

Introduction to RBM package

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October 30, 2024

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1 Overview

This document provides an introduction to the `RBM` package. The `RBM` package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the `RBM` package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code.
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 13

> which(myresult$permutation_p<=0.05)

[1] 66 239 249 263 416 459 553 583 593 608 698 786 894

> sum(myresult$bootstrap_p<=0.05)

[1] 18

> which(myresult$bootstrap_p<=0.05)

[1] 5 113 121 122 140 159 249 282 458 495 507 622 676 688 702 745 749 802

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 0

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 16

> which(myresult2$bootstrap_p<=0.05)

[1] 43 125 235 287 318 373 380 386 493 629 644 654 689 704 997 998

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM_F function: normdata_F simulates a standardized gene expression data and unifdata_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1   3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p    3000   -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 72

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 53

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 60

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 1 3 5 31 36 55 56 65 90 103 119 138 153 154 181 182 197 202 211
[20] 224 226 245 273 294 295 305 308 334 346 368 374 390 391 424 513 519 526 537
[39] 567 578 591 599 613 629 639 646 657 659 686 706 735 737 739 770 773 779 791
[58] 793 813 814 831 835 838 862 869 870 880 926 948 952 982 993

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 1 5 31 55 56 62 65 106 119 138 153 181 182 197 202 211 226 245 273
[20] 294 295 305 368 374 390 424 526 567 591 613 639 646 657 659 686 706 735 770
[39] 773 779 791 793 813 814 831 835 838 862 869 870 880 982 993

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 1 31 36 55 56 62 65 69 90 103 106 119 153 154 168 181 182 197 202
[20] 211 224 226 245 273 288 294 295 305 346 368 374 390 424 513 526 537 567 613
[39] 639 646 657 686 706 735 770 773 779 791 793 813 814 831 835 838 862 870 880
[58] 948 982 993

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

```

```

[1] 30

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 13

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 19

> which(con2_adjp<=0.05/3)

[1] 56 202 211 245 294 424 646 735 770 773 813 835 982

> which(con3_adjp<=0.05/3)

[1] 56 65 119 197 202 211 245 294 368 374 567 646 706 735 791 814 835 838 982

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1  3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p   3000   -none-  numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 51

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 51

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 46

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

```

```

[1] 3 13 15 45 62 101 160 171 174 190 231 251 270 296 329 350 351 362 456
[20] 458 471 472 474 497 501 535 545 558 560 572 580 612 647 658 661 664 670 743
[39] 749 783 786 844 846 851 865 877 897 907 929 931 978

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 3 15 45 62 101 160 174 231 251 291 296 329 350 351 362 364 383 413 426
[20] 456 471 472 474 497 501 505 524 535 545 558 560 571 572 612 658 661 670 730
[39] 743 749 783 844 846 865 877 887 897 925 929 931 978

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 15 45 62 101 160 171 174 190 231 251 296 329 338 350 351 362 458 467 472
[20] 474 501 524 535 545 558 560 572 580 612 658 661 670 715 730 731 743 749 783
[39] 877 887 897 907 923 931 932 978

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 5

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 4

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 1

```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of **RBM_T** in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the **RBM_T** function and presenting the results for further validation and investigations.

```
> system.file("data", package = "RBM")
```

```
[1] "F:/biocbuild/bbs-3.20-bioc/tmpdir/RtmpcPQI9t/Rinst19438b116410/RBM/data"
```

```
> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)
```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59032	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		

exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268

```
> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(diff_results$ordfit_pvalue<=0.05)
```

```
[1] 47
```

```
> sum(diff_results$permutation_p<=0.05)
```

```

[1] 52

> sum(diff_results$bootstrap_p<=0.05)

[1] 74

> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj_p<=0.05)

[1] 0

> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj_p<=0.05)

[1] 3

> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj_p<=0.05)

[1] 12

> diff_list_perm <- which(perm_adj_p<=0.05)
> diff_list_boot <- which(boot_adj_p<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t)
> print(sig_results_perm)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
627 cg00612467 0.04777553    0.03783457    0.05380982    0.05582291
764 cg00730260 0.90471270    0.90542290    0.91002680    0.91258610
928 cg00901493 0.03737166    0.03903724    0.04684618    0.04981432
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
627    0.04740551    0.05332965    0.05775211    0.05579710
764    0.90575890    0.88760470    0.90756300    0.90946790
928    0.04490690    0.04204062    0.05050039    0.05268215
      diff_results$ordfit_t[diff_list_perm]
627                                -1.797392
764                                -1.560713
928                                -1.982308
      diff_results$permutation_p[diff_list_perm]
627                                0
764                                0
928                                0

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t)
> print(sig_results_boot)

```


	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
95	cg00081975	0.03633894	0.04975194	0.06024723	0.05598723
146	cg00134539	0.61101320	0.53321780	0.45999340	0.46787420
259	cg00234961	0.04192170	0.04321576	0.05707140	0.05327565
280	cg00260778	0.64319890	0.60488960	0.56735060	0.53150910
285	cg00263760	0.09050395	0.10197760	0.14801710	0.12242400
632	cg00615377	0.11265030	0.16140570	0.19404450	0.17468600
677	cg00651216	0.06825629	0.12529090	0.14409190	0.13907250
802	cg00772000	0.13695750	0.18033460	0.19770870	0.17655360
833	cg00814580	0.09348613	0.09619816	0.12010440	0.11534240
851	cg00830029	0.58362500	0.59397870	0.64739610	0.67269640
911	cg00888479	0.07388961	0.07361080	0.10149800	0.09985076
979	cg00945507	0.13432250	0.23854600	0.34749760	0.28903340
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
95	0.04561792	0.05115624	0.06068253	0.06168212	
146	0.67191510	0.63137380	0.47929610	0.45428300	
259	0.04030003	0.03996053	0.05086962	0.05445672	
280	0.61920530	0.61925200	0.46753250	0.55632410	
285	0.11693600	0.10650430	0.12281160	0.12310430	
632	0.12573100	0.14483660	0.16338240	0.20130510	
677	0.07669587	0.09597587	0.11690440	0.15194540	
802	0.11228440	0.13148770	0.17138290	0.20094910	
833	0.09577040	0.11598850	0.12860890	0.14111200	
851	0.50820240	0.34657470	0.66276570	0.64634510	
911	0.08633986	0.06765189	0.09070268	0.12417730	
979	0.11848510	0.16653850	0.30718420	0.26624740	
	diff_results\$ordfit_t[diff_list_boot]				
95	-2.654324				
146	5.636263				
259	-2.833203				
280	4.337628				
285	-2.993292				
632	-3.722206				
677	-3.457874				
802	-3.169620				
833	-3.278186				
851	-2.986319				
911	-3.490240				
979	-4.968792				
	diff_results\$bootstrap_p[diff_list_boot]				
95	0				
146	0				
259	0				
280	0				
285	0				

632	0
677	0
802	0
833	0
851	0
911	0
979	0