

Package ‘BiRewire’

March 6, 2025

Version 3.38.0

Date 2022-02-10

Title High-performing routines for the randomization of a bipartite graph (or a binary event matrix), undirected and directed signed graph preserving degree distribution (or marginal totals)

Maintainer Andrea Gobbi <gobbi.andrea@mail.com>

Description Fast functions for bipartite network rewiring through N consecutive switching steps (See References) and for the computation of the minimal number of switching steps to be performed in order to maximise the dissimilarity with respect to the original network. Includes functions for the analysis of the introduced randomness across the switching steps and several other routines to analyse the resulting networks and their natural projections. Extension to undirected networks and directed signed networks is also provided. Starting from version 1.9.7 a more precise bound (especially for small network) has been implemented. Starting from version 2.2.0 the analysis routine is more complete and a visual monitoring of the underlying Markov Chain has been implemented. Starting from 3.6.0 the library can handle also matrices with NA (not for the directed signed graphs). Since version 3.27.1 it is possible to add a constraint for dsg generation: usually positive and negative arc between two nodes could be not accepted.

License GPL-3

Depends igraph, slam, Rtsne, Matrix

Suggests RUnit, BiocGenerics

Author Andrea Gobbi [aut], Francesco Iorio [aut], Giuseppe Jurman [cbt], Davide Albanese [cbt], Julio Saez-Rodriguez [cbt].

URL <http://www.ebi.ac.uk/~iorio/BiRewire>

biocViews Network

git_url <https://git.bioconductor.org/packages/BiRewire>

git_branch RELEASE_3_20

git_last_commit 6980802

git_last_commit_date 2024-10-29

Repository Bioconductor 3.20

Date/Publication 2025-03-06

Contents

BiRewire-package	2
birewire.analysis.bipartite	3
birewire.analysis.dsg	5
birewire.analysis.undirected	7
birewire.bipartite.from.incidence	9
birewire.build.dsg	10
birewire.induced.bipartite	11
birewire.load.dsg	12
birewire.rewire.bipartite	12
birewire.rewire.bipartite.and.projections	14
birewire.rewire.dsg	15
birewire.rewire.undirected	17
birewire.sampler.bipartite	19
birewire.sampler.dsg	20
birewire.sampler.undirected	21
birewire.similarity	23
birewire.similarity.dsg	24
birewire.slum.to.sparseMatrix	24
birewire.visual.monitoring.bipartite	25
birewire.visual.monitoring.dsg	27
birewire.visual.monitoring.undirected	28
BRCA_binary_matrix	30
test_dsg	31
Index	32

BiRewire-package	<i>The BiRewire package</i>
------------------	-----------------------------

Description

R package for computationally-efficient rewiring of bipartite graphs (or randomisation of 0-1 tables with prescribed marginal totals), undirected and directed signed graphs (dsg). The package provides useful functions for the analysis and the randomisation of large biological datasets that can be encoded as 0-1 tables, hence modeled as bipartite graphs by considering a 0-1 table as an incidence matrix, and for data that can be encoded as directed signed graphs such as pathways and signaling networks. Large collections of such randomised tables can be used to approximate null models, preserving event-rates both across rows and columns, for statistical significance tests of combinatorial properties of the original dataset. The package provides an interface to a sampler routine useful for generating correctly such collections. Moreover a visual monitoring for the Markov Chain underlying the swithicng algorithm has been implemented. Since version 3.6.0 the SA can be performed also using matrices with NAs. In this case the positions of the NAs are preserved as the degree distribution. This extension is limited when the tables are provided instead of the graphs and does not work for the dsg.

Details

Summary:

Package:	BiRewire
Version:	3.7.0
Date:	2017-02-27
Require:	slam, igraph, Rtsne, Matrix, R>=2.10
URL:	http://www.ebi.ac.uk/~iorio/BiRewire
License:	GPL-3

Author(s)

Andrea Gobbi [aut], Davide Albanese [cvt], Francesco Iorio [cvt], Giuseppe Jurman [cvt].
 Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

References

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* *Bioinformatics* 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* *Bioinformatics* 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* *Bioinformatics* 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Jaccard, P. (1901), *Étude comparative de la distribution florale dans une portion des Alpes et des Jura*, *Bulletin de la Société Vaudoise des Sciences Naturelles* 37: 547–579.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028 Csardi, G. and Nepusz, T (2006)

Van der Maaten, L.J.P. and Hinton, G.E., *Visualizing High-Dimensional Data Using t-SNE*. *Journal of Machine Learning Research* 9(Nov):2579-2605, 2008 *The igraph software package for complex network research*, *InterJournal, Complex Systems* <http://igraph.sf.net>

birewire.analysis.bipartite

Analysis of Jaccard similarity trends across switching steps.

Description

This function performs a sequence of *max.iter* switching steps on the input bipartite graph *g* and compute the Jaccard similarity between *g* (the initial network) and its rewired version each *step* switching steps. This procedure is performed *n.networks* times and a simple explorative plot, with mean and CI, is visualized if *display* is set to true.

Usage

```
birewire.analysis.bipartite(incidence, step=10, max.iter="n", accuracy=0.00005,
verbose=TRUE, MAXITER_MUL=10, exact=FALSE, n.networks=50, display=TRUE)
```

Arguments

incidence	Incidence matrix of the initial bipartite graph g (can be extracted from an <code>igraph</code> bipartite graph using the <code>get.incidence</code> function). Since 3.6.0 this matrix can contain also NAs and the position of such entries will be preserved by the SA;
step	10 (default): the interval (in terms of switching steps) at which the Jaccard index between g and the its current rewired version is computed;
max.iter	"n" (default) the number of switching steps to be performed (or if <code>exact==TRUE</code> the number of successful switching steps). If equal to "n" then this number is considered equal to the analytically derived lower bound presented in <i>Gobbi et al.</i> (see References): $N = e/2(1 - d) \ln((e - de)/\delta)$ if <code>exact</code> is <code>FALSE</code> , $N = e(1 - d)/2 \ln((e - de)/\delta)$ otherwise, where e is the number of edges of g and d its edge density. This bound is much lower than the empirical one proposed in <i>Milo et al. 2003</i> (see References);
accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
verbose	TRUE (default). When TRUE a progression bar is printed during computation;
MAXITER_MUL	10 (default). If <code>exact==TRUE</code> in order to prevent a possible infinite loop the program stops anyway after <code>MAXITER_MUL*max.iter</code> iterations;
exact	FALSE (default). If TRUE the program performs <code>max.iter</code> swithcing steps, otherwise the program will count also the not-performed swithcing steps;
n.networks	50 (default), the number of independent rewiring process starting from the same inital graph from which the mean value and the CI is computed.
display	TRUE (default). If TRUE two explorative plots are displayed summarizing the trend of the Jaccard index in terms of mean and confidence interval.

Details

This function performs `max.iter` switching steps (see references). In particular, at each step two edges are randomly selected from the current version of g . Let these two edges be (a, b) and (c, d) (where a and c belong to the first class of nodes whereas b and d belong to the second one), with $a \neq c$ and $b \neq d$.

If the (a, d) and (c, b) edges are not already present in the current current version of g then (a, d) and (c, b) replace (a, b) and (c, d) .

At each `step` number of switching steps the function computes the **Jaccard index** between the original graph g and its current version.

This procedure is perfomed `n.networks` times and if `display` is set to `TRUE`, two explorative plots showing the mean value of the Jaccad Index over the SS and its CI are displayed.

Value

A list containing a data.frame `data` collecting all the Jacard index computed (each row is a run of the SA), and the analytically derived lower bound N of switching steps to be performed by the switching algorithm in order to provide the revired version of g with the maximal level of achievable randomness (in terms of dissimilarity from the initial g).

Author(s)

Andrea Gobbi
 Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>
 Special thanks to:
 Davide Albanese

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Jaccard, P. (1901), *Étude comparative de la distribution florale dans une portion des Alpes et des Jura*, Bulletin de la Société Vaudoise des Sciences Naturelles 37: 547–579.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

Examples

```
library(BiRewire)
g <-graph.bipartite(rep(0:1,length=10), c(1:10))

##get the incidence matrix of g
m<-as.matrix(get.incidence(graph=g))

## set parameters
step=1
max=100*length(E(g))

## perform two different analysis using two different maximal number of switching steps
scores<-birewire.analysis.bipartite(m,step,max,n.networks=10)
scores2<-birewire.analysis.bipartite(m,step,"n",n.networks=10)
```

birewire.analysis.dsg *Analysis of Jaccard similarity trends across switching steps.*

Description

This function performs a sequence of *max.iter.pos* (and *max.iter.neg*) switching steps on the positive (and negative) part of the input dsg *g* and computes the Jaccard similarity between *g* (the initial network) and its rewired version each *step* switching steps. This procedure is performed *n.networks* times and a simple explorative plot, with mean and CI, is visualized if *display* is set to *TRUE*. The plot shows the trend of the Jaccard Index relative to the positive (and negative) part of *g*.

Usage

```
birewire.analysis.dsg(dsg, step=10, max.iter.pos='n',max.iter.neg='n',accuracy=0.00005,
  verbose=TRUE,MAXITER_MUL=10,exact=FALSE,n.networks=50,display=TRUE)
```

Arguments

dsg	The initial dsg object (see birewire.induced.bipartite). Note that the dsg must contain a list of two incidence matrices and not igraph bipartite graphs.
step	10 (default): the interval (in terms of switching steps) at which the Jaccard index between g and the its current rewired version is computed;
max.iter.pos	"n" (default) the number of switching steps to be performed (or if <code>exact==TRUE</code> the number of successful switching steps) for the positive part of g . See birewire.rewire.bipartite for more details;
max.iter.neg	"n" (default) the same of <code>max.iter.p</code> but relative to the negative part;
accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
verbose	TRUE (default). When TRUE a progression bar is printed during computation;
MAXITER_MUL	10 (default). If <code>exact==TRUE</code> in order to prevent a possible infinite loop the program stops anyway after <code>MAXITER_MUL*max.iter</code> iterations;
exact	FALSE (default). If TRUE the program performs <code>max.iter</code> swithcing steps, otherwise the program will count also the not-performed swithcing steps;
n.networks	50 (default), the number of independent rewiring process starting from the same initial graph from which the mean value and the CI is computed.
display	TRUE (default). If TRUE two explorative plots are displayed summarizing the trend of the Jaccard index in terms of mean and confidence interval.

Details

This procedure acts in the same way of [birewire.analysis.bipartite](#) but in the case of dsg. The similarity is measurwe using [birewire.similarity.dsg](#).

Value

A list containing two lists: *data* that is a list collecting all the Jacard index computed (each row is a run of the SA) for the positive and negative part, and a list with the analytically derived lower bounds N for the positive and negative part of g .

Author(s)

Andrea Gobbi
 Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

References

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T.and Jurman, G.and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Jaccard, P. (1901), *Étude comparative de la distribution florale dans une portion des Alpes et des Jura*, Bulletin de la Société Vaudoise des Sciences Naturelles 37: 547–579.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

Examples

```
library(BiRewire)
data(test_dsg)
dsg <- birewire.induced.bipartite(test_dsg, sparse=FALSE)

a=birewire.analysis.dsg(dsg, verbose=FALSE, step=1, exact=TRUE, max.iter.pos=200, max.iter.neg=50)
```

birewire.analysis.undirected

Analysis of Jaccard similarity trends across switching steps.

Description

This function performs a sequence of *max.iter* switching steps on the input undirected graph *g* and compute the Jaccard similarity between *g* (the initial network) and its rewired version each *step* switching steps. This procedure is performed *n.networks* times and a simple explorative plot, with mean and CI, is visualized if *display* is set to *TRUE*.

Usage

```
birewire.analysis.undirected(adjacency, step=10, max.iter="n", accuracy=0.00005,
verbose=TRUE, MAXITER_MUL=10, exact=FALSE, n.networks=50, display=TRUE)
```

Arguments

adjacency	Incidence matrix of the initial bipartite graph <i>g</i> (can be extracted from an igraph undirected graph using the get.adjacency function). Since 3.6.0 this matrix can contain also NAs and the position of such entries will be preserved by the SA;
step	10 (default): the interval (in terms of switching steps) at which the Jaccard index between <i>g</i> and the its current rewired version is computed;
max.iter	"n" (default) the number of switching steps to be performed (or if <i>exact==TRUE</i> the number of successful switching steps). If equal to "n" then this number is considered equal to the analytically derived lower bound presented in <i>Gobbi et al.</i> (see References): $N = e/(2d^3 - dd^2 + 2d + 2) \ln((e - de)/\delta)$ if <i>exact</i> is FALSE, $N = e(1 - d)/2 \ln((e - de)/\delta)$ otherwise, where <i>e</i> is the number of edges of <i>g</i> and <i>d</i> its edge density. This bound is much lower than the empirical one proposed in <i>Milo et al. 2003</i> (see References);

accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
verbose	TRUE (default). When TRUE a progression bar is printed during computation;
MAXITER_MUL	10 (default). If <i>exact</i> ==TRUE in order to prevent a possible infinite loop the program stops anyway after MAXITER_MUL*max.iter iterations;
exact	FALSE (default). If TRUE the program performs <i>max.iter</i> switching steps, otherwise the program will count also the not-performed switching steps;
n.networks	50 (default), the number of independent rewiring process starting from the same initial graph from which the mean value and the CI is computed.
display	TRUE (default). If TRUE two explorative plots are displayed summarizing the trend of the Jaccard index in terms of mean and confidence interval.

Details

This function performs *max.iter* switching steps (see references). In particular, at each step two edges are randomly selected from the current version of *g*. Let these two edges be (a, b) and (c, d) , with $a \neq c, b \neq d, a \neq d, b \neq c$.

If the (a, d) and (c, b) (or (a, d) and (b, d)) edges are not already present in the current version of *g* then (a, d) and (c, b) replace (a, b) and (c, d) (or (a, b) and (c, d) replace (a, c) and (b, d)). If both of the configurations are allowed, then one of them is randomly selected.

At each *step* number of switching steps the function computes the **Jaccard index** between the original graph *g* and its current version.

This procedure is performed *n.networks* times and if *display* is set to TRUE, two explorative plots showing the mean value of the Jaccard Index over the SS and its CI are displayed.

Value

A list containing a data.frame *data* collecting all the Jaccard index computed (each row is a run of the SA), and the analytically derived lower bound *N* of switching steps to be performed by the switching algorithm in order to provide the required version of *g* with the maximal level of achievable randomness (in terms of dissimilarity from the initial *g*).

Author(s)

Andrea Gobbi
 Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>
 Special thanks to:
 Davide Albanese

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Gobbi, A. and Jurman, G. (2013) *Theoretical and algorithmic solutions for null models in network theory* (Doctoral dissertation) <http://eprints-phd.biblio.unitn.it/1125/>

Jaccard, P. (1901), *Étude comparative de la distribution florale dans une portion des Alpes et des Jura*, Bulletin de la Société Vaudoise des Sciences Naturelles 37: 547–579.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

Examples

```
library(BiRewire)
g <- erdos.renyi.game(1000,0.1)
##get the incidence matrix of g
m<-as.matrix(get.adjacency(graph=g,sparse=FALSE))

## set parameters
step=1000
max=100*length(E(g))

## perform two different analysis using two different numbers of switching steps
scores<-birewire.analysis.undirected(m,step,max,n.networks=10,verbose=FALSE)
scores2<-birewire.analysis.undirected(m,step,"n",n.networks=10,verbose=FALSE)
```

```
birewire.bipartite.from.incidence
```

Converts an incidence matrix into a bipartite graph.

Description

This function creates an [igraph](#) bipartite graph from an incidence matrix.

Usage

```
birewire.bipartite.from.incidence(matrix,directed=FALSE)
```

Arguments

matrix	incidence matrix: an (n-by-m) binary matrix where rows correspond to vertices in the first class while columns correspond to vertices in the second one;
directed	Logical, if TRUE a directed graph is created.

Details

The function calls [graph.incidence](#) of package [igraph](#). See [igraph](#) documentation for more details.

Value

Bipartite *igraph* graph.

Author(s)

Andrea Gobbi
 Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

References

Csardi, G. and Nepusz, T (2006) *The igraph software package for complex network research*, Inter-Journal, Complex Systems url <http://igraph.sf.net>

Examples

```
library(igraph)
library(BiRewire)
g <- graph.bipartite( rep(0:1,length=10), c(1:10))

##gets the incidence matrix of g
m<-as.matrix(get.incidence(graph=g))

##rewire the current graph
m2=biwire.rewire.bipartite(m,100)

#create the rewired bipartite graph
g2<-biwire.bipartite.from.incidence(m2,TRUE)
```

biwire.build.dsg *Transform a dsg object in a SIF file.*

Description

The routine transforms the initial dsg (two bipartite graphs) into SIF dsg format.

Usage

```
biwire.build.dsg(dsg,delimiters=list(negative='-',positive='+'))
```

Arguments

dsg	The dsg to be converted;
delimiters	list(negative='-',positive='+') (default):a list with 'positive' and 'negative' names identifying the character encoding the relation;

Details

This function converts the dsg object into a SIF format that can be saved using `biwire.write.dsg`, an internal function, using the given delimiters for encoding the relations. It is the inverse function of [biwire.induced.bipartite](#).

Value

A dsg in SIF format.

Examples

```
data(test_dsg)
dsg=birewire.induced.bipartite(test_dsg)
tmp= birewire.rewire.dsg(dsg,verbose=FALSE)
dsg2=birewire.build.dsg(tmp)
```

```
birewire.induced.bipartite
```

Transform a SIF data frame into a dsg object (a list of positive and negative incidence matrix).

Description

The routine transforms the initial dsg graph in SIF format into a dsg object made of two bipartite graphs: one for positive edges and the other for negative edges.

Usage

```
birewire.induced.bipartite(g,delimiters=list(negative='-',positive='+'),sparse=FALSE)
```

Arguments

<code>g</code>	A dataframe in SIF format describing a dsg (for example the output of <code>birewire.load.dsg</code> ;
<code>delimiters</code>	<code>list(negative='-',positive='+')</code> (default):a list with 'positive' and 'negative' names identifying the character encoding the relation;
<code>sparse</code>	FALSE (default): if TRUE the two bipartite graphs are saved as <code>igraph</code> bipartite graphs ;

Details

This fuction extract the positive and negative part of `g` and create a dsg object that can be used for example in the rewiring algorithm. Is is the inverse function of `birewire.build.dsg`.

Value

A list of two incidence matrix or bipartite `igraph` objects.

References

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T.and Jurman, G.and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Examples

```
data(test_dsg)
dsg=birewire.induced.bipartite(test_dsg)
```

birewire.load.dsg *Read a SIF file from a given path*

Description

The routine reads a SIF file and return a R table.

Usage

```
birewire.load.dsg(path)
```

Arguments

path Path to the SIF file.

Value

A R table that can be transformred into a dsg using [birewire.induced.bipartite](#)

birewire.rewire.bipartite
Efficient rewiring of bipartite graphs

Description

Optimal implementation of the switching algorithm. It returns the rewired version of the initial bipartite graph or its incidence matrix.

Usage

```
birewire.rewire.bipartite(incidence, max.iter="n", accuracy=0.00005, verbose=TRUE,
MAXITER_MUL=10, exact=FALSE)
```

Arguments

incidence Incidence matrix of the initial bipartite graph g (can be extracted from an [igraph](#) bipartite graph using the [get.incidence](#)) function; or the entire bipartite [igraph](#) graph. Since 3.6.0, in the case the matrix is provided, such matrix can contain also NAs and the position of such entries will be preserved by the SA

max.iter "n" (default) the number of switching steps to be performed (or if *exact=TRUE* the number of **successful** switching steps). If equal to "n" then this number is considered equal to the analytically derived lower bound presented in *Gobbi et al.* (see References): $N = e/2(1 - d) \ln((e - de)/\delta)$ if *exact* is FALSE, $N = e(1 - d)/2 \ln((e - de)/\delta)$ otherwise , where e is the number of edges of g and d its edge density . This bound is much lower than the empirical one proposed in *Milo et al. 2003* (see References);

accuracy 0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fracion of common edges;

verbose	TRUE (default). When TRUE a progression bar is printed during computation.
MAXITER_MUL	10 (default). If <i>exact==TRUE</i> in order to prevent a possible infinite loop the program stops anyway after <i>MAXITER_MUL*max.iter</i> iterations;
exact	FALSE (default). If TRUE the program performs <i>max.iter</i> swithcing steps, otherwise the program will count also the not-performed swithcing steps;

Details

Main function of the package. It performs at most *max.iter* switching steps producing a rewired version of an initial bipartite graph.

Value

Incidence matrix of the rewired graphn or the *igraph* corresponding object depending on the input type.

Author(s)

Andrea Gobbi
Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

Examples

```
library(igraph)
library(BiRewire)
g <-graph.bipartite( rep(0:1,length=10), c(1:10))

##gets the incidence matrix of g
m<-as.matrix(get.incidence(graph=g))

##rewiring
m2=birewire.rewire.bipartite(m,100*length(E(g)))
##creates the corresponding bipartite graph
g2<-birewire.bipartite.from.incidence(m2,directed=TRUE)
```

```
birewire.rewire.bipartite.and.projections
```

Analysis and rewiring function processing a bipartite graphs and its two projections

Description

This function performs the same analysis of [birewire.analysis.bipartite](#) but additionally it provides in output a rewired version of the two networks resulting from the natural projections of the initial graph, together with the corresponding Jaccard index trends.

Usage

```
birewire.rewire.bipartite.and.projections(graph, step=10, max.iter="n",
accuracy=0.00005, verbose=TRUE, MAXITER_MUL=10)
```

Arguments

graph	A bipartite graph g ;
max.iter	"n" (default) the number of successful switching steps to be performed. If equal to "n" then this number is considered equal to the analytically derived lower bound $N = e(1-d)/2 \ln((e-de)/\delta)$ presented in <i>Gobbi et al.</i> (see References);
step	10 (default): the interval (in terms of switching steps) at which the Jaccard index between g and the its current rewired version is computed;
accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
verbose	TRUE (default) boolean value. If TRUE print a processing bar during the rewiring algorithm.
MAXITER_MUL	10 (default). Since N indicates the number of successful switching steps, in order to prevent a possible infinite loop the program stops anyway after $MAXITER_MUL * max.iter$ iterations ;

Details

See [birewire.analysis.bipartite](#) for details.

Value

A list containing the three sequences of Jaccard index values (`similarity_scores`, `similarity_scores.proj1`, `similarity_scores.proj2`) for the three resulting graphs respectively (rewired, `rewired.proj1`, `rewired.proj2`). The first one is the rewired version of the initial graph g , while the second and the third one are rewired versions of its natural projections.

Author(s)

Andrea Gobbi
Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* *Bioinformatics* 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* *Bioinformatics* 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Examples

```
library(igraph)
library(BiRewire)
g <- simplify(graph.bipartite( rep(0:1,length=100),
c(c(1:100),seq(1,100,3),seq(1,100,7),100,seq(1,100,13),
seq(1,100,17),seq(1,100,19),seq(1,100,23),100)))
##gets the incidence matrix of g
m<-as.matrix(get.incidence(graph=g))
## rewires g and its projections
result=birewire.rewire.bipartite.and.projections(g,step=10,max.iter="n",accuracy=0.00005)
```

birewire.rewire.dsg *Efficient rewiring of directed signed graphs*

Description

Optimal implementation of the switching algorithm. It returns the rewired version of the initial directed signed graph (dsg).

Usage

```
birewire.rewire.dsg(dsg,exact=FALSE,verbose=1,max.iter.pos='n',max.iter.neg='n',
accuracy=0.00005,MAXITER_MUL=10,path=NULL,delimitators=list(positive='+',negative='-'),check_p
```

Arguments

dsg	A dsg object: is a list of two incidence matrices (see References), "positive" and "negative", encoding the positive edges and negative edges. This list can be obtained reading a SIF file using <code>birewire.load.dsg</code> function and converting the resulting dataframe using <code>birewire.induced.bipartite</code> ;
exact	FALSE (default). If TRUE the program performs <i>max.iter</i> successful switching steps, otherwise the program will count also the not-performed switching steps;
verbose	TRUE (default). When TRUE a progression bar is printed during computation;

max.iter.pos	"n" (default) the number of switching steps to be performed on the positive part of <i>dsg</i> (or if <i>exact==TRUE</i> the number of successful switching steps). If equal to "n" then this number is considered equal to the analytically derived lower bound presented in <i>Gobbi et al.</i> (see References): $N = e/2(1 - d) \ln((e - de)/\delta)$ if <i>exact</i> is FALSE, $N = e(1 - d)/2 \ln((e - de)/\delta)$ otherwise, where <i>e</i> is the number of edges of <i>g</i> and <i>d</i> its edge density. This bound is much lower than the empirical one proposed in <i>Milo et al. 2003</i> (see References);
max.iter.neg	"n" (default) the number of switching steps to be performed on the negative part of <i>dsg</i> (or if <i>exact==TRUE</i> the number of successful switching steps). If equal to "n" then this number is considered equal to the analytically derived lower bound presented in <i>Gobbi et al.</i> (see References): $N = e/2(1 - d) \ln((e - de)/\delta)$ if <i>exact</i> is FALSE, $N = e(1 - d)/2 \ln((e - de)/\delta)$ otherwise, where <i>e</i> is the number of edges of <i>g</i> and <i>d</i> its edge density. This bound is much lower than the empirical one proposed in <i>Milo et al. 2003</i> (see References);
accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
MAXITER_MUL	10 (default). If <i>exact==TRUE</i> in order to prevent a possible infinite loop the program stops anyway after MAXITER_MUL*max.iter iterations;
path	NULL (default). If not NULL, the <i>dsg</i> is saved in <i>path</i> in SIF format;
delimiters	list(positive='+',negative=' -') (default). If <i>save.file</i> is true, the <i>dsg</i> is saved using <i>delimiters</i> as characters encoding the relations. See birewire.build.dsg for more details.
check_pos_neg	FALSE (default). if true check if there are positive and negative loops in the same arc, if so the routine will not save the <i>dsg</i> but return it anyway with a warning
in_sampler	FALSE (default).if TRUE does not print the warning due to the positive-negative check

Details

This function runs [birewire.rewire.bipartite](#) on the positive and negative part of *dsg*. See references for more details.

Value

Rewired *dsg*.

Author(s)

Andrea Gobbi: <gobbi.andrea@mail.com>

References

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast*

randomization of large genomic datasets while preserving alteration counts *Bioinformatics* 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

Examples

```
library(BiRewire)
data(test_dsg)
dsg=birewire.induced.bipartite(test_dsg)
tmp= birewire.rewire.dsg(dsg,verbose=FALSE)
```

birewire.rewire.undirected

Efficient rewiring of undirected graphs

Description

Optimal implementation of the switching algorithm. It returns the rewired version of the initial undirected graph or its adjacency matrix.

Usage

```
birewire.rewire.undirected(adjacency, max.iter="n", accuracy=0.00005,
verbose=TRUE, MAXITER_MUL=10, exact=FALSE)
```

Arguments

adjacency	An igraph undirected graph g or its adjacency matrix (can be extracted from g using get.adjacency). Since 3.6.0, if the matrix is provided, such matrix can contain also NAs and the position of such entries will be preserved by the SA
max.iter	"n" (default) the number of switching steps to be performed (or if <code>exact==TRUE</code> the number of successful switching steps). If equal to "n" then this number is considered equal to the analytically derived lower bound presented in <i>Gobbi et al.</i> (see References): $N = e/(2d^3 - 6d^2 + 2d + 2) \ln(e - de)$ if exact is FALSE, $N = e(1 - d)/2 \ln((e - de)/\delta)$ otherwise, where e is the number of edges of g and d its edge density. This bound is much lower than the empirical one proposed in <i>Milo et al. 2003</i> (see References);
accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
verbose	TRUE (default) boolean value. If TRUE print a processing bar during the rewiring algorithm.
MAXITER_MUL	10 (default). If <code>exact==TRUE</code> in order to prevent a possible infinite loop the program stops anyway after <code>MAXITER_MUL*max.iter</code> iterations;
exact	FALSE (default). If TRUE the program performs <code>max.iter</code> switching steps, otherwise the program will count also the not-performed switching steps;

Details

Performs at most *max.iter* number of rewiring steps producing a rewired version of an initial undirected graph.

Value

Adjacency matrix of the rewired graph or the relative *igraph* object depending on the input type.

Author(s)

Andrea Gobbi
 Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>
 Special thanks to: Davide Albanese

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Gobbi, A. and Jurman, G. (2013) *Theoretical and algorithmic solutions for null models in network theory* (Doctoral dissertation) <http://eprints-phd.biblio.unitn.it/1125/>
 R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

Examples

```
library(igraph)
library(BiRewire)
g <- erdos.renyi.game(1000,0.1)
##gets the incidence matrix of g
m<-as.matrix(get.adjacency(graph=g, sparse=FALSE))

## sets parameters
step=1000
max=100*length(E(g))

##rewiring
m2=birewire.rewire.undirected(m,100*length(E(g)))
##creates the corresponding bipartite graph
g2<-graph.adjacency(m2,mode="undirected")
```

 birewire.sampler.bipartite

Efficient generation of a null model for a given bipartite graph

Description

The routine samples correctly from the null model of a given bipartite graph creating a set of randomized version of the initial bipartite graph.

Usage

```
birewire.sampler.bipartite(incidence,K,path,max.iter="n", accuracy=0.00005,
  verbose=TRUE,MAXITER_MUL=10,exact=FALSE,write.sparse=TRUE)
```

Arguments

incidence	Incidence matrix of the initial bipartite graph. Since 3.6.0 this matrix can contain also NAs and the position of such entries will be preserved by the SA;
K	The number of networks that has to be generated;
path	The directory in which the routine stores the outputs;
max.iter	"n" (default) the number of switching steps to be performed (or if <i>exact==TRUE</i> the number of successful switching steps). If equal to "n" then this number is considered equal to the analytically derived lower bound presented in <i>Gobbi et al.</i> (see References): $N = e/2(1 - d) \ln((e - de)/\delta)$ if exact is FALSE, $N = e(1 - d)/2 \ln((e - de)/\delta)$ otherwise , where e is the number of edges of g and d its edge density . This bound is much lower than the empirical one proposed in <i>Milo et al. 2003</i> (see References);
accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
verbose	TRUE (default). When TRUE a progression bar is printed during computation.
MAXITER_MUL	10 (default). If <i>exact==TRUE</i> in order to prevent a possible infinite loop the program stops anyway after MAXITER_MUL*max.iter iterations;
exact	FALSE (default). If TRUE the program performs <i>max.iter</i> swithcing steps, otherwise the program will count also the not-performed swithcing steps;
write.sparse	TRUE (default). If FALSE the table is written as an R data.frame (long time and more space needed)

Details

The routine creates, starting from the given path, different subfolders in order to have maximum 1000 files for folder . Moreover the incidence matrices are saved using [write_stm_CLUTO](#) (sparse matrices) that can be loaded using [read_stm_CLUTO](#). The set is generated calling `birewire.rewire.bipartite` on the last generated graph starting from the input graph.

Author(s)

Andrea Gobbi: <gobbi.andrea@mail.com>

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

birewire.sampler.dsg *Efficient generation of a null model for a given dsg.*

Description

Efficient generation of a null model for a given dsg. The routine samples correctly from the null model of a given dsg creating a set of randomized dsgs.

Usage

```
birewire.sampler.dsg(dsg,K,path,delimiters=list(negative='-',positive='+'),exact=FALSE,
  verbose=TRUE, max.iter.pos='n',max.iter.neg='n', accuracy=0.00005,MAXITER_MUL=10,check_pos_neg=
```

Arguments

dsg	A dsg object: is a list of two incidence matrices (see References), "positive" and "negative", encoding the positive edges and negative edges. This list can be obtained reading a SIF file using <code>birewire.load.dsg</code> function and converting the resulting dataframe using <code>birewire.induced.bipartite</code> .
max.iter.pos	"n" (default) the number of switching steps to be performed on the positive part of <i>dsg</i> (or if <i>exact==TRUE</i> the number of successful switching steps). If equal to "n" then this number is considered equal to the analytically derived lower bound presented in <i>Gobbi et al.</i> (see References): $N = e/2(1 - d) \ln((e - de)/\delta)$ if <i>exact</i> is FALSE, $N = e(1 - d)/2 \ln((e - de)/\delta)$ otherwise, where <i>e</i> is the number of edges of <i>g</i> and <i>d</i> its edge density. This bound is much lower than the empirical one proposed in <i>Milo et al. 2003</i> (see References);
max.iter.neg	"n" (default) the number of switching steps to be performed on the negative part of <i>dsg</i> (or if <i>exact==TRUE</i> the number of successful switching steps). If equal to "n" then this number is considered equal to the analytically derived lower bound presented in <i>Gobbi et al.</i> (see References): $N = e/2(1 - d) \ln((e - de)/\delta)$ if <i>exact</i> is FALSE, $N = e(1 - d)/2 \ln((e - de)/\delta)$ otherwise, where <i>e</i> is the number of edges of <i>g</i> and <i>d</i> its edge density. This bound is much lower than the empirical one proposed in <i>Milo et al. 2003</i> (see References);
accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;

verbose	TRUE (default). When TRUE a progression bar is printed during computation.
MAXITER_MUL	10 (default). If <i>exact</i> == <i>TRUE</i> in order to prevent a possible infinite loop the program stops anyway after MAXITER_MUL*max.iter iterations;
exact	FALSE (default). If TRUE the program performs <i>max.iter</i> swithcing steps, otherwise the program will count also the not-performed swithcing steps;
path	The directory in which the routine stores the outputs;
K	The number of network that has to be generated;
delimiters	list(negative='-',positive='+') (default):a list with 'positive' and 'negative' names identifying the character encoding the relation used for writing the ouput with birewire.build.dsg ;
check_pos_neg	FALSE (default) : if true check if there are positive and negative loops in the same arc, if so the routine will not save the dsg and goes on with the marcov chain (it can be slower due to the chekcs);

Details

The routine creates, starting from a given path, different subfolders in order to have maximum 1000 files for folder; the SIF files are saved using `birewire.write.dsg`, an internal routine. The set is generated calling `birewire.rewire.dsg` on the last generated dsg starting from the input dsg.

Author(s)

Andrea Gobbi: <gobbi.andrea@mail.com>

References

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T.and Jurman, G.and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

birewire.sampler.undirected

Efficient generation of a null model for a given undirected graph

Description

The routine samples correctly from the null model of a given undirected graph creating a set of randomized version of the initial undirected graph.

Usage

```
birewire.sampler.undirected(adjacency,K,path,max.iter="n", accuracy=0.00005,
  verbose=TRUE,MAXITER_MUL=10,exact=FALSE,write.sparse=TRUE)
```

Arguments

adjacency	Adjacency matrix of the initial undirected graph. Since 3.6.0 this matrix can contain also NAs and the position of such entries will be preserved by the SA;
K	The number of networks that has to be generated;
path	The directory in which the routine stores the outputs;
max.iter	"n" (default) see birewire.rewire.undirected for references
accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
verbose	TRUE (default). When TRUE a progression bar is printed during computation.
MAXITER_MUL	10 (default). If <i>exact</i> == <i>TRUE</i> in order to prevent a possible infinite loop the program stops anyway after MAXITER_MUL*max.iter iterations;
exact	FALSE (default). If TRUE the program performs <i>max.iter</i> swithcing steps, otherwise the program will count also the not-performed swithcing steps;
write.sparse	TRUE (default). If FALSE the table is written as an R data.frame (long time and more space needed)

Details

The routine creates, starting from the given path, different subfolders in order to have maximum 1000 files for folder . Moreover the incidence matrices are saved using [write_stm_CLUTO](#) (sparse matrices) that can be loaded using [read_stm_CLUTO](#). The set is generated calling [birewire.rewire.undirected](#) on the last generated graph starting from the input graph.

Author(s)

Andrea Gobbi: <gobbi.andrea@mail.com>

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* *Bioinformatics* 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* *Bioinformatics* 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Gobbi, A. and Jurman, G. (2013) *Theoretical and algorithmic solutions for null models in network theory* (Doctoral dissertation) <http://eprints-phd.biblio.unitn.it/1125/>

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

`birewire.similarity` *Compute the Jaccard similarity index between two binary matrices with the same number of non-null entries and the sam row- and column-wise sums.*

Description

Compute the Jaccard similarity index between two binary matrices with the same number of non-null entries and the sam row- and column-wise sums. The function accept also two *igraph* objects.

Usage

```
birewire.similarity( m1,m2)
```

Arguments

`m1` First matrix or graph;
`m2` Second matrix or graph.

Details

The **Jaccard** index between two sets M and N is defined as:

$$|M \cup N| / |M \cap N|$$

With M and N binary matrices, the Jaccard index is computed as:

$$\frac{\sum N_{i,j} \wedge M_{i,j}}{\sum N_{i,j} \vee M_{i,j}}$$

The Jaccard index ranges between 0 and 1 and since 3.6.0 can be computed also among matrix with NAs.

Value

Returns the Jaccard similarity index between the objects.

Author(s)

Andrea Gobbi
 Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

Examples

```
library(igraph)
library(BiRewire)
g <- graph.bipartite( rep(0:1,length=10), c(1:10))
g2=birewire.rewire.bipartite(g)

birewire.similarity(get.incidence(g,sparse=FALSE),get.incidence(g2,sparse=FALSE))
birewire.similarity(g,g2)
```

biewire.similarity.dsg

Compute the Jaccard similarity index between dsg.

Description

Compute the Jaccard similarity index between dsg objects described in the same way (matrices of graphs).

Usage

```
biewire.similarity.dsg( m1,m2)
```

Arguments

m1	First dsg;
m2	Second dsg.

Details

See [biewire.similarity](#) for more details.

Value

Returns the Jaccard similarity index between the objects.

Author(s)

Andrea Gobbi
Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

Examples

```
library(BiRewire)
data(test_dsg)
dsg <- biewire.induced.bipartite(test_dsg,sparse=FALSE)
biewire.similarity.dsg(dsg,biewire.rewire.dsg(dsg))
dsg <- biewire.induced.bipartite(test_dsg,sparse=TRUE)
biewire.similarity.dsg(dsg,biewire.rewire.dsg(dsg))
```

biewire.slum.to.sparseMatrix

The function transforms a triplet sparse matrix from slum package to a Matrix sparse matrix.

Description

Transform a triplet sparse matrix from *slum* package to a *Matrix* sparse matrix that can be used by *igraph* for creating a network. This function could be used in order to analyze graphs obtained from samplers routines ([biewire.sampler.undirected](#),[biewire.sampler.dsg](#) and [biewire.sampler.bipartite](#).)

Usage

```
birewire.slum.to.sparseMatrix( simple_triplet_matrix_sparse)
```

Arguments

`simple_triplet_matrix_sparse`
A triplet sparse matrix, usually the object coming from `read_stm_CLUTO`.

Value

Returns an Matrix sparse matrix that could be used for building an `igraph` graph using `graph.adjacency`.

Author(s)

Andrea Gobbi
Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

```
birewire.visual.monitoring.bipartite
```

Visual monitoring of the Markov chain underlying the SA for directed graphs.

Description

This function generates a cascade-sampling from the model at different switching steps given in *sequence*. For each step the routine computes the pairwise Jaccard distance (1-JI) among the samples and performs, on the resulting matrix, a dimensional scaling reduction (using `Rtsne`). If *display* is set to `TRUE` the relative plot is displayed.

Usage

```
birewire.visual.monitoring.bipartite(data,accuracy=0.00005,verbose=FALSE,MAXITER_MUL=10,
  exact=FALSE,n.networks=100,perplexity=15,sequence=c(1,5,100,"n"),ncol=2,
  nrow=length(sequence)/ncol,display=TRUE)
```

Arguments

<code>data</code>	The initial bipartite graph, either an incidence matrix or an <i>igraph</i> bipartite graph object. Since 3.6.0, if the matrix is provided, such matrix can contain also NAs and the position of such entries will be preserved by the SA;
<code>accuracy</code>	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
<code>verbose</code>	TRUE (default). When TRUE a progression bar is printed during computation.
<code>MAXITER_MUL</code>	10 (default). If <i>exact</i> == <code>TRUE</code> in order to prevent a possible infinite loop the program stops anyway after <code>MAXITER_MUL*max.iter</code> iterations;
<code>exact</code>	FALSE (default). If TRUE the program performs <i>max.iter</i> switching steps, otherwise the program will count also the not-performed switching steps;
<code>n.networks</code>	100 (default): the number of network generated for each step defined in <i>sequence</i> ;

perplexity	15 (default): the value of perplexity passed to the function <code>Rtsne</code> ;
sequence	<code>c(1,5,100,"n")</code> (default) the sequence of step for wich generating a sampler (see <code>birewire.sampler.b</code>);
ncol	2 (default). The number of column in the plot;
nrow	<code>length(sequence)/ncol</code> (default). The number of row in the plot;
display	TRUE (default). If TRUE the result is displayed.

Details

For each value p in *sequence* (it that can also contain the special character "n", see `birewire.rewire.bipartite`), the routine generates n .*networks* sampled each p SS from the SA initialized with the given *data*. Pariwise distance are computed using the Jaccard distance and the resulting matrix is the input for the dimensional scaling performed by the function `Rtsne`. An explorative plot is displayed if *display* is set to TRUE.

Value

A list containing the list containing the distance matrices *dist* and the list containing the tsne results *Rtsne*.

Author(s)

Andrea Gobbi
Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Jaccard, P. (1901), *Étude comparative de la distribution florale dans une portion des Alpes et des Jura*, Bulletin de la Société Vaudoise des Sciences Naturelles 37: 547–579.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

Van der Maaten, L.J.P. and Hinton, G.E. Visualizing High-Dimensional Data Using t-SNE. Journal of Machine Learning Research 9(Nov):2579-2605, 2008

Examples

```
library(BiRewire)
g <- graph.bipartite( rep(0:1,length=100), c(1:100))
tsne = birewire.visual.monitoring.bipartite(g,display=FALSE,n.networks=10,perplexity=1)
```

birewire.visual.monitoring.dsg

Visual monitoring of the Markov chain underlying the SA for dsgs.

Description

This function generates a cascade-sampling from the model at different switching steps given in *sequence*. For each step the routine computes the pairwise Jaccard distance (1-JI) among the samples and performs, on the resulting matrix, a dimensional scaling reduction (using [Rtsne](#)). If *display* is set to *TRUE* the relative plot is displayed.

Usage

```
birewire.visual.monitoring.dsg(data,accuracy=0.00005,verbose=FALSE,MAXITER_MUL=10,exact=FALSE,n.networks=100,
sequence.pos=c(1,5,100,"n"),
sequence.neg=c(1,5,100,"n"),ncol=2,nrow=length(sequence.pos)/ncol,display=TRUE)
```

Arguments

<code>data</code>	The initial dsg either in matrix or graph formulation 9see birewire.induced.bipartite .
<code>accuracy</code>	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
<code>verbose</code>	TRUE (default). When TRUE a progression bar is printed during computation.
<code>MAXITER_MUL</code>	10 (default). If <i>exact</i> == <i>TRUE</i> in order to prevent a possible infinite loop the program stops anyway after <code>MAXITER_MUL*max.iter</code> iterations;
<code>exact</code>	FALSE (default). If TRUE the program performs <i>max.iter</i> switching steps, otherwise the program will count also the not-performed switching steps;
<code>n.networks</code>	100 (default): the number of network generated for each step defined in <i>sequence</i> ;
<code>perplexity</code>	15 (default): the value of perplexity passed to the function Rtsne ;
<code>sequence.pos</code>	<code>c(1,5,100,"n")</code> (default) the sequence of step for wich generating a sampler (see birewire.sampler.ds) for the positive part of <i>data</i>
<code>sequence.neg</code>	same as <i>sequence.pos</i> but for the negative part
<code>ncol</code>	2 (default). The number of column in the plot;
<code>nrow</code>	<code>length(sequence)/ncol</code> (default). The number of row in the plot;
<code>display</code>	TRUE (default). If TRUE the result of tsne is displayed.

Details

See [birewire.visual.monitoring.bipartite](#) for more details.

Value

A list containing the list containing the distance matrices *dist* and the list containing the tsne results *Rtsne*.

Author(s)

Andrea Gobbi
 Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

References

Iorio, F. and and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Jaccard, P. (1901), *Étude comparative de la distribution florale dans une portion des Alpes et des Jura*, Bulletin de la Société Vaudoise des Sciences Naturelles 37: 547–579.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

Van der Maaten, L.J.P. and Hinton, G.E. Visualizing High-Dimensional Data Using t-SNE. Journal of Machine Learning Research 9(Nov):2579-2605, 2008

Examples

```
library(BiRewire)
data(test_dsg)
##bigger dsg
test_dsg_2=test_dsg
test_dsg_2[,1]=paste(test_dsg_2[,1], "_", sep="")
test_dsg_2[,3]=paste(test_dsg_2[,3], "_", sep="")

dsg <- biewire.induced.bipartite(rbind(test_dsg, test_dsg_2), sparse=FALSE)

tsne = biewire.visual.monitoring.dsg(dsg, exact=TRUE, sequence.pos=c(1,2,"n",100),
  sequence.neg=c(1,2,"n",60), n.networks=50, perplexity=1)
```

biewire.visual.monitoring.undirected

Visual monitoring of the Markov chain underlying the SA for undirected graphs.

Description

This function generates a cascade-sampling from the model at different switching steps given in *sequence*. For each step the routine computes the pairwise Jaccard distance (1-JI) among the samples and performs, on the resulting matrix, a dimensional scaling reduction (using `Rtsne`). If `display` is set to `TRUE` the relative plot is displayed.

Usage

```
birewire.visual.monitoring.undirected(data, accuracy=0.00005, verbose=FALSE, MAXITER_MUL=10,
  exact=FALSE, n.networks=100, perplexity=15, sequence=c(1, 5, 100, "n"), ncol=2,
  nrow=length(sequence)/ncol, display=TRUE)
```

Arguments

data	The initial undirected graph, either an adjacency matrix or an <i>igraph</i> undirected graph object. Since 3.6.0, if the matrix is provided, such matrix can contain also NAs and the position of such entries will be preserved by the SA;
accuracy	0.00005 (default) is the desired level of accuracy reflecting the average distance between the Jaccard index at the N-th step and its analytically derived fixed point in terms of fraction of common edges;
verbose	TRUE (default). When TRUE a progression bar is printed during computation.
MAXITER_MUL	10 (default). If <i>exact==TRUE</i> in order to prevent a possible infinite loop the program stops anyway after <code>MAXITER_MUL*max.iter</code> iterations;
exact	FALSE (default). If TRUE the program performs <i>max.iter</i> swithcing steps, otherwise the program will count also the not-performed swithcing steps;
n.networks	100 (default): the number of network generated for each step defined in <i>sequence</i> ;
perplexity	15 (default): the value of perplexity passed to the function Rtsne ;
sequence	<code>c(1,5,100,"n")</code> (default) the sequence of step for wich generating a sampler (see birewire.sampler.undirected)
ncol	2 (default). The number of column in the plot;
nrow	<code>length(sequence)/ncol</code> (default). The number of row in the plot;
display	TRUE (default). If TRUE the result of tsne is displayed.

Details

For each value *p* in *sequence* (it that can also contain the special character "n", see [birewire.rewire.bipartite](#)), the routine generates *n.networks* sampled each *p* SS from the SA initialized with the given *data*. Pariwise distance are computed using the Jaccard distance and the resulting matrix is the input for the dimensional scaling performed by the function [Rtsne](#). An explorative plot is displayed if *display* is set to TRUE.

Value

A list containing the list containing the distance matrices *dist* and the list containing the tsne results *Rtsne*.

Author(s)

Andrea Gobbi
 Maintainer: Andrea Gobbi <gobbi.andrea@mail.com>

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

Iorio, F. and Bernardo-Faura, M. and Gobbi, A. and Cokelaer, T. and Jurman, G. and Saez-Rodriguez, J. (2016) *Efficient randomization of biological networks while preserving functional characterization of individual nodes* Bioinformatics 2016 1 (17):542 doi: 10.1186/s12859-016-1402-1.

Jaccard, P. (1901), *Étude comparative de la distribution florale dans une portion des Alpes et des Jura*, Bulletin de la Société Vaudoise des Sciences Naturelles 37: 547–579.

R. Milo, N. Kashtan, S. Itzkovitz, M. E. J. Newman, U. Alon (2003), *On the uniform generation of random graphs with prescribed degree sequences*, eprint arXiv:cond-mat/0312028

Van der Maaten, L.J.P. and Hinton, G.E. Visualizing High-Dimensional Data Using t-SNE. Journal of Machine Learning Research 9(Nov):2579-2605, 2008

Examples

```
library(BiRewire)
g <- erdos.renyi.game(1000,0.1)
tsne = biwire.visual.monitoring.undirected(g,display=FALSE,n.networks=10,perplexity=1)
```

BRCA_binary_matrix *TCGA Breast Cancer data*

Description

Breast cancer samples and their respective mutations downloaded from the Cancer Cancer Genome Atlas (TCGA), used in *Gobbi et al.*. Germline mutations were filtered out of the list of reported mutations; synonymous mutations and mutations identified as benign and tolerated were also removed from the dataset. The bipartite graph resulting when considering this matrix as an incidence matrix has $n_r = 757$, $n_c = 9757$, $e = 19758$ for an edge density equal to 0.27%.

Usage

```
data(BRCA_binary_matrix)
```

Source

<http://tcga.cancer.gov/dataportal/>

References

Gobbi, A. and Iorio, F. and Dawson, K. J. and Wedge, D. C. and Tamborero, D. and Alexandrov, L. B. and Lopez-Bigas, N. and Garnett, M. J. and Jurman, G. and Saez-Rodriguez, J. (2014) *Fast randomization of large genomic datasets while preserving alteration counts* Bioinformatics 2014 30 (17): i617-i623 doi: 10.1093/bioinformatics/btu474.

test_dsg

Tool example of dsg

Description

A simple dsg for testing routines.

Usage

data(test_dsg)

Index

- * **bipartite graph, incidence matrix**
 - birewire.bipartite.from.incidence, [9](#)
- * **bipartite graph, projection, rewire**
 - birewire.rewire.bipartite.and.projections, [14](#)
- * **bipartite graph, rewire**
 - birewire.rewire.bipartite, [12](#)
 - birewire.sampler.bipartite, [19](#)
- * **datasets**
 - BRCA_binary_matrix, [30](#)
 - test_dsg, [31](#)
- * **directed graph, rewire, pathway, signaling**
 - birewire.build.dsg, [10](#)
 - birewire.induced.bipartite, [11](#)
 - birewire.load.dsg, [12](#)
 - birewire.rewire.dsg, [15](#)
 - birewire.sampler.dsg, [20](#)
- * **package**
 - BiRewire-package, [2](#)
- * **rewire, bipartite graph**
 - birewire.analysis.bipartite, [3](#)
 - birewire.visual.monitoring.bipartite, [25](#)
 - birewire.visual.monitoring.dsg, [27](#)
 - birewire.visual.monitoring.undirected, [28](#)
- * **rewire, undirected graph**
 - birewire.analysis.undirected, [7](#)
- * **rewire,dsg**
 - birewire.analysis.dsg, [5](#)
- * **similarity,jaccard**
 - birewire.similarity, [23](#)
 - birewire.similarity.dsg, [24](#)
- * **slum, Matrix,sparse matrix**
 - birewire.slum.to.sparseMatrix, [24](#)
- * **undirected graph, rewire**
 - birewire.rewire.undirected, [17](#)
 - birewire.sampler.undirected, [21](#)
- birewire.analysis.undirected, [7](#)
- birewire.bipartite.from.incidence, [9](#)
- birewire.build.dsg, [10](#), [11](#), [16](#), [21](#)
- birewire.induced.bipartite, [6](#), [10](#), [11](#), [12](#), [15](#), [20](#), [27](#)
- birewire.load.dsg, [11](#), [12](#), [15](#), [20](#)
- birewire.rewire.bipartite, [6](#), [12](#), [16](#), [26](#), [29](#)
- birewire.rewire.bipartite.and.projections, [14](#)
- birewire.rewire.dsg, [15](#)
- birewire.rewire.undirected, [17](#), [22](#)
- birewire.sampler.bipartite, [19](#), [24](#), [26](#)
- birewire.sampler.dsg, [20](#), [24](#), [27](#)
- birewire.sampler.undirected, [21](#), [24](#), [29](#)
- birewire.similarity, [23](#), [24](#)
- birewire.similarity.dsg, [6](#), [24](#)
- birewire.slum.to.sparseMatrix, [24](#)
- birewire.visual.monitoring.bipartite, [25](#), [27](#)
- birewire.visual.monitoring.dsg, [27](#)
- birewire.visual.monitoring.undirected, [28](#)
- BRCA_binary_matrix, [30](#)
- get.adjacency, [7](#), [17](#)
- get.incidence, [4](#), [12](#)
- graph.adjacency, [25](#)
- graph.incidence, [9](#)
- igraph, [4](#), [6](#), [7](#), [9](#), [11](#), [12](#), [17](#), [25](#)
- read_stm_CLUTO, [19](#), [22](#), [25](#)
- Rtsne, [25–29](#)
- test_dsg, [31](#)
- write_stm_CLUTO, [19](#), [22](#)
- BiRewire (BiRewire-package), [2](#)
- BiRewire-package, [2](#)
- birewire.analysis.bipartite, [3](#), [6](#), [14](#)
- birewire.analysis.dsg, [5](#)