

Package ‘scoreInvHap’

January 20, 2026

Title Get inversion status in predefined regions

Version 1.32.0

Maintainer Dolors Pelegri-Siso <dolors.pelegri@isglobal.org>

Description scoreInvHap can get the samples' inversion status of known inversions. scoreInvHap uses SNP data as input and requires the following information about the inversion: genotype frequencies in the different haplotypes, R2 between the region SNPs and inversion status and heterozygote genotypes in the reference. The package include this data for 21 inversions.

Depends R (>= 3.6.0)

License file LICENSE

Encoding UTF-8

LazyData true

RoxygenNote 7.1.1

Imports Biostrings, methods, snpStats, VariantAnnotation,
GenomicRanges, BiocParallel, graphics, SummarizedExperiment

Suggests testthat, knitr, BiocStyle, rmarkdown

VignetteBuilder knitr

biocViews SNP, Genetics, GenomicVariation

git_url <https://git.bioconductor.org/packages/scoreInvHap>

git_branch RELEASE_3_22

git_last_commit 3779f3d

git_last_commit_date 2025-10-29

Repository Bioconductor 3.22

Date/Publication 2026-01-19

Author Carlos Ruiz [aut],
Dolors Pelegrí [aut],
Juan R. Gonzalez [aut, cre]

Contents

adaptRefs	2
checkSNPs	2

classifSNPs	3
computeScore	4
correctAlleleTable	5
getAlleleTable	5
getGenotypesTable	6
getInvStatus	6
hetRefs	7
info	7
inversionGR	7
prepareMap	8
Refs	8
scoreInvHap	9
scoreInvHapRes	10
SNPsR2	12

Index 13

adaptRefs	<i>Adapt references to imputed data</i>
-----------	---

Description

Internal

Usage

```
adaptRefs(Refs, alleletable, haploid = FALSE)
```

Arguments

Refs	List with the allele frequencies
alleletable	Data.frame with the alleles per SNP (from getAlleleTable)
haploid	Logical. If TRUE, modify references for haploid samples

Value

List with the same values than Refs but adapted to imputation data

checkSNPs	<i>Check genotype object</i>
-----------	------------------------------

Description

This function checks the genotype object before passing the SNPs to ‘scoreInvHap’. The function removes SNPs with different alleles or different allele frequencies. Nonetheless, it is possible that these SNPs could be recovered after an examination of the results. Be aware that testing of allele frequencies might fail for small datasets.

Usage

```
checkSNPs(SNPobj, checkAlleleFreqs = TRUE)
```

Arguments

- | | |
|------------------|--|
| SNPobj | List with SNPs data from plink or VCF-class. |
| checkAlleleFreqs | Should allele frequencies be check (Default: TRUE) |

Value

List containing the SNPs prepared for scoreInvHap

- **genos**: Object with genotype data ready for scoreInvHap
- **wrongAlleles**: Character vector with the SNPs discarded due to having alleles different to reference
- **wrongFreqs**: Character vector with the SNPs discarded due to having allele frequencies different to reference

Examples

```
## Run method
if(require(VariantAnnotation)){
  vcf <- readVcf(system.file("extdata", "example.vcf", package = "scoreInvHap"), "hg19")
  resList <- checkSNPs(vcf)
  resList
}
```

classifSNPs

Get similarity scores and probability

Description

This function computes the similarity scores between the sample SNPs and the haplotype's reference.

Usage

```
classifSNPs(
  genos,
  R2,
  refs,
  alleletable,
  BPPARAM = BiocParallel::SerialParam()
)

classifSNPsImpute(genos, R2, refs, BPPARAM = BiocParallel::SerialParam())
```

Arguments

- | | |
|-------------|--|
| genos | Matrix with the samples genotypes. It is the result of getGenotypesTable |
| R2 | Vector with the R2 between the SNPs and the inversion status. |
| refs | List of matrices. Each matrix has, for an SNP, the frequencies of each genotype in the different haplotypes. |
| alleletable | Data frame with the reference alleles computed with getAlleleTable. |
| BPPARAM | A BiocParallelParam instance. Used to parallelize computation |

Details

classifSNPs computes, for each individual, similarity scores for all the present haplotypes. For each SNP, we compute as many similarity scores as haplotypes present in the reference. We have defined the similarity score as the frequency of this genotype in the different haplotype population. To compute the global similarity score, we have computed a mean of the scores by SNP weighted by the R2 between the SNP and the haplotype classification.

classifSNPsImpute is a version of classifSNPs that works with posterior probabilities of imputed genotypes.

Value

List with the results:

- scores: Matrix with the similarity scores of the individuals
- numSNPs: Vector with the number of SNPs used in each computation

computeScore	<i>Compute all similarity scores for a sample</i>
--------------	---

Description

Internal

Usage

```
computeScore(geno, refs, R2)
```

Arguments

geno	Vector with the sample genotypes. It is the result of getGenotypesTable
refs	List of matrices. Each matrix has, for an SNP, the frequencies of each genotype in the different haplotypes.
R2	Vector with the R2 between the SNPs and the inversion status

Value

List with the results:

- scores: Vector with the similarity scores of the sample
- numSNPs: Numeric with the number of SNPs used in the computation

correctAlleleTable	<i>Solve genotypes discrepancies</i>
--------------------	--------------------------------------

Description

This function tries to solve discrepancies between the reference and sample genotypes. The cause of these discrepancies is that samples and references have used different strands to codify the SNP. This function get the complement genotypes for the discordant SNPs and checks if discordancies are solved.

Usage

```
correctAlleleTable(alleleTable, hetRefs, map)
```

Arguments

alleleTable	Data.frame with the alleles per SNP (from getAlleleTable)
hetRefs	Character vector with the heterozygous genotypes in the reference.
map	Data.frame with the annotation of the SNPs (from plink format)

Value

alleleTable without discrepancies between these genotypes and the references.

getAlleleTable	<i>Compute the allele table</i>
----------------	---------------------------------

Description

Get a data.frame that maps the numeric genotype of a SNPmatrix (0, 1, 2) into the real genotype. Heterozygous genotypes are ordered alphabetically.

Usage

```
getAlleleTable(map)
```

Arguments

map	Data.frame with the annotation of the SNPs (from plink format)
-----	--

Value

Data.frame with genotypes map

getGenotypesTable	<i>Get genotypes table</i>
-------------------	----------------------------

Description

Get a matrix with the sample genotypes from all SNP.

Usage

```
getGenotypesTable(geno, allele)
```

Arguments

geno	Snpmatrix (from plink format)
allele	Data.frame with the alleles per SNP (from getAlleleTable)

Value

Character matrix with the samples genotypes

getInvStatus	<i>Get the inversion status of a sample</i>
--------------	---

Description

This function estimates the inversion status of the samples using the probabilities computed in `classifSNPs`

Usage

```
getInvStatus(scores)
```

Arguments

scores	Matrix of probabilities (from <code>classifSNPs</code>)
--------	--

Value

List with the results:

- class: Vector with the most probable classification
- certainty: Vector with the certainty of the most probable classification

hetRefs	<i>Heterozygote genotypes in the references</i>
---------	---

Description

Dataset with the heterozygote genotypes of all the SNPs used in any of the references. This dataset include all the SNPs that are present inside the inversion's region in 1000 Genomes Phase 3.

Usage

hetRefs

Format

List of character vectors with the heterozygous genotypes of the SNPs present included the region of 21 inversions. Each element is named with the SNPs names.

info	<i>SNP reference description</i>
------	----------------------------------

Description

Description of the SNPs included in scoreInvHap references. The description includes the coordinates in hg19, the dbSNP identifier, the reference and alternative allele and the allele frequency in the European Samples of 1000 Genomes.

Usage

info

Format

data.frame

inversionGR	<i>Inversions' description</i>
-------------	--------------------------------

Description

Description of the 21 human inversions whose references are included in scoreInvHap. The description includes the cytogenic location, the coordinates in hg19, the number of alleles and the number of SNPs with a MAF > 5 Samples of 1000 Genomes.

Usage

inversionGR

Format

GenomicRanges with the inversions' description in the metadata

prepareMap	<i>Modify feature data from VCF</i>
------------	-------------------------------------

Description

Internal. Modify feature data from VCF to comply with scoreInvHap requirements.

Usage

```
prepareMap(vcf)
```

Arguments

vcf	VCF object
-----	------------

Value

Data.frame with the feature data

Refs	<i>Genotype frequency in references</i>
------	---

Description

Dataset with the genotype frequencies in the different haplotype populations. These frequencies have been computed using the European samples of 1000 Genomes Phase 3 data. Real inversion status have been obtained from invFEST and 1000Genomes.

Usage

```
Refs
```

Format

List of matrices for 20 inversions. Each matrix has the frequency of each genotype in each haplotype.

scoreInvHap	<i>scoreInvHap: package to get inversion status of predefined regions.</i>
-------------	--

Description

scoreInvHap can get the samples' inversion status of known inversions. scoreInvHap uses SNP data as input and requires the following information about the inversion: genotype frequencies in the different inversion groups, R2 between the region SNPs and inversion status, heterozygote genotypes in the reference, allele frequencies in the reference population and inversion frequencies. The package include this data for 21 inversions.

This is the main function of 'scoreInvHap' package. This function accepts SNPs data in a plink or a VCF format and compute the inversion prediction. The list of available inversions is included in a GenomicRanges called 'inversionGR'.

Usage

```
scoreInvHap(
  SNPlist,
  inv = NULL,
  SNPsR2,
  hetRefs,
  Refs,
  R2 = 0,
  probs = FALSE,
  BPPARAM = BiocParallel::SerialParam(),
  verbose = FALSE
)
```

Arguments

SNPlist	List with SNPs data from plink or VCF-class.
inv	Character with the name of the inversion to genotype. The available inversions are included in a table in the main vignette.
SNPsR2	Vector with the R2 of the SNPs of the region
hetRefs	Vector with the heterozygote form of the SNP in the inversion
Refs	List with the allele frequencies in the references
R2	Vector with the R2 between the SNPs and the inversion status
probs	Logical. If TRUE, scores are computed using posterior probabilities. If FALSE, scores are computed using best guess. Only applied when SNPlist is a VCF.
BPPARAM	A BiocParallelParam instance. Used to parallelize computation
verbose	Should message be shown?

Value

A scoreInvHap object

Examples

```
# See list of inversions
data(inversionGR)
inversionGR

## Run method
if(require(VariantAnnotation)){
  vcf <- readVcf(system.file("extdata", "example.vcf", package = "scoreInvHap"), "hg19")
  res <- scoreInvHap(vcf, inv = "inv7_005")
}
```

scoreInvHapRes	<i>scoreInvHapRes</i> instances
----------------	---------------------------------

Description

Container with the results of the classification pipeline

Usage

```
## S4 method for signature 'scoreInvHapRes'
classification(object, minDiff = 0, callRate = 0, inversion = TRUE)

## S4 method for signature 'scoreInvHapRes'
certainty(object)

## S4 method for signature 'scoreInvHapRes'
diffscores(object)

## S4 method for signature 'scoreInvHapRes'
maxscores(object)

## S4 method for signature 'scoreInvHapRes'
numSNPs(object)

## S4 method for signature 'scoreInvHapRes'
plotCallRate(object, callRate = 0.9, ...)

## S4 method for signature 'scoreInvHapRes'
plotScores(object, minDiff = 0.1, ...)

## S4 method for signature 'scoreInvHapRes'
propSNPs(object)

## S4 method for signature 'scoreInvHapRes'
scores(object)
```

Arguments

object	scoreInvHapRes
minDiff	Numeric with the threshold of the minimum difference between the top and the second score. Used to filter samples.
callRate	Numeric with the threshold of the minimum call rate of the samples. Used to filter samples.
inversion	Logical. If true, haplotypes classification is adapted to return inversion status. (Default: TRUE)
...	Further parameters passed to plot function.

Value

A scoreInvHapRes instance

Methods (by generic)

- classification: Get classification
- certainty: Get classification certainty
- diffscores: Get maximum similarity scores
- maxscores: Get maximum similarity scores
- numSNPs: Get number of SNPs used in computation
- plotCallRate: Plot call rate based QC
- plotScores: Plot scores based QC
- propSNPs: Get proportions of SNPs used in computation
- scores: Get similarity scores

Slots

classification Factor with the individuals classification
 scores Simmilarity scores for the different haplotypes.
 numSNPs Numeric with SNPs used to compute the scores.
 certainty Numeric with the certainty of the classification for each individual.

Examples

```
if(require(VariantAnnotation)){
  vcf <- readVcf(system.file("extdata", "example.vcf", package = "scoreInvHap"), "hg19")

  ## Create scoreInvHapRes class from pipeline
  res <- scoreInvHap(vcf, inv = "inv7_005")

  ## Print object
  res

  ## Get haplotype classification
  classification(res)

  ## Get similiraty scores
  scores(res)
}
```

SNPsR2*R2 between the SNPs and the inversion status*

Description

Dataset with R2 between the SNPs and the inversion status. This values are used to weigh similarity scores. These values have been computed using the European samples of 1000 Genomes Phase 3 data. Real inversion status have been estimated using invClust.

Usage

SNPsR2

Format

List of numeric vectors for 21 inversions

Index

* datasets

- hetRefs, [7](#)
- info, [7](#)
- inversionGR, [7](#)
- Refs, [8](#)
- SNPsR2, [12](#)

adaptRefs, [2](#)

certainty (scoreInvHapRes), [10](#)
certainty, scoreInvHapRes-method
(scoreInvHapRes), [10](#)
checkSNPs, [2](#)
classification (scoreInvHapRes), [10](#)
classification, scoreInvHapRes-method
(scoreInvHapRes), [10](#)
classifSNPs, [3](#)
classifSNPsImpute (classifSNPs), [3](#)
computeScore, [4](#)
correctAlleleTable, [5](#)

diffscores (scoreInvHapRes), [10](#)
diffscores, scoreInvHapRes-method
(scoreInvHapRes), [10](#)

getAlleleTable, [5](#)
getGenotypesTable, [6](#)
getInvStatus, [6](#)

hetRefs, [7](#)

info, [7](#)
inversionGR, [7](#)

maxscores (scoreInvHapRes), [10](#)
maxscores, scoreInvHapRes-method
(scoreInvHapRes), [10](#)

numSNPs (scoreInvHapRes), [10](#)
numSNPs, scoreInvHapRes-method
(scoreInvHapRes), [10](#)

plotCallRate (scoreInvHapRes), [10](#)
plotCallRate, scoreInvHapRes-method
(scoreInvHapRes), [10](#)

plotScores (scoreInvHapRes), [10](#)
plotScores, scoreInvHapRes-method
(scoreInvHapRes), [10](#)
prepareMap, [8](#)
propSNPs (scoreInvHapRes), [10](#)
propSNPs, scoreInvHapRes-method
(scoreInvHapRes), [10](#)

Refs, [8](#)

scoreInvHap, [9](#)
scoreInvHapRes, [10](#)
scoreInvHapRes-class (scoreInvHapRes),
[10](#)
scoreInvHapRes-methods
(scoreInvHapRes), [10](#)
scores (scoreInvHapRes), [10](#)
scores, scoreInvHapRes-method
(scoreInvHapRes), [10](#)
SNPsR2, [12](#)