

Package ‘methimpute’

January 9, 2025

Type Package

Title Imputation-guided re-construction of complete methylomes from WGBS data

Version 1.28.0

Author Aaron Taudt

Maintainer Aaron Taudt <aaron.taudt@gmail.com>

Description

This package implements functions for calling methylation for all cytosines in the genome.

Depends R (>= 3.4.0), GenomicRanges, ggplot2

Imports Rcpp (>= 0.12.4.5), methods, utils, grDevices, stats, GenomeInfoDb, IRanges, Biostrings, reshape2, minpack.lm, data.table

Suggests knitr, BiocStyle

LinkingTo Rcpp

License Artistic-2.0

LazyLoad yes

VignetteBuilder knitr

RoxygenNote 6.1.1

biocViews ImmunoOncology, Software, DNAMethylation, Epigenetics, HiddenMarkovModel, Sequencing, Coverage

git_url <https://git.bioconductor.org/packages/methimpute>

git_branch RELEASE_3_20

git_last_commit a8dc359

git_last_commit_date 2024-10-29

Repository Bioconductor 3.20

Date/Publication 2025-01-09

Contents

methimpute-package	2
arabidopsis_chromosomes	3
arabidopsis_genes	3
arabidopsis_TEs	3

arabidopsis_toydata	4
binning	4
binomialTestMethylation	5
callMethylation	6
callMethylationSeparate	7
collapseBins	8
distanceCorrelation	10
estimateTransDist	10
exportMethylome	11
extractCytosinesFromFASTA	12
getDistinctColors	13
getPosteriors	14
getStateColors	14
import	15
importRene	16
inflateMethylome	17
loadFromFiles	17
methimpute-objects	18
methimputeBinomialHMM	18
methimputeData	19
parameterScan	19
plotting	20
print.methimputeBinomialHMM	21
transCoord	22
Index	23

methimpute-package	<i>methIMPUTE: Imputation-guided methylation status calling for WGBS-seq</i>
--------------------	--

Description

methimpute is an R-package for methylation status calling in Whole-Genome Bisulfite-sequencing (WGBS-seq) data. Its powerful Hidden Markov model implementation enables imputation of methylation status calls for cytosines without any coverage.

Details

Please read the vignette for a tutorial on how to use this package. You can do this by typing `browseVignettes("methimpute")`. Here is an overview of all [plotting](#) functions.

Author(s)

Aaron Taudt

arabidopsis_chromosomes

Chromosome lengths for Arabidopsis

Description

A data.frame with chromosome lengths for Arabidopsis.

Format

A data.frame.

Examples

```
data(arabidopsis_chromosomes)
print(arabidopsis_chromosomes)
```

arabidopsis_genes

Gene coordinates for Arabidopsis (chr1)

Description

A [GRanges-class](#) object for demonstration purposes in examples of package [methimpute](#). The object contains gene coordinates of chr1 from Arabidopsis.

Format

A [GRanges-class](#) object.

Examples

```
data(arabidopsis_genes)
print(arabidopsis_genes)
```

arabidopsis_TEs

Transposable element coordinates for Arabidopsis (chr1)

Description

A [GRanges-class](#) object for demonstration purposes in examples of package [methimpute](#). The object contains transposable element coordinates of chr1 from Arabidopsis.

Format

A [GRanges-class](#) object.

Examples

```
data(arabidopsis_TEs)
print(arabidopsis_TEs)
```

arabidopsis_toydata *Toy data for Arabidopsis (200.000bp of chr1)*

Description

A `methimputeData` object for demonstration purposes in examples of package `methimpute`. The object contains the first 200.000 cytosines of chr1 from Arabidopsis.

Format

A `methimputeData` object.

Examples

```
data(arabidopsis_toydata)
print(arabidopsis_toydata)
```

binning *Methimpute binning functions*

Description

This page provides an overview of all `methimpute` binning functions.

Usage

```
binCounts(data, binsize)

binPositions(data, binsize)

binMethylome(data, binsize, contexts = "total", columns.average = NULL)
```

Arguments

<code>data</code>	A <code>GRanges-class</code> object with metadata columns 'context' and 'counts' (which is a matrix with columns 'methylated' and 'total').
<code>binsize</code>	The window size used for binning.
<code>contexts</code>	A character vector with contexts for which the binning will be done.
<code>columns.average</code>	A character vector with names of columns in data that should be averaged in bins.

Value

A `GRanges-class` object for `binCounts` and `binPositions`. A `list()` of `GRanges-class` objects for `binMethylome`.

Functions

- `binCounts`: Get the aggregated number of counts in each bin (no context).
- `binPositions`: Get the number of cytosines in each bin (total and per context).
- `binMethylome`: Get number of cytosines and aggregated counts for the specified contexts.

Examples

```
## Get some toy data
file <- system.file("data", "arabidopsis_toydata.RData",
                    package="methimpute")
data <- get(load(file))
print(data)
## Bin the data in various ways
binCounts(data, binsize=1000)
binPositions(data, binsize=1000)
binMethylome(data, binsize=1000, contexts=c("total", "CG"),
              columns.average=NULL)
```

binomialTestMethylation

Call methylation status

Description

Call methylation status of cytosines (or bins) with a binomial test.

Usage

```
binomialTestMethylation(data, conversion.rate, min.coverage = 3,
                        p.threshold = 0.05)
```

Arguments

<code>data</code>	A <code>methimputeData</code> object.
<code>conversion.rate</code>	A conversion rate between 0 and 1.
<code>min.coverage</code>	Minimum coverage to consider for the binomial test.
<code>p.threshold</code>	Significance threshold between 0 and 1.

Details

The function uses a binomial test with the specified `conversion.rate`. P-values are then multiple testing corrected with the Benjamini & Yekutieli procedure. Methylated positions are selected with the `p.threshold`.

Value

A vector with methylation statuses.

Examples

```
## Get some toy data
file <- system.file("data","arabidopsis_toydata.RData", package="methimpute")
data <- get(load(file))
data$binomial <- binomialTestMethylation(data, conversion.rate=0.998)
```

callMethylation	<i>Call methylation status</i>
-----------------	--------------------------------

Description

Call methylation status of cytosines (or bins) with a Hidden Markov Model.

Usage

```
callMethylation(data, fit.on.chrom = NULL, transDist = Inf, eps = 1,
  max.time = Inf, max.iter = Inf, count.cutoff = 500,
  verbosity = 1, num.threads = 2 + include.intermediate,
  initial.params = NULL, include.intermediate = FALSE,
  update = "independent", min.reads = 0)
```

Arguments

data	A methimputeData object.
fit.on.chrom	A character vector specifying the chromosomes on which the HMM will be fitted.
transDist	The decaying constant for the distance-dependent transition matrix. Either a single numeric or a named numeric vector, where the vector names correspond to the transition contexts. Such a vector can be obtained from estimateTransDist .
eps	Convergence threshold for the Baum-Welch algorithm.
max.time	Maximum running time in seconds for the Baum-Welch algorithm.
max.iter	Maximum number of iterations for the Baum-Welch algorithm.
count.cutoff	A cutoff for the counts to remove artificially high counts from mapping artifacts. Set to Inf to disable this filtering (not recommended).
verbosity	An integer from 1 to 5 specifying the verbosity of the fitting procedure. Values > 1 are only for debugging.
num.threads	Number of CPU to use for the computation. Parallelization is implemented on the number of states, which is 2 or 3, so setting num.threads > 3 will not give additional performance increase.
initial.params	A methimputeBinomialHMM object. This parameter is useful to continue the fitting procedure for a methimputeBinomialHMM object.
include.intermediate	A logical specifying whether or not the intermediate component should be included in the HMM.
update	One of c("independent", "constrained"). If update="independent" probability parameters for the binomial test will be updated independently. If update="constrained" the probability parameter of the intermediate component will be constrained to the mean of the unmethylated and the methylated component.

`min.reads` The minimum number of reads that a position must have to contribute in the Baum-Welch fitting procedure.

Details

The Hidden Markov model uses a binomial test for the emission densities. Transition probabilities are modeled with a distance dependent decay, specified by the parameter `transDist`.

Value

A `methimputeBinomialHMM` object.

Examples

```
## Get some toy data
file <- system.file("data", "arabidopsis_toydata.RData", package="methimpute")
data <- get(load(file))
print(data)
model <- callMethylation(data)
print(model)
```

callMethylationSeparate

Call methylation status

Description

Call methylation status of cytosines (or bins) with a separate Hidden Markov Model for each context.

Usage

```
callMethylationSeparate(data, fit.on.chrom = NULL, transDist = Inf,
  eps = 1, max.time = Inf, max.iter = Inf, count.cutoff = 500,
  verbosity = 1, num.threads = 2 + include.intermediate,
  initial.params = NULL, include.intermediate = FALSE,
  update = "independent", min.reads = 0)
```

Arguments

<code>data</code>	A <code>methimputeData</code> object.
<code>fit.on.chrom</code>	A character vector specifying the chromosomes on which the HMM will be fitted.
<code>transDist</code>	The decaying constant for the distance-dependent transition matrix. Either a single numeric or a named numeric vector, where the vector names correspond to the transition contexts. Such a vector can be obtained from <code>estimateTransDist</code> .
<code>eps</code>	Convergence threshold for the Baum-Welch algorithm.
<code>max.time</code>	Maximum running time in seconds for the Baum-Welch algorithm.
<code>max.iter</code>	Maximum number of iterations for the Baum-Welch algorithm.
<code>count.cutoff</code>	A cutoff for the counts to remove artificially high counts from mapping artifacts. Set to <code>Inf</code> to disable this filtering (not recommended).

verbosity	An integer from 1 to 5 specifying the verbosity of the fitting procedure. Values > 1 are only for debugging.
num.threads	Number of CPU to use for the computation. Parallelization is implemented on the number of states, which is 2 or 3, so setting num.threads > 3 will not give additional performance increase.
initial.params	A <code>methimputeBinomialHMM</code> object. This parameter is useful to continue the fitting procedure for a <code>methimputeBinomialHMM</code> object.
include.intermediate	A logical specifying wheter or not the intermediate component should be included in the HMM.
update	One of <code>c("independent", "constrained")</code> . If <code>update="independent"</code> probability parameters for the binomial test will be updated independently. If <code>update="constrained"</code> the probability parameter of the intermediate component will be constrained to the mean of the unmethylated and the methylated component.
min.reads	The minimum number of reads that a position must have to contribute in the Baum-Welch fitting procedure.

Details

The Hidden Markov model uses a binomial test for the emission densities. Transition probabilities are modeled with a distance dependent decay, specified by the parameter `transDist`.

Value

A `methimputeBinomialHMM` object.

Examples

```
## Get some toy data
file <- system.file("data", "arabidopsis_toydata.RData", package="methimpute")
data <- get(load(file))
print(data)
model <- callMethylationSeparate(data)
print(model)
```

collapseBins

Collapse consecutive bins

Description

The function will collapse consecutive bins which have, for example, the same combinatorial state.

Usage

```
collapseBins(data, column2collapseBy = NULL, columns2sumUp = NULL,
             columns2average = NULL, columns2getMax = NULL, columns2drop = NULL)
```


Arguments

<code>data</code>	A data.frame containing the genomic coordinates in the first three columns.
<code>column2collapseBy</code>	The number of the column which will be used to collapse all other inputs. If a set of consecutive bins has the same value in this column, they will be aggregated into one bin with adjusted genomic coordinates.
<code>columns2sumUp</code>	Column numbers that will be summed during the aggregation process.
<code>columns2average</code>	Column numbers that will be averaged during the aggregation process.
<code>columns2getMax</code>	Column numbers where the maximum will be chosen during the aggregation process.
<code>columns2drop</code>	Column numbers that will be dropped after the aggregation process.

Details

The following tables illustrate the principle of the collapsing:

Input data:

seqnames	start	end	column2collapseBy	moreColumns	columns2sumUp
chr1	0	199	2	1 10	1 3
chr1	200	399	2	2 11	0 3
chr1	400	599	2	3 12	1 3
chr1	600	799	1	4 13	0 3
chr1	800	999	1	5 14	1 3

Output data:

seqnames	start	end	column2collapseBy	moreColumns	columns2sumUp
chr1	0	599	2	1 10	2 9
chr1	600	999	1	4 13	1 6

Value

A data.frame.

Author(s)

Aaron Taudt

Examples

```
## Load example data
## Get an example multiHMM
data(arabidopsis_toydata)
df <- as.data.frame(arabidopsis_toydata)
shortdf <- collapseBins(df, column2collapseBy='context', columns2sumUp='width', columns2average=7:8)
```

distanceCorrelation *Distance correlation*

Description

Compute the distance correlation from a [methimputeData](#) object.

Usage

```
distanceCorrelation(data, distances = 0:50, separate.contexts = FALSE)
```

Arguments

`data` A [methimputeData](#) object.

`distances` An integer vector specifying the distances for which the correlation will be calculated.

`separate.contexts` A logical indicating whether contexts are treated separately. If set to TRUE, correlations will only be calculated between cytosines of the same context.

Value

A list() with an array containing the correlation values and the corresponding [ggplot](#).

Examples

```
## Get some toy data
file <- system.file("data", "arabidopsis_toydata.RData",
                    package="methimpute")
data <- get(load(file))
distcor <- distanceCorrelation(data)
print(distcor$plot)
```

estimateTransDist *transDist parameter*

Description

Obtain an estimate for the `transDist` parameter (used in function [callMethylation](#)) by fitting an exponential function to the supplied correlations (from [distanceCorrelation](#)).

Usage

```
estimateTransDist(distcor, skip = 2, plot.parameters = TRUE)
```

Arguments

distcor	The output produced by distanceCorrelation .
skip	Skip the first n cytosines for the fitting. This can be necessary to avoid periodicity artifacts due to the context definition.
plot.parameters	Whether to plot fitted parameters on to the plot or not.

Value

A list() with fitted transDist parameters and the corresponding [ggplot](#).

Examples

```
## Get some toy data
file <- system.file("data", "arabidopsis_toydata.RData",
                    package="methimpute")
data <- get(load(file))
distcor <- distanceCorrelation(data)
fit <- estimateTransDist(distcor)
print(fit)
```

exportMethylome	<i>Export a methylome</i>
-----------------	---------------------------

Description

Export a methylome as a TSV file.

Usage

```
exportMethylome(model, filename)
```

Arguments

model	A methimputeBinomialHMM object.
filename	The name of the file to be exported.

Value

NULL

Examples

```
## Not run:
## Get some toy data
file <- system.file("data", "arabidopsis_toydata.RData", package="methimpute")
data <- get(load(file))
print(data)
model <- callMethylation(data, max.iter=10)
exportMethylome(model, filename = tempfile())

## End(Not run)
```

 extractCytosinesFromFASTA

Extract cytosine coordinates

Description

Extract cytosine coordinates and context information from a FASTA file. Cytosines in ambiguous reference contexts are not reported.

Usage

```
extractCytosinesFromFASTA(file, contexts = c("CG", "CHG", "CHH"),
  anchor.C = NULL)
```

Arguments

file	A character with the file name.
contexts	The contexts that should be extracted. If the contexts are named, the returned object will use those names for the contexts.
anchor.C	A named vector with positions of the anchoring C in the contexts. This is necessary to distinguish contexts such as C*C*CG (anchor.C = 2) and *C*CCG (anchor.C = 1). Names must match the contexts. If unspecified, the first C within each context will be taken as anchor.

Value

A [GRanges-class](#) object with coordinates of extracted cytosines and meta-data column 'context'.

Examples

```
## Read a non-compressed FASTA files:
filepath <- system.file("extdata", "arabidopsis_sequence.fa.gz", package="methimpute")

## Only CG context
cytosines <- extractCytosinesFromFASTA(filepath, contexts = 'CG')
table(cytosines$context)

## Split CG context into subcontexts
cytosines <- extractCytosinesFromFASTA(filepath,
  contexts = c('DCG', 'CCG'),
  anchor.C = c(DCG=2, CCG=2))
table(cytosines$context)

## With contexts that differ only by anchor
cytosines <- extractCytosinesFromFASTA(filepath,
  contexts = c('DCG', 'CCG', 'CCG', 'CWG', 'CHH'),
  anchor.C = c(DCG=2, CCG=2, CCG=1, CWG=1, CHH=1))
table(cytosines$context)

## With named contexts
contexts <- c(CG='DCG', CG='CCG', CHG='CCG', CHG='CWG', CHH='CHH')
cytosines <- extractCytosinesFromFASTA(filepath,
```

```
contexts = contexts,  
anchor.C = c(DCG=2, CCG=2, CCG=1, CWG=1, CHH=1))  
table(cytosines$context)
```

getDistinctColors *Get distinct colors*

Description

Get a set of distinct colors selected from [colors](#).

Usage

```
getDistinctColors(n, start.color = "blue4", exclude.colors = c("white",  
"black", "gray", "grey", "\\<yellow\\>", "yellow1", "lemonchiffon"),  
exclude.brightness.above = 1, exclude.rgb.above = 210)
```

Arguments

n Number of colors to select. If *n* is a character vector, `length(n)` will be taken as the number of colors and the colors will be named by *n*.

start.color Color to start the selection process from.

exclude.colors Character vector with colors that should not be used.

exclude.brightness.above
 Exclude colors where the 'brightness' value in HSV space is above. This is useful to obtain a matt palette.

exclude.rgb.above
 Exclude colors where all RGB values are above. This is useful to exclude whitish colors.

Details

The function computes the euclidian distance between all [colors](#) and iteratively selects those that have the furthest closes distance to the set of already selected colors.

Value

A character vector with colors.

Author(s)

Aaron Taudt

Examples

```
cols <- getDistinctColors(5)  
pie(rep(1,5), labels=cols, col=cols)
```

getPosteriors	<i>Get original posteriors</i>
---------------	--------------------------------

Description

Transform the 'posteriorMeth', 'posteriorMax', and 'status' columns into original posteriors from the HMM.

Usage

```
getPosteriors(data)
```

Arguments

data The \$data entry from a [methimputeBinomialHMM](#) object.

Value

A matrix with posteriors.

getStateColors	<i>Get state colors</i>
----------------	-------------------------

Description

Get the colors that are used for plotting.

Usage

```
getStateColors(states = NULL)
```

Arguments

states A character vector.

Value

A character vector with colors.

See Also

[plotting](#)

Examples

```
cols <- getStateColors()
pie(1:length(cols), col=cols, labels=names(cols))
```

import	<i>Methimpute data import</i>
--------	-------------------------------

Description

This page provides an overview of all **methimpute** data import functions.

Usage

```
importBSMAP(file, chrom.lengths = NULL, skip = 1, contexts = c(CG =
  "NNCGN", CHG = "NNCHG", CHH = "NNCHH"))
```

```
importMethylpy(file, chrom.lengths = NULL, skip = 1, contexts = c(CG
  = "CGN", CHG = "CHG", CHH = "CHH"))
```

```
importBSSeeker(file, chrom.lengths = NULL, skip = 0)
```

```
importBismark(file, chrom.lengths = NULL, skip = 0)
```

Arguments

file	The file to import.
chrom.lengths	A data.frame with chromosome names in the first, and chromosome lengths in the second column. Only chromosomes named in here will be returned. Alternatively a tab-separated file with such a data.frame (with headers).
skip	The number of lines to skip. Usually 1 if the file contains a header and 0 otherwise.
contexts	A character vector of the contexts that are to be assigned. Since some programs report 5-letter contexts, this parameter can be used to obtain a reduced number of contexts. Will yield contexts CG, CHG, CHH by default. Set contexts=NULL to obtain all available contexts.

Value

A **methimputeData** object.

Functions

- `importBSMAP`: Import a BSMAP methylation extractor file.
- `importMethylpy`: Import a Methylpy methylation extractor file.
- `importBSSeeker`: Import a BSSeeker methylation extractor file.
- `importBismark`: Import a Bismark methylation extractor file.

Examples

```
## Get an example file in BSSeeker format
file <- system.file("extdata", "arabidopsis_bsseeker.txt.gz", package="methimpute")
data(arabidopsis_chromosomes)
bsseeker.data <- importBSSeeker(file, chrom.lengths=arabidopsis_chromosomes)
```

```
## Get an example file in Bismark format
file <- system.file("extdata", "arabidopsis_bismark.txt", package="methimpute")
data(arabidopsis_chromosomes)
arabidopsis_chromosomes$chromosome <- sub('chr', '', arabidopsis_chromosomes$chromosome)
bismark.data <- importBismark(file, chrom.lengths=arabidopsis_chromosomes)

## Get an example file in BSMAP format
file <- system.file("extdata", "arabidopsis_BSMAP.txt", package="methimpute")
data(arabidopsis_chromosomes)
bsmap.data <- importBSMAP(file, chrom.lengths=arabidopsis_chromosomes)

## Get an example file in Methylypy format
file <- system.file("extdata", "arabidopsis_methylypy.txt", package="methimpute")
data(arabidopsis_chromosomes)
arabidopsis_chromosomes$chromosome <- sub('chr', '', arabidopsis_chromosomes$chromosome)
methylypy.data <- importMethylypy(file, chrom.lengths=arabidopsis_chromosomes)
```

importRene

Import a Rene methylation extractor file

Description

Import a Rene methylation extractor file into a [GRanges-class](#) object.

Usage

```
importRene(file, chrom.lengths = NULL, skip = 1)
```

Arguments

file	The file to import.
chrom.lengths	A data.frame with chromosome names in the first, and chromosome lengths in the second column. Only chromosomes named in here will be returned. Alternatively a tab-separated file with such a data.frame (with headers).
skip	The number of lines to skip. Usually 1 if the file contains a header and 0 otherwise.

Value

A [methimputeData](#) object.

Examples

```
## Get an example file in Rene format
file <- system.file("extdata", "arabidopsis_rene.txt", package="methimpute")
data(arabidopsis_chromosomes)
rene.data <- methimpute::importRene(file, chrom.lengths=arabidopsis_chromosomes)
```

inflateMethylome	<i>Inflate an imported methylation extractor file</i>
------------------	---

Description

Inflate an imported methylation extractor file to contain all cytosine positions. This is useful to obtain a full methylome, including non-covered cytosines, because most methylation extractor programs only report covered cytosines.

Usage

```
inflateMethylome(methylome, methylome.full)
```

Arguments

`methylome` A [GRanges-class](#) with methylation counts.
`methylome.full` A [GRanges-class](#) with positions for all cytosines or a file with such an object.

Value

The `methylome.full` object with added metadata column 'counts'.

Examples

```
## Get an example file in BSSeeker format
file <- system.file("extdata", "arabidopsis_bsseeker.txt.gz", package="methimpute")
bsseeker.data <- importBSSeeker(file)
bsseeker.data

## Inflate to full methylome (including non-covered sites)
data(arabidopsis_toydata)
full.methylome <- inflateMethylome(bsseeker.data, arabidopsis_toydata)
full.methylome
```

loadFromFiles	<i>Load methimpute objects from file</i>
---------------	---

Description

Wrapper to load [methimpute](#) objects from file and check the class of the loaded objects.

Usage

```
loadFromFiles(files, check.class = c("GRanges", "methimputeBinomialHMM"))
```

Arguments

`files` A list of [GRanges-class](#) or [methimputeBinomialHMM](#) objects or a character vector with files that contain such objects.
`check.class` Any combination of `c('GRanges', 'methimputeBinomialHMM')`. If any of the loaded objects does not belong to the specified class, an error is thrown.

Value

A list of [GRanges-class](#) or [methimputeBinomialHMM](#) objects.

Examples

```
## Get some files that you want to load
file <- system.file("data", "arabidopsis_toydata.RData",
                    package="methimpute")

## Load and print
data <- loadFromFiles(file)
print(data)
```

methimpute-objects *methimpute objects*

Description

[methimpute](#) defines several objects.

- [methimputeData](#): Returned by [importBSSeeker](#), [importBismark](#) and [inflateMethylome](#).
- [methimputeBinomialHMM](#): Returned by [callMethylation](#).

methimputeBinomialHMM *methimputeBinomialHMM*

Description

The `methimputeBinomialHMM` is a `list()` which contains various entries (see Value section). The main entry of this object is `$data`, which contains the methylation status calls and posterior values. See Details for a description of all columns.

Details

The `$data` entry in this object contains the following columns:

- `context` The sequence context of the cytosine.
- `counts` Counts for methylated and total number of reads at each position.
- `distance` The distance in base-pairs from the previous to the current cytosine.
- `transitionContext` Transition context in the form "previous-current".
- `posteriorMax` Maximum posterior value of the methylation status call, can be interpreted as the confidence in the call.
- `posteriorMeth` Posterior value of the "methylated" component.
- `posteriorUnmeth` Posterior value of the "unmethylated" component.
- `status` Methylation status.
- `rc.meth.lvl` Recalibrated methylation level, calculated as $r\$data\$rc.meth.lvl = r\$data\$params\$emissionParams * r\$data\$posteriorUnmeth + r\$params\$emissionParams\$Methylated[data\$context,] * r\$data\$posteriorMeth$, where `r` is the `methimputeBinomialHMM` object.

Value

A list() with the following entries:

convergenceInfo	A list() with information about the convergence of the model fitting procedure.
params	A list() with fitted and non-fitted model parameters.
params.initial	A list() with initial values for the model parameters.
data	A GRanges-class with cytosine positions and methylation status calls.
segments	The data entry where coordinates of consecutive cytosines with the same methylation status have been merged.

See Also

[methimpute-objects](#)

methimputeData	<i>methimputeData</i>
----------------	-----------------------

Description

A [GRanges-class](#) object containing cytosine coordinates with meta-data columns 'context' and 'counts'.

See Also

[methimpute-objects](#)

parameterScan	<i>Perform a parameter scan</i>
---------------	---------------------------------

Description

Perform a parameter scan for an arbitrary parameter.

Usage

```
parameterScan(f, param, values, ...)
```

Arguments

f	A function for which to perform the scan.
param	A character with the parameter for which to perform the scan.
values	A vector with parameter values for which to perform the scan.
...	Other parameters passed through to f.

Value

A data.frame with loglikelihood values.

Description

This page provides an overview of all [methimpute](#) plotting functions.

Usage

```
plotHistogram(model, total.counts, binwidth = 1)

plotScatter(model, datapoints = 1000)

plotTransitionProbs(model)

plotConvergence(model)

plotEnrichment(model, annotation, windowsize = 100, insidewindows = 20,
  range = 1000, category.column = NULL, plot = TRUE,
  df.list = NULL)

plotPosteriorDistance(model, datapoints = 1e+06, binwidth = 5,
  max.coverage.y = 0, min.coverage.x = 3, xmax = 200,
  xbreaks.interval = xmax/10, cutoffs = NULL)
```

Arguments

<code>model</code>	A methimputeBinomialHMM object.
<code>total.counts</code>	The number of total counts for which the histogram is to be plotted.
<code>binwidth</code>	The bin width for the histogram/boxplot.
<code>datapoints</code>	The number of randomly selected datapoints for the plot.
<code>annotation</code>	A GRanges-class object with coordinates for the annotation.
<code>windowsize</code>	Resolution in base-pairs for the curve upstream and downstream of the annotation.
<code>insidewindows</code>	Number of data points for the curve inside the annotation.
<code>range</code>	Distance upstream and downstream for which the enrichment profile is calculated.
<code>category.column</code>	The name of a column in data that will be used for facetting of the plot.
<code>plot</code>	Logical indicating whether a plot or the underlying data.frame is to be returned.
<code>df.list</code>	A <code>list()</code> of data.frames, output from <code>plotEnrichment(..., plot=FALSE)</code> . If specified, option data will be ignored.
<code>max.coverage.y</code>	Maximum coverage for positions on the y-axis.
<code>min.coverage.x</code>	Minimum coverage for positions on the x-axis.
<code>xmax</code>	Upper limit for the x-axis.
<code>xbreaks.interval</code>	Interval for breaks on the x-axis.
<code>cutoffs</code>	A vector with values that are plotted as horizontal lines. The names of the vector must match the context levels in <code>data\$context</code> .

Value

A `ggplot` object.

Functions

- `plotHistogram`: Plot a histogram of count values and fitted distributions.
- `plotScatter`: Plot a scatter plot of read counts colored by methylation status.
- `plotTransitionProbs`: Plot a heatmap of transition probabilities.
- `plotConvergence`: Plot the convergence of the probability parameters.
- `plotEnrichment`: Plot an enrichment profile around an annotation.
- `plotPosteriorDistance`: Maximum posterior vs. distance to nearest covered cytosine.

Examples

```
## Get some toy data
file <- system.file("data", "arabidopsis_toydata.RData",
                    package="methimpute")
data <- get(load(file))
print(data)
model <- callMethylation(data)
## Make nice plots
plotHistogram(model, total.counts=5)
plotScatter(model)
plotTransitionProbs(model)
plotConvergence(model)
plotPosteriorDistance(model$data)

## Get annotation data and make an enrichment profile
# Note that this looks a bit ugly because our toy data
# has only 200000 datapoints.
data(arabidopsis_genes)
plotEnrichment(model, annotation=arabidopsis_genes)
```

```
print.methimputeBinomialHMM
      Print model object
```

Description

Print model object

Usage

```
## S3 method for class 'methimputeBinomialHMM'
print(x, ...)
```

Arguments

<code>x</code>	A <code>methimputeBinomialHMM</code> object.
<code>...</code>	Ignored.

Value

An invisible NULL.

transCoord	<i>Transform genomic coordinates</i>
------------	--------------------------------------

Description

Add two columns with transformed genomic coordinates to the [GRanges-class](#) object. This is useful for making genomewide plots.

Usage

```
transCoord(gr)
```

Arguments

gr A [GRanges-class](#) object.

Value

The input [GRanges-class](#) with two additional metadata columns 'start.genome' and 'end.genome'.

Index

arabidopsis_chromosomes, 3
arabidopsis_genes, 3
arabidopsis_TEs, 3
arabidopsis_toydata, 4

binCounts (binning), 4
binMethylome (binning), 4
binning, 4
binomialTestMethylation, 5
binPositions (binning), 4

callMethylation, 6, 10, 18
callMethylationSeparate, 7
collapseBins, 8
colors, 13

distanceCorrelation, 10, 10, 11

estimateTransDist, 6, 7, 10
exportMethylome, 11
extractCytosinesFromFASTA, 12

getDistinctColors, 13
getPosteriors, 14
getStateColors, 14
ggplot, 10, 11, 21

import, 15
importBismark, 18
importBismark (import), 15
importBSMAP (import), 15
importBSSeeker, 18
importBSSeeker (import), 15
importMethylpy (import), 15
importRene, 16
inflateMethylome, 17, 18

loadFromFiles, 17

methimpute, 3, 4, 15, 17, 18, 20
methimpute (methimpute-package), 2
methimpute-objects, 18
methimpute-package, 2
methimputeBinomialHMM, 6–8, 11, 14, 17, 18, 18, 20, 21

methimputeData, 4–7, 10, 15, 16, 18, 19

parameterScan, 19
plotConvergence (plotting), 20
plotEnrichment (plotting), 20
plotHistogram (plotting), 20
plotPosteriorDistance (plotting), 20
plotScatter (plotting), 20
plotting, 2, 14, 20
plotTransitionProbs (plotting), 20
print.methimputeBinomialHMM, 21

transCoord, 22