

Package ‘ReadqPCR’

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Title Read qPCR data

Description The package provides functions to read raw RT-qPCR data of different platforms.

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Depends R(>= 2.14.0), Biobase, methods

Suggests qpcR

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License LGPL-3

LazyLoad yes

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ReadqPCR-package *Read qPCR data*

Description

The package provides functions to read raw RT-qPCR data of different platforms.

Details

Package: ReadqPCR
Type: Package
Version: 1.5.3
Date: 2013-03-23
Depends: R(>= 2.14.0), Biobase, methods, affy
Imports: Biobase
Suggests: qpcR
License: LGPL-3
LazyLoad: yes

```
library(ReadqPCR)
```

Author(s)

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References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

Examples

```
## some examples are given in the vignette  
## Not run:  
library(ReadqPCR)  
vignette("ReadqPCR")  
  
## End(Not run)
```

CyclesSet-class	<i>Class CyclesSet</i>
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Description

Class to contain and describe raw fluorescence data. Extends eSet

Creating Objects

```
new("CyclesSet")
```

Slots

assayData: Object of class AssayData containing the raw data, which will be a matrix of fluorescence values.

phenoData: Object of class AnnotatedDataFrame containing phenotypic data for the samples.

annotation A character string identifying the annotation that may be used for the CyclesSet instance.

protocolData: Object of class AnnotatedDataFrame containing protocol data for the samples.

featureData Object of class AnnotatedDataFrame containing feature-level (e.g., probeset-level) information.

experimentData: Object of class "MIAME" containing experiment-level information.

.__classVersion__: Object of class Versions describing the R and Biobase version number used to create the instance. Intended for developer use.

Methods

exprs signature(object = "CyclesSet"): extracts the matrix with the fluorescence values.

exprs<- signature(object = "CyclesSet", value = "matrix"): replaces the matrix with the fluorescence values.

Author(s)

Nor Izayu Abdul Rahman, Matthias Kohl <Matthias.Kohl@stamats.de>

References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

See Also

[eSet](#)

merge

*Generic Function for Merging eSet and AnnotatedDataFrame***Description**

Generic function for the merging an eSet and an AnnotatedDataFrame; that is, the information given in the AnnotatedDataFrame is merged into the appropriate slot of the given eSet. This can be slot phenoData (default), featureData, or protocolData.

Usage

```
merge(x, y, ...)
## S4 method for signature 'eSet,AnnotatedDataFrame'
merge(x, y, eSet.slot = "phenoData",
      by = intersect(names(pData(x)), names(pData(y))),
      by.x = by, by.y = by,
      all = FALSE, all.x = all, all.y = all,
      sort = FALSE, suffixes = c(".x", ".y"),
      incomparables = NULL, ...)
## S4 method for signature 'AnnotatedDataFrame,eSet'
merge(x, y, eSet.slot = "phenoData",
      by = intersect(names(pData(x)), names(pData(y))),
      by.x = by, by.y = by,
      all = FALSE, all.x = all, all.y = all,
      sort = FALSE, suffixes = c(".x", ".y"),
      incomparables = NULL, ...)
```

Arguments

x	object of class eSet or AnnotatedDataFrame
y	object of class eSet or AnnotatedDataFrame
eSet.slot	name of the slot of the given eSet which is merged with the given AnnotatedDataFrame. This can be "phenoData" (default), "featureData", or "protocolData"
by	specifications of the columns used for merging.
by.x	specifications of the columns used for merging.
by.y	specifications of the columns used for merging.
all	logical; all = L is shorthand for all.x = L and all.y = L, where L is either TRUE or FALSE.
all.x	logical; if TRUE, then extra rows will be added to the output, one for each row in x that has no matching row in y. These rows will have NAs in those columns that are usually filled with values from y. The default is FALSE, so that only rows with data from both x and y are included in the output.
all.y	logical; analogous to all.x.
sort	logical. Should the result be sorted on the by columns?
suffixes	a character vector of length 2 specifying the suffixes to be used for making unique the names of columns in the result which not used for merging (appearing in by etc).
incomparables	values which cannot be matched. See match .
...	additional arguments to be passed to or from methods.

Details

For details on the arguments see [merge](#).

Value

Object of class "CyclesSet".

Methods

x = "eSet", y = "AnnotatedDataFrame": merge y into specified slot of x.

x = "AnnotatedDataFrame", y = "eSet": merge x into specified slot of y.

Author(s)

Nor Izayu Abdul Rahman, Matthias Kohl <Matthias.Kohl@stamats.de>

See Also

[merge](#), [eSet](#)

Examples

```
path <- system.file("exData", package = "ReadqPCR")

LC480.example <- file.path(path, "LC480_Example.txt")
## Read in the raw qPCR data from file "LC480_Example.txt"
## with maximum cycle to be read in the values is 45 (default).
cycData <- read.LC480(file = LC480.example)

LC480.SamInfo <- file.path(path, "LC480_Example_SampleInfo.txt")
# Read in the sample information data from file "LC480_Example_SampleInfo.txt".
samInfo <- read.LC480SampleInfo(LC480.SamInfo)

cycData1 <- merge(cycData, samInfo)
```

qpcR2CyclesSet

Transform a dataset from qpcR Package into CyclesSet

Description

Transform a dataset from package **qpcR** into an object of class "CyclesSet".

Usage

```
qpcR2CyclesSet(x, cyc = 1, cycleThreshold)
```

Arguments

x the name of the dataset from package **qpcR** to be transformed.
cyc the column number which contains cycle data.
cycleThreshold maximum number of cycles which the data will be counted and transformed.

Details

Allows the user to transform a dataset from package **qpcR** into an object of class "CyclesSet" class, alongside phenotypic data.

Value

Object of class "CyclesSet".

Author(s)

Nor Izayu Abdul Rahman, Matthias Kohl <Matthias.Kohl@tamats.de>

References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

See Also

[reps](#), [CyclesSet-class](#)

Examples

```
library(qpcR)

## Transform the reps dataset from qpcR Package
## with maximum cycle to be read in the value is 45.
repsdata <- qpcR2CyclesSet(reps, cyc=1, cycleThreshold=45)

## Transform the batsch1 dataset from qpcR Package
## with maximum cycle to be read in the value is 40.
batsch1data <- qpcR2CyclesSet(batsch1, cyc=1, cycleThreshold=40)
```

qPCRBatch-class

Class qPCRBatch

Description

Class to Contain and Describe raw and normalised qPCR Data, as Cq or delta-Cq values. Extends eSet

Creating Objects

```
new("qPCRBatch")
```

Slots

assayData: Object of class AssayData containing the raw data, which will be at minimum a matrix of Cq values. This slot can also hold a matrix of well.info values if these are present in the input file read in by read.qPCR or read.taqman

phenoData: Object of class AnnotatedDataFrame containing phenotypic data for the samples.

annotation A character string identifying the annotation that may be used for the qPCRBatch instance.

protocolData: Object of class AnnotatedDataFrame containing protocol data for the samples.

featureData Object of class AnnotatedDataFrame containing feature-level (e.g., probeset-level) information.

experimentData: Object of class "MIAME" containing experiment-level information.

.__classVersion__: Object of class Versions describing the R and Biobase version number used to create the instance. Intended for developer use.

Methods

exprs signature(object = "qPCRBatch"): extracts the Cq expression matrix.

exprs<- signature(object = "qPCRBatch", value = "matrix"): replaces the Cq expression matrix.

se.exprs signature(object = "qPCRBatch"): extracts the expression matrix with SDs of Cq values.

se.exprs<- signature(object = "qPCRBatch", value = "matrix"): replaces the expression matrix with SDs of Cq values.

exprs.well.order signature(object = "qPCRBatch"): extracts the Cq well order matrix (if it exists).

exprs.well.order<- signature(object = "qPCRBatch", value = "matrix"): replaces the Cq well order matrix.

effs signature(object = "qPCRBatch"): extracts the efficiency matrix (if it exists).

effs<- signature(object = "qPCRBatch", value = "matrix"): replaces the efficiency matrix.

se.effs signature(object = "qPCRBatch"): extracts the matrix with the standard errors/deviations of the efficiencies (if it exists).

se.effs<- signature(object = "qPCRBatch", value = "matrix"): replaces the matrix with the standard errors/deviations of the efficiencies.

Note

This class is better described in the vignette.

Author(s)

James Perkins <jimrperkins@gmail.com>

References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

See Also[eSet](#)

`read.LC480`*Read in raw qPCR data of Light Cycler 480*

Description

Reads in raw qPCR data of Light Cycler 480 and uses the data to populate an object of class "CyclesSet".

Usage

```
read.LC480(file, colNames = c("Sample position", "Sample name",
                             "Program number", "Segment number",
                             "Cycle number", "Acquisition time",
                             "Acquisition temperature",
                             "Fluorescence data"),
           cycleThreshold = 45, fileType = "txt", skip = 1,
           header = TRUE, sep = "\t", quote = "\"", dec = ".",
           fill = TRUE, comment.char = "")
```

Arguments

<code>file</code>	the name of the file to be read in.
<code>colNames</code>	a character vector of names to be assumed for the columns.
<code>cycleThreshold</code>	maximum number of cycles which will be read in.
<code>fileType</code>	the type of the file.
<code>skip</code>	integer: the number of lines of the data file to skip before beginning to read data.
<code>header</code>	a logical value indicating whether the file contains the names of the variables as its first line. If missing, the value is determined from the file format: header is set to TRUE if and only if the first row contains one fewer field than the number of columns.
<code>sep</code>	the field separator character. Values on each line of the file are separated by this character. If <code>sep = ""</code> (the default for read.table) the separator is 'white space', that is one or more spaces, tabs, newlines or carriage returns.
<code>quote</code>	the set of quoting characters. To disable quoting altogether, use <code>quote = ""</code> . See scan for the behaviour on quotes embedded in quotes. Quoting is only considered for columns read as character, which is all of them unless <code>colClasses</code> is specified.
<code>dec</code>	the character used in the file for decimal points.
<code>fill</code>	logical. If TRUE then in case the rows have unequal length, blank fields are implicitly added. See read.table .
<code>comment.char</code>	character: a character vector of length one containing a single character or an empty string. Use <code>""</code> to turn off the interpretation of comments altogether.

Details

Allows the user to read in qPCR fluorescence data from Light Cycler 480 which has been exported to a txt-file, alongside phenotypic data.

Value

Object of class "CyclesSet".

Author(s)

Nor Izayu Abdul Rahman, Matthias Kohl <Matthias.Kohl@stamats.de>

References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

See Also

[read.table](#)

[read.LC480SampleInfo](#) for reading in sample information of qPCR data from Light Cycler and produce an object of Class "CyclesSet".

[merge](#) for merging the phenodata from [read.LC480](#) and [read.LC480SampleInfo](#) and produce an object of Class "CyclesSet".

[read.qPCR](#) and [read.taqman](#) for reading in the RT-qPCR data consisting of Cq values.

[eSet](#), [CyclesSet-class](#)

Examples

```
path <- system.file("exData", package = "ReadqPCR")
LC480.example <- file.path(path, "LC480_Example.txt")

## Read in the raw qPCR data from file "LC480_Example.txt"
## with maximum cycle to be read in the values is 45 (default).
cycData <- read.LC480(file = LC480.example)

## Read in the data from file "LC480_Example.txt"
## with maximum cycle to be read in the values is 50.
rawdata <- read.LC480(file=LC480.example, cycleThreshold=50)
```

`read.LC480SampleInfo` *Read sample information data of a qPCR experiment from Light Cycler 480*

Description

Reads sample information data of a qPCR experiment from Light Cycler 480 which is in txt-file and uses the data to populate an object of Class "AnnotatedDataFrame".

Usage

```
read.LC480SampleInfo(file, removeEmptyCols = TRUE,
                    header = TRUE, sep = "\t", quote = "\"",
                    dec = ".", fill = TRUE, comment.char = "#",
                    skip = 0)
```

Arguments

<code>file</code>	the name of the file to be read in.
<code>removeEmptyCols</code>	a logical value which indicates whether the empty column(s) should be removed or not.
<code>header</code>	a logical value indicating whether the file contains the names of the variables as its first line. If missing, the value is determined from the file format: header is set to TRUE if and only if the first row contains one fewer field than the number of columns.
<code>sep</code>	the field separator character. Values on each line of the file are separated by this character. If <code>sep = ""</code> (the default for <code>read.table</code>) the separator is ‘white space’, that is one or more spaces, tabs, newlines or carriage returns.
<code>quote</code>	the set of quoting characters. To disable quoting altogether, use <code>quote = ""</code> . See <code>scan</code> for the behaviour on quotes embedded in quotes. Quoting is only considered for columns read as character, which is all of them unless <code>colClasses</code> is specified.
<code>dec</code>	the character used in the file for decimal points.
<code>fill</code>	logical. If TRUE then in case the rows have unequal length, blank fields are implicitly added. See <code>read.table</code> .
<code>comment.char</code>	character: a character vector of length one containing a single character or an empty string. Use <code>"#"</code> to turn off the interpretation of comments altogether.
<code>skip</code>	integer: the number of lines of the data file to skip before beginning to read data.

Details

Allows the user to read in sample information data of a qPCR experiment from Light Cycler 480 which is in txt-file.

Value

Object of class "AnnotatedDataFrame".

Author(s)

Nor Izayu Abdul Rahman, Matthias Kohl <Matthias.Kohl@stamats.de>

References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

See Also[read.table](#)[read.LC480](#) for reading in the qPCR data from Light Cycler and produce an object of Class "CyclesSet".[merge](#) for merging the phenodata from [read.LC480](#) and [read.LC480SampleInfo](#) and produce an object of Class "CyclesSet".[read.qPCR](#) and [read.taqman](#) for reading in the RT-qPCR data.[AnnotatedDataFrame-class](#)**Examples**

```

path <- system.file("exData", package = "ReadqPCR")
LC480.SamInfo <- file.path(path, "LC480_Example_SampleInfo.txt")

# Read in the sample information data from file "LC480_Example_SampleInfo.txt".
samInfo <- read.LC480SampleInfo(LC480.SamInfo)

```

read.qPCR

*Read user formatted qPCR data and produce a qPCRBatch***Description**

Reads RT-qPCR data in format specified in the [ReadqPCR](#) vignette and uses the data to populate an object of class "qPCRBatch".

Usage

```

read.qPCR(filename = character(0),
          phenoData = new("AnnotatedDataFrame"),
          notes = "",
          verbose = FALSE)

```

Arguments

filename	file name (must be formatted as shown in vignette).
phenoData	an AnnotatedDataFrame object, a character of length one, or a data.frame.
notes	notes.
verbose	verbosity flag. If true more messages are given to the user on the processing steps

Details

Permits the user to read in qPCR Cq value data in a predefined format (more details on this format in the [ReadqPCR](#) package vignette), alongside phenotypic data and further notes about the data. If `phenoData` is a data.frame, it is converted to an [AnnotatedDataFrame](#). If it is NULL then a default object of class [AnnotatedDataFrame](#) is created, whose `pData` is a data.frame with rownames being the names of the samples, and with one column `sample` with an integer index. More details on how technical replicates are handled in the [ReadqPCR](#) package vignette

Value

Object of class "qPCRBatch".

Author(s)

James Perkins <jimrperkins@gmail.com>

References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

See Also

ExpressionSet-class

Examples

```
path <- system.file("exData", package = "ReadqPCR")
qPCR.example <- file.path(path, "qPCR.example.txt")
qPCRBatch.qPCR <- read.qPCR(qPCR.example)
```

read.taqman

Read Taqman qPCR data and produce a qPCRBatch

Description

Reads Taqman RT-qPCR data and uses the data to populate an object of class "qPCRBatch".

Usage

```
read.taqman(...,
  filenames = character(0),
  phenoData = new("AnnotatedDataFrame"),
  notes = "",
  verbose = FALSE)
```

Arguments

...	file names separated by comma.
filenames	file names in a character vector.
phenoData	an AnnotatedDataFrame object, a character of length one, or a data.frame.
notes	notes.
verbose	verbosity flag. If true more messages are given to the user on the processing steps

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