Number of MCMC draw.
prior.gamma	Prior probability for the indicator variable gamma of each data set.
sdev	Standard deviation of random walk proposal for the latent variable.
beta.var.scale	A positive value to control the scale of covariance matrix of the proposed beta.
thin	A parameter to thin the MCMC chain in order to reduce autocorrelation. Discard all but every 'thin'th sampling values. When thin=1, all sampling values are kept.
pp.cutoff	Posterior probability cutoff for the indicator variable gamma. The BIC and deviance ratio will be calculated by setting parameter beta to zero when the posterior probability of gamma $\leq$ cutoff.

**Value**

A list named 'fit'. fit[[i]] is an object return by iClusterBayes, corresponding to the ith element in K. Each component of fit has the following elements.

alpha	Intercept parameter.
beta	Information parameter.
beta.pp	Posterior probability of beta. The higher the beta.pp, the more likely the beta should be included in the model.
gamma.ar	Acceptance ratio for parameter gamma.
beta.ar	Acceptance ratio for parameter beta.
Z.ar	Acceptance ratio for the latent variable.
clusters	Cluster assignment.
centers	Cluster center.
meanZ	Latent variable.
BIC	Bayesian information criterion.
dev.ratio	See dev.ratio defined in glmnet package.

**Author(s)**

Qianxing Mo <qianxing.mo@moffitt.org>

**References**

Mo Q, Shen R, Guo C, Vannucci M, Chan KS, Hilsenbeck SG. (2018). A fully Bayesian latent variable model for integrative clustering analysis of multi-type omics data. *Biostatistics* 19(1):71-86.

**See Also**

[iClusterBayes](#), [plotHMBayes](#), [iClusterPlus](#), [tune.iClusterPlus](#), [plotHeatmap](#)

**Examples**

```
### see the users' guide iManu1.pdf
```

---

tune.iClusterPlus      *Integrative clustering of multiple genomic data*

---

**Description**

Given multiple genomic data (e.g., copy number, gene expression, DNA methylation) measured in the same set of samples, tune.iClusterPlus uses a series of lambda values to fit a regularized latent variable model based clustering that generates an integrated cluster assignment based on joint inference across data.

**Usage**

```
tune.iClusterPlus(cpus=8,dt1,dt2=NULL,dt3=NULL,dt4=NULL,
  type=c("gaussian","binomial","poisson","multinomial"),
  K=2,alpha=c(1,1,1,1),n.lambda=NULL,scale.lambda=c(1,1,1,1),
  n.burnin=200,n.draw=200,maxiter=20,sdev=0.05,eps=1.0e-4)
```

**Arguments**

cpus	Number of CPU used for parallel computation.
dt1	A data matrix. The rows represent samples, and the columns represent genomic features.
dt2	A data matrix. The rows represent samples, and the columns represent genomic features.
dt3	A data matrix. The rows represent samples, and the columns represent genomic features.
dt4	A data matrix. The rows represent samples, and the columns represent genomic features.
type	data type, which can be "gaussian", "binomial", "poisson", and "multinomial".
K	The number of eigen features. Given K, the number of cluster is K+1.
alpha	Vector of elasticnet penalty terms. At this version of iClusterPlus, elasticnet is not used. Therefore, all the elements of alpha are set to 1.
n.lambda	Number of lambda are tuned.
scale.lambda	A value between (0,1); the actual lambda values will be scale.lambda multiplying the lambda values of the uniform design.
n.burnin	Number of MCMC burnin.
n.draw	Number of MCMC draw.
maxiter	Maximum iteration for the EM algorithm.
sdev	standard deviation of random walk proposal.
eps	EM algorithm convergence criterion.

**Value**

A list with the two elements 'fit' and 'lambda', where fit itself is a list and lambda is a matrix. Each row of lambda is the lambda values used to fit iClusterPlus model. Each component of fit is an object return by iClusterPlus, one-to-one corresponding to the row of lambda. Each component of fit has the following objects.

alpha	Intercept parameter for the genomic features.
beta	Information parameter for the genomic features. The rows and the columns represent the genomic features and the coefficients for the latent variable, respectively.
clusters	Cluster assignment.
centers	Cluster centers.
meanZ	Latent variable.

**Author(s)**

Qianxing Mo <qianxing.mo@moffitt.org>, Ronglai Shen <shenr@mskcc.org>

## References

Qianxing Mo, Sijian Wang, Venkatraman E. Seshan, Adam B. Olshen, Nikolaus Schultz, Chris Sander, R. Scott Powers, Marc Ladanyi, and Ronglai Shen. (2012). Pattern discovery and cancer gene identification in integrated cancer genomic data. *Proc. Natl. Acad. Sci. USA* 110(11):4245-50.

## See Also

[plotiCluster](#), [iClusterPlus](#), [iCluster2](#), [iCluster](#), [compute.pod](#)

## Examples

```
### see the users' guide iManu1.pdf
```

---

UM

*Uveal Melanoma data*

---

## Description

This is a subset of the uveal melanoma (UM) multi-omics data from the cancer genome atlas (TCGA) study (2018), which were re-analyzed by Mo et al. (2021).

## Usage

```
data(UM)
```

## Format

Data matrices of somatic mutation, DNA copy number, methylation and mRNA expression for 80 UM primary samples.

## Details

The TCGA UM multi-omics data (version 2016\_01\_28) were obtained from the Firebrowse portal (<http://firebrowse.org/>, accessed on 19 December 2018). The level 3 multi-omics data were processed for iCluster analysis, which were detailed in the Materials and Methods of Mo et al. (2021).

## Value

mut02	Somatic mutation data matrix with 0 representing wild type and 1 representing somatic mutation. Genes with mutation rate $\geq 2\%$ in the 80 samples are kept in the data matrix.
cn	Copy number regions, which were generated by merging the log2 ratios of chromosome segments using the CNregions function.
methy25	Methylation data matrix made of the top 25% most variable genes.
mrna25	mRNA expression data matrix made of the top 25% most variable genes.
methy25Anno	Annotation for the genes in the methylation data matrix methy25.
clin4	Clinical data of the 80 UM samples.

## References

Robertson, A.G.; Shih, J.; Yau, C.; Gibb, E.A.; Oba, J.; Mungall, K.L.; Hess, J.M.; Uzunangelov, V.; Walter, V.; Danilova, L.; et al. Integrative Analysis Identifies Four Molecular and Clinical Subsets in Uveal Melanoma. 2018. *Cancer Cell* 33 (1), 151.

Mo, Q; Wan, L; Schell, MJ; Jim, H; Tworoger, SS; G Peng, G. Integrative analysis identifies multi-omics signatures that drive molecular classification of uveal melanoma. 2021. *Cancers* 13 (24), 6168

---

utility

*Utility functions for iClusterPlus package*

---

## Description

Some utility functions for processing the results produced by iClusterPlus methods.

## Usage

```
getBIC(resultList)
getDevR(resultList)
getClusters(resultList)
iManual(view=TRUE)
```

## Arguments

resultList	A list object as shown in the following example.
view	A logical value TRUE or FALSE

## Value

getBIC	produce a matrix containing the BIC value for each lambda and K; the rows correspond to the lambda (vector) and the columns correspond to the K latent variables.
getDevR	produce a matrix containing the deviance ratio for each lambda and K; the rows correspond to the lambda (vector) and the columns correspond to the K latent variables.
getClusters	produce a matrix containing the cluster assignments for the samples under each K; the rows correspond to the samples; the columns correspond to the K latent variables.
iManual	Open the iClusterPlus User's Guide.

## Author(s)

Qianxing Mo <qianxing.mo@moffitt.org>

## References

Qianxing Mo, Sijian Wang, Venkatraman E. Seshan, Adam B. Olshen, Nikolaus Schultz, Chris Sander, R. Scott Powers, Marc Ladanyi, and Ronglai Shen. (2012). Pattern discovery and cancer gene identification in integrated cancer genomic data. *Proc. Natl. Acad. Sci. USA* (invited revision).

### See Also

[tune.iClusterPlus](#), [iClusterPlus](#), [iCluster2](#)

### Examples

```
### see the users' guide iManual.pdf

#data(simuResult)
#BIC = getBIC(simuResult)
#devR = getDevR(simuResult)
#clusters = getClusters(simuResult)
```

---

variation.hg18.v10.nov.2010

*Human genome variants of the NCBI 36 (hg18) assembly*

---

### Description

Human genome variants of the NCBI 36 (hg18) assembly

### Usage

```
data(variation.hg18.v10.nov.2010)
```

### Format

data frame

### Value

```
variation.hg18.v10.nov.2010
  Human genome variants of the NCBI 36 (hg18) assembly
```

### References

[http://projects.tcag.ca/variation/tableview.asp?table=DGV\\_Content\\_Summary.txt](http://projects.tcag.ca/variation/tableview.asp?table=DGV_Content_Summary.txt)

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