

# Introduction to RBM package

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October 14, 2015

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## 1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

## 2 Getting started

The RBM package can be installed and loaded through the following R code.  
Install the RBM package with:

```
> source("http://bioconductor.org/biocLite.R")
> biocLite("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The  $p$ -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
<code>ordfit_t</code>	1000	-none-	numeric
<code>ordfit_pvalue</code>	1000	-none-	numeric
<code>ordfit_beta0</code>	1000	-none-	numeric
<code>ordfit_beta1</code>	1000	-none-	numeric
<code>permutation_p</code>	1000	-none-	numeric
<code>bootstrap_p</code>	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```
[1] 0
```

```

> which(myresult$permutation_p<=0.05)

integer(0)

> sum(myresult$bootstrap_p<=0.05)

[1] 18

> which(myresult$bootstrap_p<=0.05)

[1] 142 155 208 236 342 544 620 625 738 758 786 820 821 868 883 892 899 966

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 0

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 17

> which(myresult2$bootstrap_p<=0.05)

[1] 104 120 190 209 236 373 436 516 520 608 626 772 777 798 867 917 970

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM\_F function: normdata\_F simulates a standardized gene expression data and unifdata\_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none- numeric
ordfit_pvalue 3000   -none- numeric
ordfit_beta1  3000   -none- numeric
permutation_p 3000   -none- numeric
bootstrap_p   3000   -none- numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 53

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 69

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 57

> which(myresult_F$permutation_p[, 1]<=0.05)

[1]  4 15 20 62 78 94 99 116 132 148 151 159 182 210 231 248 313 364 381
[20] 387 393 397 399 400 410 445 455 500 583 587 594 636 669 717 726 729 733 735
[39] 745 748 764 784 827 837 839 866 916 921 936 955 957 963 997

> which(myresult_F$permutation_p[, 2]<=0.05)

[1]  4 15 18 20 61 62 78 99 102 116 124 132 148 151 159 182 191 231 234
[20] 240 248 353 364 379 387 393 397 399 400 410 412 428 430 445 447 455 462 488
[39] 500 583 587 589 594 615 636 658 669 679 717 726 729 733 735 736 745 748 764
[58] 784 797 827 829 837 839 916 921 936 955 957 963

> which(myresult_F$permutation_p[, 3]<=0.05)

[1]  4 15 16 18 62 78 99 102 116 124 132 148 151 159 182 231 234 240 248
[20] 379 387 393 397 399 400 410 445 455 486 500 583 587 589 594 636 669 717 729
[39] 735 745 748 764 767 784 797 822 827 837 839 851 916 936 946 952 955 957 963

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 6

```

```

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 11

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 6

> which(con2_adjp<=0.05/3)

[1] 99 159 182 387 399 500 587 729 748 784 839

> which(con3_adjp<=0.05/3)

[1] 15 148 159 399 500 587

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

      Length Class  Mode
ordfit_t      3000  -none- numeric
ordfit_pvalue 3000  -none- numeric
ordfit_beta1  3000  -none- numeric
permutation_p 3000  -none- numeric
bootstrap_p   3000  -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 47

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 51

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 35

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 7 12 18 40 87 90 93 128 134 136 142 154 180 181 243 259 284 285 329
[20] 331 346 347 391 470 543 557 571 583 593 598 609 665 675 690 697 698 757 760
[39] 773 795 812 833 838 878 936 944 971

```

```

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 1 7 12 18 40 87 90 93 134 136 142 152 158 178 180 234 237 243 259
[20] 329 331 391 401 425 447 470 475 490 571 593 594 598 609 665 675 697 698 757
[39] 760 789 795 812 818 820 825 869 878 907 936 944 971

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 7 12 18 40 87 90 93 118 128 134 180 243 331 346 391 470 490 571 593
[20] 598 665 675 697 698 757 760 773 818 833 849 878 907 936 944 949

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 9

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 8

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 5

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM\_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM\_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "/tmp/RtmpEWeGtL/Rinst4f7a499a8abf/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

```

```

      IlmnID          Beta      exmdata2[, 2]      exmdata3[, 2]
cg00000292: 1  Min.    :0.01058  Min.    :0.01187  Min.    :0.009103
cg00002426: 1  1st Qu.:0.04111  1st Qu.:0.04407  1st Qu.:0.041543
cg00003994: 1  Median :0.08284  Median :0.09531  Median :0.087042
cg00005847: 1  Mean    :0.27397  Mean    :0.28872  Mean    :0.283729
cg00006414: 1  3rd Qu.:0.52135  3rd Qu.:0.59032  3rd Qu.:0.558575
cg00007981: 1  Max.    :0.97069  Max.    :0.96937  Max.    :0.970155
(Other)    :994
exmdata4[, 2]      exmdata5[, 2]      exmdata6[, 2]      exmdata7[, 2]
Min.    :0.01019  Min.    :0.01108  Min.    :0.01937  Min.    :0.01278
1st Qu.:0.04092  1st Qu.:0.04059  1st Qu.:0.05060  1st Qu.:0.04260
Median :0.09042  Median :0.08527  Median :0.09502  Median :0.09362
Mean    :0.28508  Mean    :0.28482  Mean    :0.27348  Mean    :0.27563
3rd Qu.:0.57502  3rd Qu.:0.57300  3rd Qu.:0.52099  3rd Qu.:0.52240
Max.    :0.96658  Max.    :0.97516  Max.    :0.96681  Max.    :0.95974
NA's    :1
exmdata8[, 2]
Min.    :0.01357
1st Qu.:0.04387
Median :0.09282
Mean    :0.28679
3rd Qu.:0.57217
Max.    :0.96268

```

```

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

```

```

      Length Class  Mode
ordfit_t      1000  -none- numeric
ordfit_pvalue 1000  -none- numeric
ordfit_beta0  1000  -none- numeric
ordfit_beta1  1000  -none- numeric
permutation_p 1000  -none- numeric
bootstrap_p   1000  -none- numeric

```

```

> sum(diff_results$ordfit_pvalue<=0.05)

```

```
[1] 45
```

```

> sum(diff_results$permutation_p<=0.05)

```

```
[1] 72
```

```

> sum(diff_results$bootstrap_p<=0.05)

```

```
[1] 58
```

```
> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")  
> sum(ordfit_adj_p<=0.05)
```

```
[1] 0
```

```
> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")  
> sum(perm_adj_p<=0.05)
```

```
[1] 20
```

```
> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")  
> sum(boot_adj_p<=0.05)
```

```
[1] 3
```

```
> diff_list_perm <- which(perm_adj_p<=0.05)  
> diff_list_boot <- which(boot_adj_p<=0.05)  
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t  
> print(sig_results_perm)
```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
19	cg00016968	0.80628480	NA	0.81440820	0.83623180
97	cg00083937	0.53046980	0.60529020	0.62733150	0.65623920
103	cg00094319	0.73784280	0.73532960	0.75574900	0.73830220
106	cg00095674	0.07076291	0.05045181	0.03861991	0.03337576
131	cg00121904	0.15449580	0.17949750	0.23608110	0.24354150
146	cg00134539	0.61101320	0.53321780	0.45999340	0.46787420
189	cg00176210	0.28756520	0.39161870	0.44272520	0.44725330
245	cg00224508	0.04479948	0.04972043	0.04152814	0.04189373
251	cg00230368	0.05546448	0.04403809	0.04143668	0.03345086
280	cg00260778	0.64319890	0.60488960	0.56735060	0.53150910
285	cg00263760	0.09050395	0.10197760	0.14801710	0.12242400
504	cg00489401	0.12241640	0.20361160	0.22400680	0.21123070
627	cg00612467	0.04777553	0.03783457	0.05380982	0.05582291
764	cg00730260	0.90471270	0.90542290	0.91002680	0.91258610
772	cg00743372	0.03922780	0.02919634	0.02187972	0.02568053
848	cg00826384	0.05721674	0.05612171	0.06644259	0.06358381
851	cg00830029	0.58362500	0.59397870	0.64739610	0.67269640
887	cg00862290	0.43640520	0.54047160	0.60786800	0.56325950
928	cg00901493	0.03737166	0.03903724	0.04684618	0.04981432
979	cg00945507	0.13432250	0.23854600	0.34749760	0.28903340
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
19	0.80831380	0.73306440	0.82968340	0.84917800	
97	0.55974270	0.43157020	0.64046990	0.57876990	
103	0.67349260	0.73510200	0.75715920	0.78981220	

106	0.04693030	0.06837343	0.04534005	0.03709488
131	0.17352980	0.12564280	0.18193170	0.20847670
146	0.67191510	0.63137380	0.47929610	0.45428300
189	0.34106080	0.33765930	0.41252110	0.37024890
245	0.04208405	0.05284988	0.03775905	0.03955271
251	0.04921680	0.06053175	0.04160748	0.04809040
280	0.61920530	0.61925200	0.46753250	0.55632410
285	0.11693600	0.10650430	0.12281160	0.12310430
504	0.18750380	0.15103450	0.20825840	0.19061830
627	0.04740551	0.05332965	0.05775211	0.05579710
764	0.90575890	0.88760470	0.90756300	0.90946790
772	0.02796053	0.03512214	0.02575992	0.02093909
848	0.05230160	0.06119713	0.06542751	0.06240686
851	0.50820240	0.34657470	0.66276570	0.64634510
887	0.50259740	0.40111730	0.56646700	0.54552980
928	0.04490690	0.04204062	0.05050039	0.05268215
979	0.11848510	0.16653850	0.30718420	0.26624740

diff\_results\$ordfit\_t[diff\_list\_perm]

19	-2.446404
97	-2.541586
103	-2.268711
106	3.100324
131	-3.451679
146	5.394750
189	-3.097987
245	1.962457
251	2.189114
280	4.170347
285	-3.093997
504	-2.352810
627	-2.239498
764	-1.808081
772	2.416991
848	-2.314412
851	-2.841244
887	-3.217939
928	-2.716443
979	-4.750997

diff\_results\$permutation\_p[diff\_list\_perm]

19	0
97	0
103	0
106	0
131	0
146	0

```

189          0
245          0
251          0
280          0
285          0
504          0
627          0
764          0
772          0
848          0
851          0
887          0
928          0
979          0

```

```

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t)
> print(sig_results_boot)

```

```

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
146 cg00134539 0.6110132  0.53321780  0.4599934  0.46787420
259 cg00234961 0.0419217  0.04321576  0.0570714  0.05327565
979 cg00945507 0.1343225  0.23854600  0.3474976  0.28903340
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
146  0.67191510  0.63137380  0.47929610  0.45428300
259  0.04030003  0.03996053  0.05086962  0.05445672
979  0.11848510  0.16653850  0.30718420  0.26624740
      diff_results$ordfit_t[diff_list_boot]
146          5.394750
259         -4.052697
979         -4.750997
      diff_results$bootstrap_p[diff_list_boot]
146          0
259          0
979          0

```