

# Package ‘SCOPE’

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**Type** Package

**Title** A normalization and copy number estimation method for single-cell DNA sequencing

**Version** 1.19.0

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**Description** Whole genome single-cell DNA sequencing (scDNA-seq) enables characterization of copy number profiles at the cellular level. This circumvents the averaging effects associated with bulk-tissue sequencing and has increased resolution yet decreased ambiguity in deconvolving cancer subclones and elucidating cancer evolutionary history. ScDNA-seq data is, however, sparse, noisy, and highly variable even within a homogeneous cell population, due to the biases and artifacts that are introduced during the library preparation and sequencing procedure. Here, we propose SCOPE, a normalization and copy number estimation method for scDNA-seq data. The distinguishing features of SCOPE include: (i) utilization of cell-specific Gini coefficients for quality controls and for identification of normal/diploid cells, which are further used as negative control samples in a Poisson latent factor model for normalization; (ii) modeling of GC content bias using an expectation-maximization algorithm embedded in the Poisson generalized linear models, which accounts for the different copy number states along the genome; (iii) a cross-sample iterative segmentation procedure to identify breakpoints that are shared across cells from the same genetic background.

**Depends** R (>= 3.6.0), GenomicRanges, IRanges, Rsamtools, GenomeInfoDb, BSgenome.Hsapiens.UCSC.hg19

**Imports** stats, grDevices, graphics, utils, DescTools, RColorBrewer, gplots, foreach, parallel, doParallel, DNACopy, BSgenome, Biostrings, BiocGenerics, S4Vectors

**Suggests** knitr, rmarkdown, WGSmapp, BSgenome.Hsapiens.UCSC.hg38, BSgenome.Mmusculus.UCSC.mm10, testthat (>= 2.1.0)

**VignetteBuilder** knitr

**biocViews** SingleCell, Normalization, CopyNumberVariation, Sequencing, WholeGenome, Coverage, Alignment, QualityControl, DataImport, DNASEq

**License** GPL-2

**LazyData** true

**RoxygenNote** 6.1.1**Encoding** UTF-8**git\_url** <https://git.bioconductor.org/packages/SCOPE>**git\_branch** devel**git\_last\_commit** 8calafd**git\_last\_commit\_date** 2024-10-29**Repository** Bioconductor 3.21**Date/Publication** 2025-04-09

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---

coverageObj.scopeDemo *Pre-stored coverageObj.scope data for demonstration purposes*

---

## Description

Pre-stored coverageObj.scope data for demonstration purposes

**Usage**

```
coverageObj.scopeDemo
```

**Format**

Pre-computed using whole genome sequencing data of three single cells from 10X Genomics Single-Cell CNV solution

---

get_bam_bed	<i>Get bam file directories, sample names, and whole genomic bins</i>
-------------	---

---

**Description**

Get bam file directories, sample names, and whole genomic bins from .bed file

**Usage**

```
get_bam_bed(bamdir, sampname, hgrep = "hg19", resolution = 500,
            sex = FALSE)
```

**Arguments**

bamdir	vector of the directory of a bam file. Should be in the same order as sample names in sampname.
sampname	vector of sample names. Should be in the same order as bam directories in bamdir.
hgref	reference genome. This should be 'hg19', 'hg38' or 'mm10'. Default is human genome hg19.
resolution	numeric value of fixed bin-length. Default is 500. Unit is "kb".
sex	logical, whether to include sex chromosomes. Default is FALSE.

**Value**

A list with components

bamdir	A vector of bam directories
sampname	A vector of sample names
ref	A GRanges object specifying whole genomic bin positions

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```

library(WGSmapp)
library(BSgenome.Hsapiens.UCSC.hg38)
bamfolder <- system.file('extdata', package = 'WGSmapp')
bamFile <- list.files(bamfolder, pattern = '*.dedup.bam$')
bamdir <- file.path(bamfolder, bamFile)
samname_raw <- sapply(strsplit(bamFile, '.', fixed = TRUE), '[', 1)
bambedObj <- get_bam_bed(bamdir = bamdir, samname = samname_raw,
                        href = "hg38")

bamdir <- bambedObj$bamdir
samname_raw <- bambedObj$samname
ref_raw <- bambedObj$ref

```

---

get\_coverage\_scDNA      *Get read coverage from single-cell DNA sequencing*

---

**Description**

Get read coverage for each genomic bin across all single cells from scDNA-seq. Blacklist regions, such as segmental duplication regions and gaps near telomeres/centromeres will be masked prior to getting coverage.

**Usage**

```
get_coverage_scDNA(bambedObj, mapqthres, seq, href = "hg19")
```

**Arguments**

bambedObj	object returned from get_bam_bed
mapqthres	mapping quality threshold of reads
seq	the sequencing method to be used. This should be either 'paired-end' or 'single-end'
href	reference genome. This should be 'hg19', 'hg38' or 'mm10'. Default is human genome hg19.

**Value**

Y	Read depth matrix
---	-------------------

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

## Examples

```
library(WGSmapp)
library(BSgenome.Hsapiens.UCSC.hg38)
bamfolder <- system.file('extdata', package = 'WGSmapp')
bamFile <- list.files(bamfolder, pattern = '*.dedup.bam$')
bamdir <- file.path(bamfolder, bamFile)
samname_raw <- sapply(strsplit(bamFile, '.', fixed = TRUE), '[', 1)
bambedObj <- get_bam_bed(bamdir = bamdir,
                        samname = samname_raw,
                        hgrep = "hg38")

# Getting raw read depth
coverageObj <- get_coverage_scDNA(bambedObj,
                                  mapqthres = 40,
                                  seq = 'paired-end',
                                  hgrep = "hg38")

Y_raw <- coverageObj$Y
```

---

get\_gc

*Compute GC content*

---

## Description

Compute GC content for each bin

## Usage

```
get_gc(ref, hgrep = "hg19")
```

## Arguments

ref	GRanges object returned from <code>get_bam_bed</code>
hgrep	reference genome. This should be 'hg19', 'hg38' or 'mm10'. Default is human genome hg19.

## Value

gc	Vector of GC content for each bin/target
----	--

## Author(s)

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
## Not run:
library(WGSmapp)
library(BSgenome.Hsapiens.UCSC.hg38)
bamfolder <- system.file('extdata', package = 'WGSmapp')
bamFile <- list.files(bamfolder, pattern = '*.dedup.bam$')
bamdir <- file.path(bamfolder, bamFile)
samname_raw <- sapply(strsplit(bamFile, '.'), fixed = TRUE), '[' , 1)
bambedObj <- get_bam_bed(bamdir = bamdir,
                        samname = samname_raw,
                        hgrep = "hg38")

bamdir <- bambedObj$bamdir
samname_raw <- bambedObj$samname
ref_raw <- bambedObj$ref

gc <- get_gc(ref_raw, hgrep = "hg38")

## End(Not run)
```

---

get\_gini

---

*Compute Gini coefficients for single cells*


---

**Description**

Gini index is defined as two times the area between the Lorenz curve and the diagonal.

**Usage**

```
get_gini(Y)
```

**Arguments**

Y                      raw read depth matrix after quality control procedure

**Value**

Gini                      Vector of Gini coefficients for single cells from scDNA-seq

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
Gini <- get_gini(Y_sim)
```

---

get_mapp	<i>Compute mappability</i>
----------	----------------------------

---

### Description

Compute mappability for each bin. Note that scDNA sequencing is whole-genome amplification and the mappability score is essential to determine variable binning method. Mappability track for 100-mers on the GRCh37/hg19 human reference genome from ENCODE is pre-saved. Compute the mean of mappability scores that overlapped reads map to bins, weighted by the width of mappability tracks on the genome reference. Use liftOver utility to calculate mappability for hg38, which is pre-saved as well. For mm10, there are two workarounds: 1) set all mappability to 1 to avoid extensive computation; 2) adopt QC procedures based on annotation results, e.g., filter out bins within black list regions, which generally have low mappability.

### Usage

```
get_mapp(ref, hgrep = "hg19")
```

### Arguments

ref	GRanges object returned from get_bam_bed
hgrep	reference genome. This should be 'hg19', 'hg38' or 'mm10'. Default is human genome hg19.

### Value

mapp	Vector of mappability for each bin/target
------	---

### Author(s)

Rujin Wang <rujin@email.unc.edu>

### Examples

```
## Not run:
library(WGSmapp)
library(BSgenome.Hsapiens.UCSC.hg38)
bamfolder <- system.file('extdata', package = 'WGSmapp')
bamFile <- list.files(bamfolder, pattern = '*.dedup.bam$')
bamdir <- file.path(bamfolder, bamFile)
samname_raw <- sapply(strsplit(bamFile, '.', fixed = TRUE), '[', 1)
bambedObj <- get_bam_bed(bamdir = bamdir,
                        samname = samname_raw,
                        hgrep = "hg38")

bamdir <- bambedObj$bamdir
samname_raw <- bambedObj$samname
ref_raw <- bambedObj$ref
```

```
mapp <- get_mapp(ref_raw, hgrep = "hg38")  
## End(Not run)
```

---

get\_samp\_QC                      *Get QC metrics for single cells*

---

### Description

Perform QC step on single cells.

### Usage

```
get_samp_QC(bambedObj)
```

### Arguments

bambedObj                      object returned from get\_bam\_bed

### Value

QCmetric                      A matrix containing total number/proportion of reads, total number/proportion of mapped reads, total number/proportion of mapped non-duplicate reads, and number/proportion of reads with mapping quality greater than 20

### Author(s)

Rujin Wang <rujin@email.unc.edu>

### Examples

```
library(WGSmapp)  
library(BSgenome.Hsapiens.UCSC.hg38)  
bamfolder <- system.file('extdata', package = 'WGSmapp')  
bamFile <- list.files(bamfolder, pattern = '*.dedup.bam$')  
bamdir <- file.path(bamfolder, bamFile)  
samname_raw <- sapply(strsplit(bamFile, '.'), fixed = TRUE), '[' , 1)  
bambedObj <- get_bam_bed(bamdir = bamdir,  
                          samname = samname_raw,  
                          hgrep = "hg38")  
QCmetric_raw = get_samp_QC(bambedObj)
```



---

iCN_sim	<i>A post cross-sample segmentation integer copy number matrix returned by SCOPE in the demo</i>
---------	--

---

**Description**

A post cross-sample segmentation integer copy number matrix returned by SCOPE in the demo

**Usage**

```
iCN_sim
```

**Format**

A post cross-sample segmentation integer copy number matrix of five toy cells returned by SCOPE

---

initialize_ploidy	<i>Ploidy pre-initialization</i>
-------------------	----------------------------------

---

**Description**

Pre-estimate ploidies across all cells

**Usage**

```
initialize_ploidy(Y, Yhat, ref, maxPloidy = 6, minPloidy = 1.5,
                 minBinWidth = 5, SoS.plot = FALSE)
```

**Arguments**

Y	raw read depth matrix after quality control procedure
Yhat	normalized read depth matrix
ref	GRanges object after quality control procedure
maxPloidy	maximum ploidy candidate. Defalut is 6
minPloidy	minimum ploidy candidate. Defalut is 1.5
minBinWidth	the minimum number of bins for a changed segment. Defalut is 5
SoS.plot	logical, whether to generate ploidy pre-estimation plots. Default is FALSE.

**Value**

ploidy.SoS	Vector of pre-estimated ploidies for each cell
------------	--

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```

Gini <- get_gini(Y_sim)

# first-pass CODEX2 run with no latent factors
normObj.sim <- normalize_codex2_ns_noK(Y_qc = Y_sim,
                                       gc_qc = ref_sim$gc,
                                       norm_index = which(Gini<=0.12))

Yhat.noK.sim <- normObj.sim$Yhat
beta.hat.noK.sim <- normObj.sim$beta.hat
fGC.hat.noK.sim <- normObj.sim$fGC.hat
N.sim <- normObj.sim$N

# Ploidy initialization
ploidy.sim <- initialize_ploidy(Y = Y_sim,
                              Yhat = Yhat.noK.sim,
                              ref = ref_sim)

ploidy.sim

```

---

```
initialize_ploidy_group
```

*Group-wise ploidy pre-initialization*

---

**Description**

Pre-estimate ploidies across cells with shared clonal memberships

**Usage**

```
initialize_ploidy_group(Y, Yhat, ref, groups,
                      maxPloidy = 6, minPloidy = 1.5,
                      minBinWidth = 5, SoS.plot = FALSE)
```

**Arguments**

Y	raw read depth matrix after quality control procedure
Yhat	normalized read depth matrix
ref	GRanges object after quality control procedure
groups	clonal membership labels for each cell
maxPloidy	maximum ploidy candidate. Default is 6
minPloidy	minimum ploidy candidate. Default is 1.5
minBinWidth	the minimum number of bins for a changed segment. Default is 5
SoS.plot	logical, whether to generate ploidy pre-estimation plots. Default is FALSE.

**Value**

ploidy.SoS	Vector of group-wise pre-estimated ploidies for each cell
------------	---

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
Gini <- get_gini(Y_sim)

# first-pass CODEX2 run with no latent factors
normObj.sim <- normalize_codex2_ns_noK(Y_qc = Y_sim,
                                       gc_qc = ref_sim$gc,
                                       norm_index = which(Gini<=0.12))

Yhat.noK.sim <- normObj.sim$Yhat
beta.hat.noK.sim <- normObj.sim$beta.hat
fGC.hat.noK.sim <- normObj.sim$fGC.hat
N.sim <- normObj.sim$N

# Group-wise ploidy initialization
clones <- c("normal", "tumor1", "normal", "tumor1", "tumor1")
ploidy.sim.group <- initialize_ploidy_group(Y = Y_sim, Yhat = Yhat.noK.sim,
                                           ref = ref_sim, groups = clones)

ploidy.sim.group
```

---

normalize\_codex2\_ns\_noK

*Normalization of read depth without latent factors under the case-control setting*

---

**Description**

Assuming that all reads are from diploid regions, fit a Poisson generalized linear model to normalize the raw read depth data from single-cell DNA sequencing, without latent factors under the case-control setting.

**Usage**

```
normalize_codex2_ns_noK(Y_qc, gc_qc, norm_index)
```

**Arguments**

Y_qc	read depth matrix after quality control
gc_qc	vector of GC content for each bin after quality control
norm_index	indices of normal/diploid cells

**Value**

A list with components

Yhat	A list of normalized read depth matrix
fGC.hat	A list of estimated GC content bias matrix
beta.hat	A list of estimated bin-specific bias vector
N	A vector of cell-specific library size factor, which is computed from the genome-wide read depth data

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
Gini <- get_gini(Y_sim)
# first-pass CODEX2 run with no latent factors
normObj.sim <- normalize_codex2_ns_noK(Y_qc = Y_sim,
                                       gc_qc = ref_sim$gc,
                                       norm_index = which(Gini<=0.12))
```

---

normalize_scope	<i>Normalization of read depth with latent factors using Expectation-Maximization algorithm under the case-control setting</i>
-----------------	--

---

**Description**

Fit a Poisson generalized linear model to normalize the raw read depth data from single-cell DNA sequencing, with latent factors under the case-control setting. Model GC content bias using an expectation-maximization algorithm, which accounts for the different copy number states.

**Usage**

```
normalize_scope(Y_qc, gc_qc, K, norm_index, T, ploidyInt,
               beta0, minCountQC = 20)
```

**Arguments**

Y_qc	read depth matrix after quality control
gc_qc	vector of GC content for each bin after quality control
K	Number of latent Poisson factors
norm_index	indices of normal/diploid cells

T	a vector of integers indicating number of CNV groups. Use BIC to select optimal number of CNV groups. If $T = 1$ , assume all reads are from normal regions so that EM algorithm is not implemented. Otherwise, we assume there is always a CNV group of heterozygous deletion and a group of null region. The rest groups are representative of different duplication states.
ploidyInt	a vector of initialized ploidy return from <code>initialize_ploidy</code> . Users are also allowed to provide prior-knowledge ploidies as the input and to manually tune a few cells that have poor fitting
beta0	a vector of initialized bin-specific biases returned from CODEX2 without latent factors
minCountQC	the minimum read coverage required for normalization and EM fitting. Defalut is 20

**Value**

A list with components

Yhat	A list of normalized read depth matrix with EM
alpha.hat	A list of absolute copy number matrix
fGC.hat	A list of EM estimated GC content bias matrix
beta.hat	A list of EM estimated bin-specific bias vector
g.hat	A list of estimated Poisson latent factor
h.hat	A list of estimated Poisson latent factor
AIC	AIC for model selection
BIC	BIC for model selection
RSS	RSS for model selection
K	Number of latent Poisson factors

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
Gini <- get_gini(Y_sim)

# first-pass CODEX2 run with no latent factors
normObj.sim <- normalize_codex2_ns_noK(Y_qc = Y_sim,
                                       gc_qc = ref_sim$gc,
                                       norm_index = which(Gini<=0.12))

Yhat.noK.sim <- normObj.sim$Yhat
beta.hat.noK.sim <- normObj.sim$beta.hat
fGC.hat.noK.sim <- normObj.sim$fGC.hat
N.sim <- normObj.sim$N

# Ploidy initialization
ploidy.sim <- initialize_ploidy(Y = Y_sim,
```

```

                                Yhat = Yhat.noK.sim,
                                ref = ref_sim)
ploidy.sim

normObj.scope.sim <- normalize_scope(Y_qc = Y_sim, gc_qc = ref_sim$gc,
                                    K = 1, ploidyInt = ploidy.sim,
                                    norm_index = which(Gini<=0.12), T = 1:5,
                                    beta0 = beta.hat.noK.sim)
Yhat.sim <- normObj.scope.sim$Yhat[[which.max(normObj.scope.sim$BIC)]]
fGC.hat.sim <- normObj.scope.sim$fGC.hat[[which.max(normObj.scope.sim$BIC)]]

```

---

normalize\_scope\_foreach

*Normalization of read depth with latent factors using Expectation-Maximization algorithm under the case-control setting in parallel*

---

## Description

Fit a Poisson generalized linear model to normalize the raw read depth data from single-cell DNA sequencing, with latent factors under the case-control setting. Model GC content bias using an expectation-maximization algorithm, which accounts for the different copy number states.

## Usage

```

normalize_scope_foreach(Y_qc, gc_qc, K, norm_index, T,
                       ploidyInt, beta0, minCountQC = 20, nCores = NULL)

```

## Arguments

Y_qc	read depth matrix after quality control
gc_qc	vector of GC content for each bin after quality control
K	Number of latent Poisson factors
norm_index	indices of normal/diploid cells
T	a vector of integers indicating number of CNV groups. Use BIC to select optimal number of CNV groups. If T = 1, assume all reads are from normal regions so that EM algorithm is not implemented. Otherwise, we assume there is always a CNV group of heterozygous deletion and a group of null region. The rest groups are representative of different duplication states.
ploidyInt	a vector of initialized ploidy return from initialize_ploidy. Users are also allowed to provide prior-knowledge ploidies as the input and to manually tune a few cells that have poor fitting
beta0	a vector of initialized bin-specific biases returned from CODEX2 without latent factors
minCountQC	the minimum read coverage required for normalization and EM fitting. Default is 20
nCores	number of cores to use. If NULL, number of cores is detected. Default is NULL.

**Value**

A list with components

Yhat	A list of normalized read depth matrix with EM
alpha.hat	A list of absolute copy number matrix
fGC.hat	A list of EM estimated GC content bias matrix
beta.hat	A list of EM estimated bin-specific bias vector
g.hat	A list of estimated Poisson latent factor
h.hat	A list of estimated Poisson latent factor
AIC	AIC for model selection
BIC	BIC for model selection
RSS	RSS for model selection
K	Number of latent Poisson factors

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
Gini <- get_gini(Y_sim)

# first-pass CODEX2 run with no latent factors
normObj.sim <- normalize_codex2_ns_noK(Y_qc = Y_sim,
                                       gc_qc = ref_sim$gc,
                                       norm_index = which(Gini<=0.12))

Yhat.noK.sim <- normObj.sim$Yhat
beta.hat.noK.sim <- normObj.sim$beta.hat
fGC.hat.noK.sim <- normObj.sim$fGC.hat
N.sim <- normObj.sim$N

# Ploidy initialization
ploidy.sim <- initialize_ploidy(Y = Y_sim,
                              Yhat = Yhat.noK.sim,
                              ref = ref_sim)

ploidy.sim

# Specify nCores = 2 only for checking examples
normObj.scope.sim <- normalize_scope_foreach(Y_qc = Y_sim,
                                             gc_qc = ref_sim$gc,
                                             K = 1, ploidyInt = ploidy.sim,
                                             norm_index = which(Gini<=0.12), T = 1:5,
                                             beta0 = beta.hat.noK.sim, nCores = 2)
Yhat.sim <- normObj.scope.sim$Yhat[[which.max(normObj.scope.sim$BIC)]]
fGC.hat.sim <- normObj.scope.sim$fGC.hat[[which.max(normObj.scope.sim$BIC)]]
```

---

normalize\_scope\_group *Group-wise normalization of read depth with latent factors using Expectation-Maximization algorithm and shared clonal memberships*

---

### Description

Fit a Poisson generalized linear model to normalize the raw read depth data from single-cell DNA sequencing, with latent factors and shared clonal memberships. Model GC content bias using an expectation-maximization algorithm, which accounts for clonal specific copy number states.

### Usage

```
normalize_scope_group(Y_qc, gc_qc, K, norm_index, groups, T,
                    ploidyInt, beta0, minCountQC = 20)
```

### Arguments

Y_qc	read depth matrix after quality control
gc_qc	vector of GC content for each bin after quality control
K	Number of latent Poisson factors
norm_index	indices of normal/diploid cells using group/clone labels
groups	clonal membership labels for each cell
T	a vector of integers indicating number of CNV groups. Use BIC to select optimal number of CNV groups. If $T = 1$ , assume all reads are from normal regions so that EM algorithm is not implemented. Otherwise, we assume there is always a CNV group of heterozygous deletion and a group of null region. The rest groups are representative of different duplication states.
ploidyInt	a vector of group-wise initialized ploidy return from <code>initialize_ploidy_group</code> . Users are also allowed to provide prior-knowledge ploidies as the input and to manually tune a few cells/clones that have poor fitting
beta0	a vector of initialized bin-specific biases returned from CODEX2 without latent factors
minCountQC	the minimum read coverage required for normalization and EM fitting. Defalut is 20

### Value

A list with components

Yhat	A list of normalized read depth matrix with EM
alpha.hat	A list of absolute copy number matrix
fGC.hat	A list of EM estimated GC content bias matrix
beta.hat	A list of EM estimated bin-specific bias vector
g.hat	A list of estimated Poisson latent factor



h.hat	A list of estimated Poisson latent factor
AIC	AIC for model selection
BIC	BIC for model selection
RSS	RSS for model selection
K	Number of latent Poisson factors

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
Gini <- get_gini(Y_sim)

# first-pass CODEX2 run with no latent factors
normObj.sim <- normalize_codex2_ns_noK(Y_qc = Y_sim,
                                       gc_qc = ref_sim$gc,
                                       norm_index = which(Gini<=0.12))

Yhat.noK.sim <- normObj.sim$Yhat
beta.hat.noK.sim <- normObj.sim$beta.hat
fGC.hat.noK.sim <- normObj.sim$fGC.hat
N.sim <- normObj.sim$N

# Group-wise ploidy initialization
clones <- c("normal", "tumor1", "normal", "tumor1", "tumor1")
ploidy.sim.group <- initialize_ploidy_group(Y = Y_sim, Yhat = Yhat.noK.sim,
                                           ref = ref_sim, groups = clones)
ploidy.sim.group

normObj.scope.sim.group <- normalize_scope_group(Y_qc = Y_sim,
                                                gc_qc = ref_sim$gc,
                                                K = 1, ploidyInt = ploidy.sim.group,
                                                norm_index = which(clones=="normal"),
                                                groups = clones,
                                                T = 1:5,
                                                beta0 = beta.hat.noK.sim)
Yhat.sim.group <- normObj.scope.sim.group$Yhat[[which.max(
  normObj.scope.sim.group$BIC)]]
fGC.hat.sim.group <- normObj.scope.sim.group$fGC.hat[[which.max(
  normObj.scope.sim.group$BIC)]]
```

---

normObj.scopeDemo

*Pre-stored normObj.scope data for demonstration purposes*

---

**Description**

Pre-stored normObj.scope data for demonstration purposes

**Usage**

```
normObj.scopeDemo
```

**Format**

Pre-computed by SCOPE using pre-stored data Y\_sim

---

perform_qc	<i>Quality control for cells and bins</i>
------------	---

---

**Description**

Perform QC step on single cells and bins.

**Usage**

```
perform_qc(Y_raw, sampname_raw, ref_raw, QCmetric_raw,
           cov_thresh = 0, minCountQC = 20,
           mapq20_thresh = 0.3, mapp_thresh = 0.9,
           gc_thresh = c(20, 80), nMAD = 3)
```

**Arguments**

Y_raw	raw read count matrix returned from <a href="#">get_coverage_scDNA</a>
sampname_raw	sample names for quality control returned from <a href="#">get_bam_bed</a>
ref_raw	raw GRanges object with corresponding GC content and mappability for quality control returned from <a href="#">get_bam_bed</a>
QCmetric_raw	a QC metric for single cells returned from <a href="#">get_samp_QC</a>
cov_thresh	scalar variable specifying the lower bound of read count summation of each cell. Default is 0
minCountQC	the minimum read coverage required for normalization and EM fitting. Default is 20
mapq20_thresh	scalar variable specifying the lower threshold of proportion of reads with mapping quality greater than 20. Default is 0.3
mapp_thresh	scalar variable specifying mappability of each genomic bin. Default is 0.9
gc_thresh	vector specifying the lower and upper bound of GC content threshold for quality control. Default is 20-80
nMAD	scalar variable specifying the number of MAD from the median of total read counts adjusted by library size for each cell. Default is 3

**Value**

A list with components

Y	read depth matrix after quality control
sampname	sample names after quality control
ref	A GRanges object specifying whole genomic bin positions after quality control
QCmetric	A data frame of QC metric for single cells after quality control

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
Y_raw <- coverageObj.scopeDemo$Y
sampname_raw <- rownames(QCmetric.scopeDemo)
ref_raw <- ref.scopeDemo
QCmetric_raw <- QCmetric.scopeDemo
qcObj <- perform_qc(Y_raw = Y_raw, sampname_raw = sampname_raw,
                   ref_raw = ref_raw, QCmetric_raw = QCmetric_raw)
```

---

plot_EM_fit	<i>Visualize EM fitting for each cell.</i>
-------------	--

---

**Description**

A pdf file containing EM fitting results and plots is generated.

**Usage**

```
plot_EM_fit(Y_qc, gc_qc, norm_index, T, ploidyInt, beta0,
            minCountQC = 20, filename)
```

**Arguments**

Y_qc	read depth matrix across all cells after quality control
gc_qc	vector of GC content for each bin after quality control
norm_index	indices of normal/diploid cells
T	a vector of integers indicating number of CNV groups. Use BIC to select optimal number of CNV groups. If T = 1, assume all reads are from normal regions so that EM algorithm is not implemented. Otherwise, we assume there is always a CNV group of heterozygous deletion and a group of null region. The rest groups are representative of different duplication states.
ploidyInt	a vector of initialized ploidy return from initialize_ploidy

beta0            a vector of initialized bin-specific biases returned from CODEX2 without latent factors

minCountQC      the minimum read coverage required for EM fitting. Defalut is 20

filename         the name of output pdf file

### Value

pdf file with EM fitting results and two plots: log likelihood, and BIC versus the number of CNV groups.

### Author(s)

Rujin Wang <rujin@email.unc.edu>

### Examples

```
Gini <- get_gini(Y_sim)
# first-pass CODEX2 run with no latent factors
normObj.sim <- normalize_codex2_ns_noK(Y_qc = Y_sim,
                                       gc_qc = ref_sim$gc,
                                       norm_index = which(Gini<=0.12))

Yhat.noK.sim <- normObj.sim$Yhat
beta.hat.noK.sim <- normObj.sim$beta.hat
fGC.hat.noK.sim <- normObj.sim$fGC.hat
N.sim <- normObj.sim$N

# Ploidy initialization
ploidy.sim <- initialize_ploidy(Y = Y_sim,
                              Yhat = Yhat.noK.sim,
                              ref = ref_sim)

ploidy.sim

plot_EM_fit(Y_qc = Y_sim, gc_qc = ref_sim$gc,
            norm_index = which(Gini<=0.12), T = 1:7,
            ploidyInt = ploidy.sim,
            beta0 = beta.hat.noK.sim,
            filename = 'plot_EM_fit_demo.pdf')
```

---

plot\_iCN

*Plot post-segmentation copy number profiles of integer values*

---

### Description

Show heatmap of inferred integer copy-number profiles by SCOPE with cells clustered by hierarchical clustering

**Usage**

```
plot_iCN(iCNmat, ref, Gini, annotation = NULL,  
        plot.dendrogram = TRUE, show.names = FALSE, filename)
```

**Arguments**

iCNmat	inferred integer copy-number matrix by SCOPE, with each column being a cell and each row being a genomic bin
ref	GRanges object after quality control procedure
Gini	vector of Gini coefficients for each cell, with the same order as that of cells in columns of iCNmat
annotation	vector of annotation for each cell, with the same order as that of cells in columns of iCNmat. Default is NULL.
plot.dendrogram	logical, whether to plot the dendrogram. Default is TRUE.
show.names	logical, whether to show cell names by y axis. Default is FALSE.
filename	name of the output png file

**Value**

png file with integer copy-number profiles across single cells with specified annotations

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
Gini <- get_gini(Y_sim)  
plot_iCN(iCNmat = iCN_sim,  
        ref = ref_sim,  
        Gini = Gini,  
        filename = 'plot_iCN_demo')
```

---

QCmetric.scopeDemo      *Pre-stored QCmetric data for demonstration purposes*

---

**Description**

Pre-stored QCmetric data for demonstration purposes

**Usage**

```
QCmetric.scopeDemo
```

**Format**

Pre-computed using whole genome sequencing data of three single cells from 10X Genomics Single-Cell CNV solution

---

ref.scopeDemo	<i>Pre-stored 500kb-size reference genome for demonstration purposes</i>
---------------	--

---

**Description**

Pre-stored 500kb-size reference genome for demonstration purposes

**Usage**

ref.scopeDemo

**Format**

Pre-computed using whole genome sequencing data with GC content and mappability scores

---

ref_sim	<i>A reference genome in the toy dataset</i>
---------	--

---

**Description**

A reference genome in the toy dataset

**Usage**

ref\_sim

**Format**

A GRanges object with 1544 bins and 1 metadata column of GC content

---

segment_CBScs	<i>Cross-sample segmentation</i>
---------------	----------------------------------

---

**Description**

SCOPE offers a cross-sample Poisson likelihood-based recursive segmentation, enabling shared breakpoints across cells from the same genetic background.

**Usage**

```
segment_CBScs(Y, Yhat, sampname, ref, chr,
              mode = "integer", max.ns)
```

**Arguments**

Y	raw read depth matrix after quality control procedure
Yhat	normalized read depth matrix
sampname	vector of sample names
ref	GRanges object after quality control procedure
chr	chromosome name. Make sure it is consistent with the reference genome.
mode	format of returned copy numbers. Only integer mode is supported for scDNA-seq data.
max.ns	a number specifying how many rounds of nested structure searching would be performed. Defalut is 0.

**Value**

A list with components

poolcall	Cross-sample CNV callings indicating shared breakpoints
finalcall	Final cross-sample segmented callset of CNVs with genotyping results
image.orig	A matrix giving logarithm of normalized z-scores
image.seg	A matrix of logarithm of estimated copy number over 2
iCN	A matrix of inferred integer copy number profiles

**Author(s)**

Rujin Wang <rujin@email.unc.edu>

**Examples**

```
Yhat.sim <- normObj.scopeDemo$Yhat[[which.max(normObj.scopeDemo$BIC)]]
segment_cs_chr1 <- segment_CBScs(Y = Y_sim, Yhat = Yhat.sim,
                                sampname = colnames(Y_sim),
                                ref = ref_sim, chr = 'chr1', max.ns = 1)
```

---

Y\_sim

*A read count matrix in the toy dataset*

---

**Description**

A read count matrix in the toy dataset

**Usage**

Y\_sim

**Format**

A read count matrix with 1544 bins and 39 cells



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