

Package ‘ggcyto’

May 26, 2026

Type Package

Title Visualize Cytometry data with ggplot

Version 1.41.0

Date 2015-11-02

Author Mike Jiang

Maintainer Mike Jiang <mike@ozette.com>

Description With the dedicated fortify method implemented for flowSet, ncdffFlowSet and GatingSet classes, both raw and gated flow cytometry data can be plotted directly with ggplot. ggcyto wrapper and some customized layers also make it easy to add gates and population statistics to the plot.

VignetteBuilder knitr

Depends methods, ggplot2(>= 3.5.0), flowCore(>= 1.41.5), ncdffFlow(>= 2.17.1), flowWorkspace(>= 4.3.1)

Imports plyr, scales, hexbin, data.table, RColorBrewer, gridExtra, rlang

Suggests testthat, flowWorkspaceData, knitr, rmarkdown, flowStats, openCyto, flowViz, ggridges, vdiffr

License file LICENSE

URL <https://github.com/RGLab/ggcyto/issues>

biocViews ImmunoOncology, FlowCytometry, CellBasedAssays, Infrastructure, Visualization

Collate 'AllClasses.R' 'autoplot.R' 'axis_inverse_trans.R'
'compute_stats.R' 'faust_gating_plot.R' 'fortify.R'
'fortify_fs.R' 'geom_gate.R' 'geom_hvline.R'
'geom_multi_range.R' 'geom_overlay.R' 'geom_stats.R'
'getFlowFrame.R' 'ggcyto.R' 'ggcyto_GatingLayout.R'
'ggcyto_GatingSet.R' 'ggcyto_flowSet.R' 'labs.R' 'ggcyto_par.R'
'ggplot_data_frame.R' 'merge_quad_gates.R' 'replace_data.R'
'scales_flowCore_fasinh.R' 'scales_flowJo_biexp.R'
'scales_flowJo_fasinh.R' 'scales_logicle.R' 'stat_position.R'
'transform_gate.R' 'utility.R'

RoxygenNote 7.2.1

Roxygen list(markdown=TRUE)

git_url <https://git.bioconductor.org/packages/ggcyto>

git_branch devel

git_last_commit 6c11f2e

git_last_commit_date 2026-04-28

Repository Bioconductor 3.24

Date/Publication 2026-05-25

Contents

as.ggplot	3
autoplot.flowSet	3
axis_x_inverse_trans	6
compute_stats	6
faust_gating_plot	7
flowCore_asinh_trans	8
fortify.cytoframe	9
fortify.ellipsoidGate	10
fortify.filterList	10
fortify.multiRangeGate	11
fortify.polygonGate	12
fortify.rectangleGate	12
fortify_fs	13
gate_null	14
geom_gate	14
geom_hvline	16
geom_multi_range	17
geom_overlay	18
geom_stats	19
getFlowFrame	20
ggcyto-class	21
ggcyto_add	23
ggcyto_arrange	24
ggcyto_par_default	24
ggcyto_par_set	25
is.ggcyto	26
is.ggcyto_flowSet	26
is.ggcyto_par	27
labs_cyto	27
marginalFilter	28
merge.quad.gates	29
print.ggcyto	30
print.ggcyto_GatingLayout	30
replace_data	31
scales_flowjo_biexp	32
scales_flowjo_fasinh	33
scale_x_flowCore_fasinh	33
scale_x_logicle	34
stats_null	35
stat_position	35
transform-gate	37

as.ggplot	<i>It fortifies the data, fills some default settings and returns a regular ggplot object.</i>
-----------	--

Description

The original data format is preserved during the ggcyto constructor because they still need to be used during the plot building process. This function is usually called automatically in the print/plot method of ggcyto. Sometime it is useful to coerce it to ggplot explicitly by user so that it can be used as a regular ggplot object.

Usage

```
as.ggplot(x, pre_binning = FALSE)
```

Arguments

x	ggcyto object with the data that has not yet been fortified to data.frame.
pre_binning	whether to pass the binned data to ggplot to avoid the overhead to scaling the original raw data for geom_hex layer

Value

ggplot object

Examples

```
data(GvHD)
fs <- GvHD[1:3]
#construct the `ggcyto` object (inherits from `ggplot` class)
p <- ggcyto(fs, aes(x = `FSC-H`)) + geom_histogram()
class(p) # a ggcyto object
p$data # data has not been fortified
p1 <- as.ggplot(p) # convert it to a ggplot object explicitly
class(p1)
p1$data # data is fortified
```

autoplot.flowSet	<i>Plot cytometry data in one or two dimension with the ggcyto package.</i>
------------------	---

Description

Overloaded autoplot methods for the cytometry data structure: flowFrame or flowSet, Gatinghierarchy, GatingSet. It plots the cytometry data with geom_histogram, geom_density or geom_hex. When autoplot is called on a GatingSet/Gatinghierarchy, the second argument should be a gate or population node. And the dimensions(channels/markers) are deduced from the gate dimensions.

Usage

```

## S3 method for class 'flowSet'
autoplot(object, x, y = NULL, bins = 30, ...)

## S3 method for class 'ncdfFlowList'
autoplot(object, ...)

## S3 method for class 'cytosep'
autoplot(object, ...)

## S3 method for class 'cytoframe'
autoplot(object, ...)

## S3 method for class 'flowFrame'
autoplot(object, x, ...)

## S3 method for class 'GatingSetList'
autoplot(object, ...)

## S3 method for class 'GatingSet'
autoplot(
  object,
  gate,
  x = NULL,
  y = "SSC-A",
  bins = 30,
  axis_inverse_trans = TRUE,
  ...
)

## S3 method for class 'GatingHierarchy'
autoplot(
  object,
  gate,
  y = "SSC-A",
  bool = FALSE,
  arrange.main = sampleNames(object),
  arrange = TRUE,
  merge = TRUE,
  projections = list(),
  strip.text = c("parent", "gate"),
  path = "auto",
  ...
)

```

Arguments

object	The data source. A core cytometry data structure. A flowFrame, flowSet, GatingSet or GatingHierarchy object
x	define the x dimension of the plot (not used when object is a GatingSet). When object is a flowFrame, it can be missing, which plots 1d density plot on all the channels.

y	define the y dimension of the plot. Default is NULL, which means 1d density-plot.
bins	passed to geom_hex
...	other arguments passed to ggplot
gate	the gate to be plotted
axis_inverse_trans	logical flag indicating whether to add axis_x_inverse_trans and axis_y_inverse_trans layers.
bool	whether to plot boolean gates
arrange.main	the main title of the arranged plots
arrange	whether to use arrangeGrob to put multiple plots in the same page
merge	whether to merge multiple gates into the same panel when they share the same parent and projections
projections	a list of customized projections
strip.text	either "parent" (the parent population name) or "gate" (the gate name). The latter usually is used when merge is FALSE
path	the gating path format (passed to gs_get_pop_paths)

Value

a ggcyto object

Examples

```
library(flowCore)
data(GvHD)
fs <- GvHD[subset(pData(GvHD), Patient %in%5:7 & Visit %in% c(5:6))["name"]]

#1d- density plot
autoplot(fs, x = "SSC-H")

#1d- density plot on all channels
autoplot(fs[[1]])

#2d plot: default geom_hex plot
autoplot(fs, x = 'FSC-H', y = 'SSC-H')

#autoplot for GatingSet
dataDir <- system.file("extdata", package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual", full = TRUE))
autoplot(gs, "CD3+")
#display axis values in transformed scale
autoplot(gs, "CD3+", axis_inverse_trans = FALSE)

#autoplot for GatingHierarchy
gh <- gs[[1]]
autoplot(gh) # by default the strip.text shows the parent population

#To display the gate name
#autoplot(gh, strip.text = "gate")
```

`axis_x_inverse_trans` *Display ggcyto axis labels using their raw values (as stored in the data structure)*

Description

It is essentially a dummy continuous scale and will be instantiated by `'+.ggcyto_GatingSet'` with `'breaks'` and `'lables'` customized.

Usage

```
axis_x_inverse_trans(...)
```

```
axis_y_inverse_trans(...)
```

Arguments

`...` common continuous scale parameters passed to `'continuous_scale'` (not used currently)

Value

a `raw_scale` object that inherits scale class.

Examples

```
dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
p <- ggcyto(gs, aes(x = CD4, y = CD8), subset = "CD3+") + geom_hex(bins = 64)
p <- p + geom_gate("CD4") + geom_stats() #plot CD4 gate and it is stats
p
p + axis_x_inverse_trans() #inverse transform the x axis into raw scale
```

`compute_stats` *compute the statistics of the cell population defined by gates*

Description

It calls the underlining stats routine and merge it with the label position calculated by `stat_position` as well as the `pData` of `flowSet`.

Usage

```
compute_stats(fs = NULL, gates, type = "percent", value = NULL, ...)
```

Arguments

fs	flowSet. can be NULL when precalculated 'value' is provided
gates	a list of filters
type	a vector of strings to specify the stats types. can be any or multiple values of "percent", "count", "gate_name", or "MFI" (MFI is currently not supported yet).
value	the pre-calculated stats value. when supplied, the stats computing is skipped.
...	other arguments passed to stat_position function

Details

This function is usually not called directly by user but used by ggcyto when geom_stat layer is added.

Value

a data.table that contains percent and centroid locations as well as pData that used as data for geom_btext layer.

Examples

```
data(GvHD)
fs <- GvHD[1:4]
rect.g <- rectangleGate(list("FSC-H" = c(300,500), "SSC-H" = c(50,200)), filterId = "P1")
rect.gates <- sapply(sampleNames(fs), function(sn)rect.g)
compute_stats(fs, rect.gates)
compute_stats(fs, rect.gates, type = c("gate_name", "percent"))
```

faust_gating_plot *plot faust gating schemes*

Description

plot faust gating schemes

Usage

```
faust_gating_plot(gh, start_node, end_node, ...)
```

Arguments

start_node	faust start node
end_node	the terminal leaf node generated by faust

Examples

```
## Not run:
gs=load_gs("~/Downloads/ics")

end_node = "/S/LV/L/CD4+/CD3+/CD8-/TNF+/CD107a-/IL4-/IFNg+/IL2+/CD154-/IL17a-"
start_node = "/S/LV/L"
gh=gs[[1]]
p = faust_gating_plot(gh, start_node, end_node, bins=128)
plot(ggcyto_arrange(p, nrow=1))

## End(Not run)
```

flowCore_asinht_trans *Inverse hyperbolic sine transformation(flowCore version).*

Description

Used to construct inverse hyperbolic sine transform object.

Usage

```
flowCore_asinht_trans(..., n = 6, equal.space = FALSE)
```

Arguments

...	parameters passed to arcsinhTransform
n	desired number of breaks (the actual number will be different depending on the data range)
equal.space	whether breaks at equal-spaced intervals

Value

asinht transformation object

Examples

```
trans.obj <- flowCore_asinht_trans(equal.space = TRUE)
data <- 1:1e3
brks.func <- trans.obj[["breaks"]]
brks <- brks.func(data)
brks # fasinh space displayed at raw data scale

#transform it to verify it is equal-spaced at transformed scale
trans.func <- trans.obj[["transform"]]
brks.trans <- trans.func(brks)
brks.trans
```

fortify.cytoframe	<i>Convert a flowFrame/flowSet/GatingSet to a ggplot-compatible data.table</i>
-------------------	--

Description

It extracts events matrices and appends the pData to it so that ggplot can use the pData for facetting.

Usage

```
## S3 method for class 'cytoframe'  
fortify(model, ...)  
  
## S3 method for class 'flowFrame'  
fortify(model, data, ...)  
  
## S3 method for class 'flowSet'  
fortify(model, data, ...)  
  
## S3 method for class 'cytoset'  
fortify(model, ...)  
  
## S3 method for class 'ncdfFlowList'  
fortify(model, ...)  
  
## S3 method for class 'GatingSetList'  
fortify(model, ...)  
  
## S3 method for class 'GatingSet'  
fortify(model, ...)
```

Arguments

model	flowFrame, flowSet or GatingSet
...	not used.
data	not used.

Value

data.table
data.table
data.table

Examples

```
dataDir <- system.file("extdata",package="flowWorkspaceData")  
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))  
  
attr(gs, "subset") <- "CD4" #must attach subset information to GatingSet object before fortifying it  
fortify(gs)
```

```
fs <- gs_pop_get_data(gs, "CD8")
fortify(fs)#fs is a flowSet/ncdfFlowSet

fr <- fs[[1]]
fortify(fr)#fr is a flowFrame
```

`fortify.ellipsoidGate` *Convert a ellipsoidGate to a data.table useful for ggplot*

Description

It interpolates the ellipsoidGate to polygongate before fortifying it.

Usage

```
## S3 method for class 'ellipsoidGate'
fortify(model, data = NULL, ...)
```

Arguments

<code>model</code>	ellipsoidGate
<code>data</code>	data range used for polygon interpolation.
<code>...</code>	not used.

Value

data.table

Examples

```
## Defining the gate
cov <- matrix(c(6879, 3612, 3612, 5215), ncol=2,
              dimnames=list(c("FSC-H", "SSC-H"), c("FSC-H", "SSC-H")))
mean <- c("FSC-H"=430, "SSC-H"=175)
eg <- ellipsoidGate(filterId= "myEllipsoidGate", .gate=cov, mean=mean)
fortify(eg)
```

`fortify.filterList` *Convert a filterList to a data.table useful for ggplot*

Description

It tries to merge with pData that is associated with filterList as attribute 'pd'

Usage

```
## S3 method for class 'filterList'
fortify(model, data = NULL, nPoints = NULL, ...)
```

Arguments

model	filterList
data	not used
nPoints	not used
...	not used.

Value

data.table

Examples

```
dataDir <- system.file("extdata", package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual", full = TRUE))
gates <- gs_pop_get_gate(gs, "CD4")
gates <- as(gates, "filterList") #must convert list to filterList in order for the method to dispatch properly
fortify(gates)
```

fortify.multiRangeGate

Convert a multiRangeGate to a data.table useful for ggplot

Description

It converts the boundaries slot into a data.table

Usage

```
## S3 method for class 'multiRangeGate'
fortify(model, data = NULL, ...)
```

Arguments

model	multiRangeGate
data	Not used
...	not used.
nPoints	not used

Value

data.table

Examples

```
mrq = multiRangeGate(ranges = list(min=c(100, 350), max=c(250, 400)))
fortify(mrq)
```

fortify.polygonGate *Convert a polygonGate to a data.table useful for ggplot*

Description

It converts the boundaries slot into a data.table

Usage

```
## S3 method for class 'polygonGate'
fortify(model, data = NULL, nPoints = NULL, ...)
```

Arguments

model	polygonGate
data	data range used to reset off-bound gate coordinates to prevent interpolating on the extremely large space unnecessarily.
nPoints	not used
...	not used.

Value

data.table

Examples

```
sqrct <- matrix(c(300,300,600,600,50,300,300,50),ncol=2,nrow=4)
colnames(sqrct) <- c("FSC-H","SSC-H")
pg <- polygonGate(filterId="nonDebris", .gate= sqrct)
fortify(pg)
```

fortify.rectangleGate *Convert a rectangleGate to a data.table useful for ggplot*

Description

For 2d rectangleGate, it is converted to a polygonGate first and then dispatch to the fortify method for polygonGate. for 1d, uses geom_vline/hline format.

Usage

```
## S3 method for class 'rectangleGate'
fortify(model, data = NULL, ...)
```

Arguments

model	rectangleGate
data	data range used for polygon interpolation.
...	not used.

Value

data.table

Examples

```
#2d rectangleGate
rect.g <- rectangleGate(list("FSC-H" = c(300,500), "SSC-H" = c(50,200)))
fortify(rect.g)
#1d gate
rg <- rectangleGate(list("FSC-H" = c(300,500)))
fortify(rg)
```

fortify_fs

Fortify a model into flowSet object

Description

The method provides a universe interface to convert a generic R object into a flowSet useful for ggcyto

Usage

```
fortify_fs(model, data, ...)

## S3 method for class 'flowSet'
fortify_fs(model, data, ...)

## Default S3 method:
fortify_fs(model, data, ...)

## S3 method for class 'flowFrame'
fortify_fs(model, data, ...)

## S3 method for class 'cytoframe'
fortify_fs(model, data, ...)

## S3 method for class 'GatingSetList'
fortify_fs(model, data, ...)

## S3 method for class 'GatingSet'
fortify_fs(model, data, ...)
```

Arguments

model	flow object(flowFrame or GatingSet) to be converted to flowSet. when it is a GatingSet, it must contain the subset information stored as 'subset' attribute.
data	original dataset, if needed
...	other arguments passed to methods

Value

a flowSet/ncdfFlowSet object

Examples

```
data(GvHD)
fr <- GvHD[[1]]
fortify_fs(fr)

dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
attr(gs, "subset") <- "CD4"
fortify_fs(gs)
```

gate_null	<i>clear all the geom_gate() layer previously added</i>
-----------	---

Description

clear all the geom_gate() layer previously added

Usage

```
gate_null()
```

Examples

```
dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
#autoplot display pop stats by default
p <- autoplot(gs, "CD4")
#it is easy to remove the default gate
p <- p + gate_null()
#and add a new one
p <- p + geom_gate("CD8")
p
```

geom_gate	<i>Add a gate layer to a ggcyto plot.</i>
-----------	---

Description

When 'data' is a gate (or flowCore filter) or a list of gates or a filterList object. When it is used directly with 'ggplot', pdata of the flow data must be supplied through 'pd' argument explicitly in order for the gates to be dispatched to each panel. However It is not necessary when used with 'ggcyto' wrapper since the latter will attach pData automatically.

Usage

```
geom_gate(data, ...)

## S3 method for class 'filterList'
geom_gate(data, pd, nPoints = 100, ...)

## S3 method for class 'filter'
geom_gate(data, mapping = NULL, fill = NA, colour = "red", nPoints = 100, ...)
```

Arguments

data	a filter (Currently only rectangleGate (1d or 2d), polygonGate, ellipsoidGate are supported.) or a list of these gates or filterList or character specifying a gated cell population in the GatingSet
...	other arguments
pd	pData (data.frame) that has rownames represents the sample names used as key to be merged with filterList
nPoints	used for interpolating polygonGates to prevent them from losing shape when truncated by axis limits
mapping	The aesthetic mapping
fill	fill color for the gate. Not filled by default.
colour	default is red

Details

When 'data' is a character, it construct an abstract geom layer for a character that represents nodes in a Gating tree and will be instantiated later as a specific geom_gate layer or layers based on the gates extracted from the given GatingSet object.

Value

a geom_gate layer

Examples

```
data(GvHD)
fs <- GvHD[subset(pData(GvHD), Patient %in%5:7 & Visit %in% c(5:6))][["name"]]
p <- ggcyto(fs, aes(x = `FSC-H`, y = `SSC-H`))
p <- p + geom_hex(bins = 128)
rect.g <- rectangleGate(list("FSC-H" = c(300,500), "SSC-H" = c(50,200)))
#constructor for a list of filters
rect.gates <- sapply(sampleNames(fs), function(sn)rect.g)
p + geom_gate(rect.gates)

dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
p <- ggcyto(gs, aes(x = CD4, y = CD8), subset = "CD3+") + geom_hex(bins = 64)
# add gate layer by gate name
p + geom_gate("CD4")
```

geom_hvline *Vertical or horizontal line.*

Description

This geom is based on the source code of ' [geom_hline](#) and [geom_vline](#).

Usage

```
geom_hvline(
  mapping = NULL,
  data = NULL,
  position = "identity",
  show.legend = FALSE,
  ...
)
```

Arguments

mapping	The aesthetic mapping, usually constructed with aes or aes_string . Only needs to be set at the layer level if you are overriding the plot defaults.
data	A layer specific dataset - only needed if you want to override the plot defaults.
position	The position adjustment to use for overlapping points on this layer
show.legend	should a legend be drawn? (defaults to FALSE)
...	other arguments passed on to layer . This can include aesthetics whose values you want to set, not map. See layer for more details.

Details

The goal is to determine the line to be either vertical or horizontal based on the 1-d data provided in this layer.

Value

a geom_hvline layer

Aesthetics

@section Aesthetics: [geom_vline\(\)](#) understands the following aesthetics. Required aesthetics are displayed in bold and defaults are displayed for optional aesthetics:

- xintercept
- **alpha** → NA
- **colour** → via theme()
- **group** → inferred
- **linetype** → via theme()
- **linewidth** → via theme()

Learn more about setting these aesthetics in `vignette("ggplot2-specs")`.

Examples

```
p <- ggplot(mtcars, aes(x = wt, y = mpg)) + geom_point()
# vline
p + geom_hvline(data = data.frame(wt= 3))
# hline
p + geom_hvline(data = data.frame(mpg= 20))
```

geom_multi_range *Draw multi-ranges as multiple rectangles on 1D or 2D plot*

Description

This geom is based on the source code of [geom_rect](#)

Usage

```
geom_multi_range(
  mapping = NULL,
  data = NULL,
  stat = "identity",
  position = "identity",
  ...,
  linejoin = "mitre",
  na.rm = FALSE,
  show.legend = NA,
  inherit.aes = TRUE
)
```

Arguments

mapping	The aesthetic mapping, usually constructed with aes or aes_string . Only needs to be set at the layer level if you are overriding the plot defaults.
data	A layer specific dataset - only needed if you want to override the plot defaults.
position	The position adjustment to use for overlapping points on this layer
...	other arguments passed on to layer . This can include aesthetics whose values you want to set, not map. See layer for more details.
show.legend	should a legend be drawn? (defaults to FALSE)

Details

The goal is to determine the line to be either vertical or horizontal based on the data provided in this layer. Also convert input 1D intervals to geom_rect acceptable shapes

Value

a geom_rect layer

Aesthetics

@section Aesthetics: `geom_vline()` understands the following aesthetics. Required aesthetics are displayed in bold and defaults are displayed for optional aesthetics:

- `xintercept`
- `alpha` → NA
- `colour` → via `theme()`
- `group` → inferred
- `linetype` → via `theme()`
- `linewidth` → via `theme()`

Learn more about setting these aesthetics in `vignette("ggplot2-specs")`.

<code>geom_overlay</code>	<i>Overlay a population on an existing ggcyto plot analogous to backgating.</i>
---------------------------	---

Description

It is useful for "backgating" plots.

Usage

```
geom_overlay(data, ...)
```

Arguments

<code>data</code>	a filter (Currently only <code>rectangleGate</code> (1d or 2d), <code>polygonGate</code> , <code>ellipsoidGate</code> are supported.) or a list of these gates or <code>filterList</code> or character specifying a gated cell population in the <code>GatingSet</code>
<code>...</code>	other arguments mapping, The mapping aesthetic mapping data a <code>polygonGate</code> fill <code>polygonGate</code> is not filled by default <code>colour</code> default is red <code>pd</code> <code>pData</code> (<code>data.frame</code>) that has rownames represents the sample names used as key to be merged with <code>filterList</code>

Value

a `geom_overlay` layer

Examples

```
library(ggcyto)
dataDir <- system.file("extdata", package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual", full = TRUE))
p <- autoplot(gs, "CD3+")

# add a flowSet as the overlay
fs <- gs_pop_get_data(gs, "DPT")
p + geom_overlay(data = fs, size = 0.3, alpha = 0.7)
```

```
# add overlay layer by gate name
p + geom_overlay(data = "DNT", size = 0.3, alpha = 0.7)

#add overlay for 1d densityplot
p <- ggcyto(gs, aes(x = CD4), subset = "CD3+") + geom_density(aes(y = ..count..))
p + geom_overlay("DNT", aes(y = ..count..), fill = "red")
```

geom_stats

Add a population statistics layer to a ggcyto plot.

Description

This is a virtual layer and will be instantiated as `geom_label` layer within `ggcyto.+` operator.

Usage

```
geom_stats(
  gate = NULL,
  ...,
  value = NULL,
  type = "percent",
  negated = FALSE,
  adjust = 0.5,
  location = "gate",
  label.padding = unit(0.05, "lines"),
  label.size = 0,
  digits = 3
)
```

Arguments

gate	a 'filterList' or character (represent as a population node in GatingSet) if not supplied, ggcyto then tries to parse the gate from the first <code>geom_gate</code> layer.
...	other arguments passed to <code>geom_label</code> layer
value	the pre-calculated stats value. when supplied, the stats computing is skipped.
type	a vector of strings to specify the stats types. can be any or multiple values of "percent", "count", "gate_name", or "MFI" (MFI is currently not supported yet).
negated	whether the gate needs to be negated
adjust	see details for stat_position
location	see details for stat_position
label.padding, label.size	arguments passed to <code>geom_label</code> layer
digits	control the stats format

Details

So it is dedicated for `ggcyto` context and thus cannot be added to `ggplot` object directly.

Value

a `geom_popStats` layer

Examples

```

dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
p <- ggcyto(gs, aes(x = CD4, y = CD8), subset = "CD3+") + geom_hex(bins = 64)
p
# add gate and stats layer
p + geom_gate("CD4") + geom_stats()

# display gate name
p + geom_gate(c("CD4", "CD8")) + geom_stats(type = "gate_name")
# display gate name and percent
p + geom_gate(c("CD4", "CD8")) + geom_stats(type = c("gate_name", "percent"))

```

getFlowFrame

extract flowFrame data structure from the given R object

Description

Mainly to get the channel and marker information.

Usage

```
getFlowFrame(x)
```

Arguments

x flowSet, ncdfFlowList, GatingSet, GatingHierarchy, or GatingSetList

Value

a flowFrame. When x is a ncdfFlowSet or GatingSet that is associated with ncdfFlowSet, the raw event data is not read and an empty flowFrame is returned.

Examples

```

data(GvHD)
fs <- GvHD[1:2]
getFlowFrame(fs)# fs is a flowSet

dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
getFlowFrame(gs)# gs is a GatingSet

```

ggcyto-class

*Plot cytometry data using the ggcyto API***Description**

ggcyto() initializes a ggcyto object that inherits ggplot class. Similarly the + operator can be used to add layers to the existing ggcyto object.

Usage

```
ggcyto(data = NULL, ...)

## S3 method for class 'GatingSet'
ggcyto(data, mapping, subset = "_parent_", ...)

## S3 method for class 'GatingSetList'
ggcyto(data, ...)

## S3 method for class 'GatingHierarchy'
ggcyto(data, ...)

## S3 method for class 'flowSet'
ggcyto(data, mapping, filter = NULL, max_nrow_to_plot = 50000, ...)
```

Arguments

data	The data source. A core cytometry data structure. (flowSet, flowFrame, ncdf-FlowSet, GatingSet or GatingHierarchy)
...	other arguments passed to specific methods
mapping	default list of aesthetic mappings (these can be colour, size, shape, line type – see individual geom functions for more details)
subset	character that specifies the node path or node name in the case of GatingSet. Default is "parent", which will be substituted with the actual node name based on the geom_gate layer to be added later.
filter	a flowcore gate object or a function that takes a flowSet and channels as input and returns a data-dependent flowcore gate. The gate is used to filter the flow data before it is plotted.
max_nrow_to_plot	the maximum number of cells to be plotted. When the actual data exceeds it, The subsampling process will be triggered to speed up plotting. Default is 5e4. To turn off the subsampling, simply set it to a large enough number or Inf.

Details

To invoke ggcyto:

- ggcyto(fs, aes(x, y, <other aesthetics>))

Value

ggcyto object

Examples

```

data(GvHD)
fs <- GvHD[1:3]
#construct the `ggcyto` object (inherits from `ggplot` class)
p <- ggcyto(fs, aes(x = `FSC-H`))
p + geom_histogram()

# display density/area
p + geom_density()
p + geom_area(stat = "density")

# 2d scatter plot
p <- ggcyto(fs, aes(x = `FSC-H`, y = `SSC-H`))
p + geom_hex(bins = 128)
# do it programatically through aes_string and variables
col1 <- "`FSC-H`" #note that the dimension names with special characters needs to be quoted by backticks
col2 <- "`SSC-H`"
ggcyto(fs, aes_string(col1,col2)) + geom_hex()

## More flowSet examples
fs <- GvHD[subset(pData(GvHD), Patient %in%5:7 & Visit %in% c(5:6))][["name"]]
# 1d histogram/densityplot
p <- ggcyto(fs, aes(x = `FSC-H`))
#facet_wrap(~name)` is used automatically
p1 <- p + geom_histogram()
p1
#overwriting the default faceting
p1 + facet_grid(Patient~Visit)

#display density
p + geom_density()

#you can use ggridges package to display stacked density plot
require(ggridges)
#stack by fcs file ('name')
p + geom_density_ridges(aes(y = name)) + facet_null() #facet_null is used to remove the default facet_wrap (by 'name')
#or to stack by Visit and facet by patient
p + geom_density_ridges(aes(y = Visit)) + facet_grid(~Patient)

# 2d scatter/dot plot
p <- ggcyto(fs, aes(x = `FSC-H`, y = `SSC-H`))
p <- p + geom_hex(bins = 128)
p

## GatingSet
dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
# 2d plot
ggcyto(gs, aes(x = CD4, y = CD8), subset = "CD3+") + geom_hex(bins = 64)

# 1d plot
ggcyto(gs, aes(x = CD4), subset = "CD3+") + geom_density()

```

ggcyto_add

*overloaded '+' method for ggcyto***Description**

It tries to copy pData from ggcyto object to the gate layers so that the gate layer does not need to have pd to be supplied explicitly by users. It also calculates population statistics when geom_stats layer is added. It supports addition ggcyto layers such as 'ggcyto_par' and 'labs_cyto'.

Usage

e1 + e2

Arguments

e1 An object of class ggcyto or a class inheriting from ggcyto, such as ggcyto_flowSet, ggcyto_GatingSet, or ggcyto_GatingLayout. In the case of ggcyto_GatingLayout, the component of e2 will be added to each subsidiary plot.

e2 A component to add to e1

Value

ggcyto object

Examples

```
## flowSet
data(GvHD)
fs <- GvHD[subset(pData(GvHD), Patient %in% 5:7 & Visit %in% c(5:6))][["name"]]
p <- ggcyto(fs, aes(x = `FSC-H`, y = `SSC-H`)) + geom_hex(bins = 128)
#add rectangleGate layer (2d)
rect.g <- rectangleGate(list("FSC-H" = c(300,500), "SSC-H" = c(50,200)))
rect.gates <- sapply(sampleNames(fs), function(sn)rect.g)
p + geom_gate(rect.gates) + geom_stats()

## GatingSet
dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
p <- ggcyto(gs, aes(x = CD4, y = CD8), subset = "CD3+") + geom_hex(bins = 64)
p <- p + geom_gate("CD4") + geom_stats() #plot CD4 gate and it is stats
p
p + axis_x_inverse_trans() #inverse transform the x axis into raw scale

## GatingLayout
#autoplot for GatingSet
dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
gh <- gs[[1]]
p <- autoplot(gh)
class(p)
# customize the font size of strip text for each ggcyto plots contained in GatingLayout object
p + theme(strip.text = element_text(size = 14))
```

ggcyto_arrange *Arrange a list of ggplot objects into gtable*

Description

It is usually implicitly invoked by print and show method and can be called by user when the further manipulation is needed,

Usage

```
ggcyto_arrange(x, ...)
```

Arguments

x ggcyto_gate_layout, which is essentially a list of ggplot objects that were previously stored as ggcyto_gate_layout object by autoplot function.

... other arguments passed to arrangeGrob

Value

gtable

Examples

```
## Not run:
# get ggcyto_GatingLayout object from first sample
res <- autoplot(gs[[1]], nodes, bins = 64)
class(res)
# arrange it as one-row gtable object
gt <- ggcyto_arrange(res, nrow = 1)
gt
# do the same to the second sample
gt2 <- ggcyto_arrange(autoplot(gs[[2]], nodes, bins = 64), nrow = 1)
# combine the two and print it on the same page
gt3 <- gridExtra::gtable_rbind(gt, gt2)
plot(gt3)

## End(Not run)
```

ggcyto_par_default *Return The default ggcyto settings*

Description

Return The default ggcyto settings

Usage

```
ggcyto_par_default()
```

Value

a list of default settings for ggcyto

Examples

```
ggcyto_par_default()
```

```
ggcyto_par_set          Set some default parameters for ggcyto
```

Description

Use this function to modify ggcyto parameters These are the regular (or to be instantiated as) scales, labs, facet objects. They can be added as a single layer to the plot for the convenience.

Usage

```
ggcyto_par_set(...)
```

Arguments

... a list of element name, element pairings that modify the existing parameter settings

Value

a list of new settings for ggcyto

elements

The individual elements are:

limits	can be "data"(default) or "instrument" or a list of numeric limits for x and y (e.g. <code>list(x = c(0, 4000))</code>)
facet	the regular facet object
hex_fill	default scale_fill_gradientn for geom_hex layer
lab	labs_cyto object

Examples

```
library(ggcyto)
dataDir <- system.file("extdata", package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual", full = TRUE))

p <- ggcyto(gs, aes(x = CD4, y = CD8), subset = "CD3+")
# 2d plot
p <- p + geom_hex(bins = 64)
p

#use instrument range by overwriting the default limits settings
p + ggcyto_par_set(limits = "instrument")

#manually set limits
myPars <- ggcyto_par_set(limits = list(x = c(0,3.2e3), y = c(-10, 3.5e3)))
p + myPars# or xlim(0,3.2e3) + ylim(-10, 3.5e3)
```

is.ggcyto	<i>Reports whether x is a ggcyto object</i>
-----------	---

Description

Reports whether x is a ggcyto object

Usage

```
is.ggcyto(x)
```

Arguments

x An object to test

Value

TRUE/FALSE

Examples

```
data(GvHD)
fs <- GvHD[1:2]
p <- ggcyto(fs, aes(x = `FSC-H`))
is.ggcyto(p)
```

is.ggcyto_flowSet	<i>Reports whether x is a ggcyto_flowSet object</i>
-------------------	---

Description

Reports whether x is a ggcyto_flowSet object

Usage

```
is.ggcyto_flowSet(x)
```

Arguments

x An object to test

Value

TRUE or FALSE

Examples

```
data(GvHD)
fs <- GvHD[1:2]
p <- ggcyto(fs, aes(x = `FSC-H`))
is.ggcyto_flowSet(p)
```

is.ggcyto_par	<i>Reports whether x is a ggcyto_par object</i>
---------------	---

Description

Reports whether x is a ggcyto_par object

Usage

```
is.ggcyto_par(x)
```

Arguments

x	An object to test
---	-------------------

Value

TRUE or FALSE

Examples

```
myPar <- ggcyto_par_set(limits = "instrument")
is.ggcyto_par(myPar)
```

labs_cyto	<i>Change axis labels and legend titles</i>
-----------	---

Description

The actual labels text will be instantiated when it is added to ggcyto plot.

Usage

```
labs_cyto(labels = "both")
```

Arguments

labels	default labels for x, y axis. Can be "channel" , "marker", or "both" (default)
--------	--

Value

a list

Examples

```

dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))

# default is "both"
p <- ggcyto(gs, aes(x = CD4, y = CD8), subset = "CD3+") + geom_hex(bins = 64)
p

#use marker name as x,y labs
p + labs_cyto("marker")

#use channel name as x,y labs
p + labs_cyto("channel")

```

marginalFilter	<i>Generate a marginal gate.</i>
----------------	----------------------------------

Description

It simply constructs an `boundaryFilter` that removes the marginal events. It can be passed directly to `ggcyto` constructor. See the examples for details.

Usage

```
marginalFilter(fs, dims, ...)
```

Arguments

<code>fs</code>	flowSet (not used.)
<code>dims</code>	the channels involved
<code>...</code>	arguments passed to boundaryFilter

Value

an `boundaryFilter`

Examples

```

data(GvHD)
fs <- GvHD[1]
chnls <- c("FSC-H", "SSC-H")
#before removign marginal events
summary(fs[, chnls])

# create marginal filter
g <- marginalFilter(fs, chnls)
g

#after remove marginal events
fs.clean <- Subset(fs, g)
summary(fs.clean[, chnls])

#pass the function directly to ggcyto

```

```

dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
# with marginal events
ggcyto(gs, aes(x = CD4, y = CD8), subset = "CD3+") + geom_hex(bins = 64)

# using marginalFilter to remove these events
ggcyto(gs, aes(x = CD4, y = CD8), subset = "CD3+", filter = marginalFilter) + geom_hex(bins = 64)

```

merge.quad.gates	<i>extend the original flowWorkspace:::mergeGates function to restore quadGate when applicable</i>
------------------	--

Description

For internal usage.

Usage

```

## S3 method for class 'quad.gates'
merge(gh, pops, bool = TRUE)

```

Arguments

gh	a GatingHierarchy
pops	a vector of population names
bool	whether to deal with boolean gate

Value

a nested list of data structure that captures the information of parent, grouped populations (with the same projections) and the reconstructed quadGate object and the respective quadrant pattern

Examples

```

library(flowWorkspace)
dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(file.path(dataDir, "gs_manual"))
#get the GatingHierarchy object
gh <- gs[[1]]
pops <- gs_pop_get_children(gh, "CD4")
grps <- ggcyto:::merge.quad.gates(gh, pops)
length(grps) # pops are grouped into two
grps[[1]] # each group is annotaed with quadGate information

ggcyto:::merge.quad.gates(gh, gs_pop_get_children(gh, "CD3+")) # cd3 subsets are not coercible to quadgate th

```

print.ggcyto	<i>Draw ggcyto on current graphics device.</i>
--------------	--

Description

A wrapper for print.ggplot. It converts the ggcyto to conventional ggplot object before printing it. This is usually invoked automatically when a ggcyto object is returned to R console.

Usage

```
## S3 method for class 'ggcyto'
print(x, ...)

## S3 method for class 'ggcyto'
plot(x, ...)

## S3 method for class 'ggcyto'
show(object)
```

Arguments

x	ggcyto object to display
...	other arguments not used by this method
object	ggcyto object

Value

nothing

print.ggcyto_GatingLayout	<i>print method for ggcyto_gate_layout class</i>
---------------------------	--

Description

print method for ggcyto_gate_layout class

Usage

```
## S3 method for class 'ggcyto_GatingLayout'
print(x, ...)

## S3 method for class 'ggcyto_GatingLayout'
show(object)
```

Arguments

x	ggcyto_gate_layout, which is essentially a list of ggplot objects that were previously stored as ggcyto_gate_layout object by autoplot function.
...	other arguments passed to arrangeGrob
object	ggcyto_GatingLayout

Value

nothing

replace_data	<i>replace current cytometry data</i>
--------------	---------------------------------------

Description

It essentially reconstructs the entire ggcyto plot object based on the new data and the original mapping and layers recorded in the plot object.

Usage

```
e1 %>% e2
```

Arguments

e1	the ggcyto object
e2	the new cytometry data . It can be 'GatingSet' or 'flowSet'.

Value

the new ggcyto object

Examples

```
dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_bcell_auto",full = TRUE))
gs1 <- gs[1]
gs2 <- gs[2]

#construct the ggcyto object for gs1
p <- ggcyto(gs1, aes(cd24, cd38)) + geom_hex(bins = 128)
p <- p + geom_gate("Transitional") #add gate
#customize the stats layer
p <- p + geom_stats(type = "count", size = 6, color = "white", fill = "black", adjust = 0.3)
#customize the layer
p <- p + labs_cyto("channel")
#customize the axis limits
p <- p + ggcyto_par_set(limits = "instrument")
#add another population as the overlay dots
p <- p + geom_overlay("IgD-CD27-", col = "black", size = 1.2, alpha = 0.4)
p

#replace the data with gs2 and see the same visual effect
p %>% gs2
```

scales_flowjo_biexp *Add a flowJo biexponential scale to the x or y axes of a ggcyto plot.*

Description

Add a flowJo biexponential scale to the x or y axes of a ggcyto plot.

Usage

```
scale_x_flowjo_biexp(
  ...,
  maxValue = 262144,
  widthBasis = -10,
  pos = 4.5,
  neg = 0,
  equal.space = FALSE
)

scale_y_flowjo_biexp(
  ...,
  maxValue = 262144,
  widthBasis = -10,
  pos = 4.5,
  neg = 0,
  equal.space = FALSE
)
```

Arguments

... common continuous scale parameters passed to 'continuous_scale' (not used currently)

maxValue, widthBasis, pos, neg
 see 'help(flowjo_biexp)'

equal.space whether to display the breaks in equal.space format

Value

ScaleContinuous object

Examples

```
data(GvHD)
fr <- GvHD[[1]]
p <- ggcyto(fr, aes(x = `FL1-H`)) + geom_density()
#display at raw scale
p
#display at transformed scale
p + scale_x_flowjo_biexp(maxValue = 1e4, widthBasis = 0)
```

`scales_flowjo_fasinh` *Add a flowJo inverse hyperbolic sine scale to the x or y axes of a ggcyto plot.*

Description

Add a flowJo inverse hyperbolic sine scale to the x or y axes of a ggcyto plot.

Usage

```
scale_x_flowjo_fasinh(..., m = 4, t = 1200)
```

```
scale_y_flowjo_fasinh(..., m = 4, t = 1200)
```

Arguments

`...` common continuous scale parameters passed to 'continuous_scale' (not used currently)

`m, t` see 'help(flowjo_fasinh)'

Value

ScaleContinuous object

Examples

```
data(GvHD)
fr <- GvHD[[1]]
p <- ggcyto(fr, aes(x = `FL1-H`)) + geom_density()
#display at raw scale
p
#display at transformed scale
p + scale_x_flowjo_fasinh(t = 1e4)
```

`scale_x_flowCore_fasinh`

Add a flowCore inverse hyperbolic sine scale to the x or y axes of a ggcyto plot.

Description

Add a flowCore inverse hyperbolic sine scale to the x or y axes of a ggcyto plot.

Usage

```
scale_x_flowCore_fasinh(..., a = 1, b = 1, c = 0)
```

```
scale_y_flowCore_fasinh(..., a = 1, b = 1, c = 0)
```

Arguments

... common continuous scale parameters passed to 'continuous_scale' (not used currently)

a, b, c see 'help(arcsinhTransform)'

Value

ScaleContinuous object

Examples

```
data(GvHD)
fr <- GvHD[[1]]
p <- ggcyto(fr, aes(x = `FL1-H`)) + geom_density()
#display at raw scale
p
#display at transformed scale
p + scale_x_flowCore_fasinh(a = 2)
```

scale_x_logicle *Add a logicle scale to the x or y axes of a ggcyto plot.*

Description

Add a logicle scale to the x or y axes of a ggcyto plot.

Usage

```
scale_x_logicle(..., w = 0.5, t = 262144, m = 4.5, a = 0)
```

```
scale_y_logicle(..., w = 0.5, t = 262144, m = 4.5, a = 0)
```

Arguments

... common continuous scale parameters passed to 'continuous_scale' (not used currently)

w, t, m, a see 'help(logicleTransform)'

Value

ScaleContinuous object

Examples

```
data(GvHD)
fr <- GvHD[[1]]
p <- ggcyto(fr, aes(x = `FL1-H`)) + geom_density()
#display at raw scale
p
#display at transformed scale
p + scale_x_logicle(t = 1e4)
```

stats_null	<i>clear all the geom_stats() layer previously added</i>
------------	--

Description

clear all the geom_stats() layer previously added

Usage

```
stats_null()
```

Examples

```
dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
#autoplot display pop stats by default
p <- autoplot(gs, "CD4")
#it is easy to remove the default stats
p <- p + stats_null()
#and add a new one
p <- p + geom_stats(type = "count")
```

stat_position	<i>Compute the positions of the population statistics based on the geometric gate centroid for a ggcyto plot.</i>
---------------	---

Description

It is usually not called directly by user but mainly used by compute_stats function (which is called by ggcyto add method when geom_states layer is added).

Usage

```
stat_position(gate, ...)

## S3 method for class 'filter'
stat_position(
  gate,
  negated = FALSE,
  adjust = 0.5,
  location = "gate",
  data_range = NULL,
  limits = NULL,
  ...
)
```

Arguments

gate	a flowCore filter
...	other arguments
negated	logical indicating whether position needs to be moved to negative side of gate
adjust	see details
location	see details
data_range	a two-row data.frame representing the actual data range. Each column is a range for a specific channel. First row is min, Second row is max.
limits	used to fix the gate range

Details**Specifying location for statistical annotation:**

The `adjust` and `location` arguments allow for a few different ways to adjust the location of the statistical annotation for a gate on a ggcyto plot. The valid values for `location` are "gate" (default), "data", "plot", and "fixed".

Relative location:

If `location` is not "fixed", the starting position of the annotation will be determined with respect to a rectangular window whose bounds are determined in the following way:

- For `location = "gate"`, the window will be set by the range of the data in the gate
- For `location = "data"`, the window will be set by the range of values in all of the data on the plot (provided by `data_range`)
- For `location = "plot"`, the window will be set by the axis limits of the plot (adjusted by `ggcyto_par_set`)

This starting position can then be adjusted by passing values in a vector to the `adjust` parameter, where they will be interpreted as relative proportions of the window dimension, where 0.0 represents the lower bound of the dimension and 1.0 represents the upper bound. So, for a 2-D plot, `adjust=c(0,0)` places the annotation at the lower left corner of this window and `adjust=c(1,1)` places it at the upper right corner.

As another example, for a 2-D gate, if `location = "gate"` and `adjust=c(0.25, 0.75)`, the statistical annotation will be placed 1/4 of the way across the x-range of the gate and 3/4 of the way across the y-range of the gate.

The `adjust` argument will also accept values less than 0.0 or greater than 1.0. This can be an easy way to simply move the annotation outside of a gate so it does not obstruct the view of the data within. For example, `location = "gate"` and `adjust=c(-0.2, 1.2)` will move the annotation outside of the upper left corner of the gate range.

Fixed location:

If `location = "fixed"`, the numeric vector passed to `adjust` will be interpreted as values on the data scales of the plot to provide an explicit location for the annotation.

For example, if the annotation should be at the location 3000, 5000 on the plot, that could be done with `location="fixed"` and `adjust = c(3000,5000)`.

Default:

The default behavior if no values are provided to `location` or `adjust` will be to place the annotation at the center of the range of the data in the gate.

Value

a data.table of gate centroid coordinates

Examples

```
data(GvHD)
fs <- GvHD[1:4]
rect.g <- rectangleGate(list("FSC-H" = c(300,500), "SSC-H" = c(50,200)))
rect.gates <- sapply(sampleNames(fs), function(sn)rect.g)
stat_position(rect.gates)
```

transform-gate	<i>rescale methods for gates</i>
----------------	----------------------------------

Description

rescale the gate coordinates with the transformation provided

Usage

```
transform(`_data`, ...)  
rescale_gate(gate, trans, param)
```

Arguments

<code>_data</code>	the filter or filterList object. Currently support polygonGate, ellipsoidGate, rectangleGate and quadGate.
<code>...</code>	trans the transformation function or transformList object param the parameter/dimension to be transformed. When trans is transformList object, param is not needed since it is derived from transformList.
<code>gate</code>	gate object
<code>trans</code>	the transformation function
<code>param</code>	the parameter/dimension to be transformed.

Value

the transformed filter/filterList object

Index

- + (ggcyto_add), 23
- + , ggcyto_GatingLayout , ANY-method (ggcyto_add), 23
- + , ggcyto_GatingLayout-method (ggcyto_add), 23
- + , ggcyto_GatingSet , ANY-method (ggcyto_add), 23
- + , ggcyto_GatingSet-method (ggcyto_add), 23
- + , ggcyto_flowSet , ANY-method (ggcyto_add), 23
- + , ggcyto_flowSet-method (ggcyto_add), 23
- + . ggcyto_GatingLayout (ggcyto_add), 23
- + . ggcyto_GatingSet (ggcyto_add), 23
- + . ggcyto_flowSet (ggcyto_add), 23
- + . ggcyto_ncdfFlowList (ggcyto_add), 23
- %% (replace_data), 31
- %% , ggcyto-method (replace_data), 31
- %% , ggcyto_GatingLayout , ANY-method (replace_data), 31
- %% , ggcyto_GatingLayout-method (replace_data), 31

- aes, 16, 17
- aes_string, 16, 17
- alpha, 16, 18
- as.ggplot, 3
- autoplot (autoplot.flowSet), 3
- autoplot.flowSet, 3
- axis_x_inverse_trans, 5, 6
- axis_y_inverse_trans (axis_x_inverse_trans), 6

- boundaryFilter, 28

- colour, 16, 18
- compute_stats, 6

- faust_gating_plot, 7
- flowCore_asinht_trans, 8
- fortify (fortify.cytoframe), 9
- fortify.cytoframe, 9
- fortify.ellipsoidGate, 10
- fortify.filterList, 10

- fortify.multiRangeGate, 11
- fortify.polygonGate, 12
- fortify.rectangleGate, 12
- fortify_fs, 13

- gate_null, 14
- geom_gate, 14
- geom_hline, 16
- geom_hvline, 16
- geom_multi_range, 17
- geom_overlay, 18
- geom_rect, 17
- geom_stats, 19
- geom_vline, 16
- getFlowFrame, 20
- ggcyto (ggcyto-class), 21
- ggcyto-class, 21
- ggcyto.default (ggcyto-class), 21
- ggcyto.flowSet (ggcyto-class), 21
- ggcyto.GatingHierarchy (ggcyto-class), 21
- ggcyto.GatingSet (ggcyto-class), 21
- ggcyto.GatingSetList (ggcyto-class), 21
- ggcyto_add, 23
- ggcyto_arrange, 24
- ggcyto_flowSet-class (ggcyto-class), 21
- ggcyto_GatingLayout-class (ggcyto-class), 21
- ggcyto_GatingSet-class (ggcyto-class), 21
- ggcyto_par_default, 24
- ggcyto_par_set, 25, 36
- group, 16, 18
- gs_get_pop_paths, 5

- is.ggcyto, 26
- is.ggcyto_flowSet, 26
- is.ggcyto_par, 27

- labs_cyto, 27
- layer, 16, 17
- linetype, 16, 18
- linewidth, 16, 18

- marginalFilter, 28

`merge.quad.gates`, 29

`plot.ggcyto` (`print.ggcyto`), 30

`print,ggcyto-method` (`print.ggcyto`), 30

`print.ggcyto`, 30

`print.ggcyto_GatingLayout`, 30

`replace_data`, 31

`rescale_gate` (`transform-gate`), 37

`scale_x_flowCore_fasinh`, 33

`scale_x_flowJo_biexp`
 (`scales_flowjo_biexp`), 32

`scale_x_flowjo_biexp`
 (`scales_flowjo_biexp`), 32

`scale_x_flowJo_fasinh`
 (`scales_flowjo_fasinh`), 33

`scale_x_flowjo_fasinh`
 (`scales_flowjo_fasinh`), 33

`scale_x_logicle`, 34

`scale_y_flowCore_fasinh`
 (`scale_x_flowCore_fasinh`), 33

`scale_y_flowJo_biexp`
 (`scales_flowjo_biexp`), 32

`scale_y_flowjo_biexp`
 (`scales_flowjo_biexp`), 32

`scale_y_flowJo_fasinh`
 (`scales_flowjo_fasinh`), 33

`scale_y_flowjo_fasinh`
 (`scales_flowjo_fasinh`), 33

`scale_y_logicle` (`scale_x_logicle`), 34

`scales_flowjo_biexp`, 32

`scales_flowjo_fasinh`, 33

`show,ggcyto-method` (`print.ggcyto`), 30

`show,ggcyto_GatingLayout-method`
 (`print.ggcyto_GatingLayout`), 30

`show.ggcyto` (`print.ggcyto`), 30

`show.ggcyto_GatingLayout`
 (`print.ggcyto_GatingLayout`), 30

`stat_position`, 19, 35

`stats_null`, 35

`transform` (`transform-gate`), 37

`transform,filter-method`
 (`transform-gate`), 37

`transform,filterList-method`
 (`transform-gate`), 37

`transform-gate`, 37