

# Package ‘gespeR’

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**Title** Gene-Specific Phenotype EstimatorR

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**Description** Estimates gene-specific phenotypes from off-target confounded RNAi  
screens. The phenotype of each siRNA is modeled based on on-targeted and  
off-targeted genes, using a regularized linear regression model.

**License** GPL-3

**URL** <http://www.cbg.ethz.ch/software/gespeR>

**Collate** 'Phenotypes-class.R' 'TargetRelations-class.R'  
'gespeR-class.R' 'gespeR-concordance.R' 'gespeR-functions.R'  
'gespeR-generics.R' 'gespeR-methods.R' 'gespeR-package.R'

**NeedsCompilation** no

## R topics documented:

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gespeR-package                      *Package: Gene-Specific Phenotype Estimator*

---

### Description

This package provides a model to deconvolute off-target confounded RNAi knockdown phenotypes, and methods to investigate concordance between ranked lists of (estimated) phenotypes. The regularized linear regression model can be fitted using two different strategies. (a) Cross-validation over regularization parameters optimising the mean-squared-error of the model and (b) stability selection of covariates (genes) based on a method by Nicolai Meinshausen et al.

### Author(s)

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### References

Fabian Schmich et. al, Deconvoluting Off-Target Confounded RNA Interference Screens (2014).

### See Also

[gespeR](#)

**Examples**

```
# Read phenotypes
phenos <- lapply(LETTERS[1:4], function(x) {
  sprintf("Phenotypes_screen_%s.txt", x)
})
phenos <- lapply(phenos, function(x) {
  Phenotypes(system.file("extdata", x, package="gespeR"),
             type = "SSP",
             col.id = 1,
             col.score = 2)
})
phenos
plot(phenos[[1]])

# Read target relations
tr <- lapply(LETTERS[1:4], function(x) {
  sprintf("TR_screen_%s.rds", x)
})
tr <- lapply(tr, function(x) {
  TargetRelations(system.file("extdata", x, package="gespeR"))
})
tr[[1]]
tempfile <- paste(tempfile(pattern = "file", tmpdir = tempdir()), ".rds", sep="")
tr[[1]] <- unloadValues(tr[[1]], writeValues = TRUE, path = tempfile)
tr[[1]]
tr[[1]] <- loadValues(tr[[1]])
tr[[1]]

# Fit gespeR models with cross validation
res.cv <- lapply(1:length(phenos), function(i) {
  gespeR(phenotypes = phenos[[i]],
        target.relations = tr[[i]],
        mode = "cv",
        alpha = 0.5,
        ncores = 1)
})
summary(res.cv[[1]])
res.cv[[1]]
plot(res.cv[[1]])

# Extract scores
ssp(res.cv[[1]])
gsp(res.cv[[1]])
head(scores(res.cv[[1]]))

# Fit gespeR models with stability selection
res.stab <- lapply(1:length(phenos), function(i) {
  gespeR(phenotypes = phenos[[i]],
        target.relations = tr[[i]],
        mode = "stability",
        nbootstrap = 100,
        fraction = 0.67,
```

```

        threshold = 0.75,
        EV = 1,
        weakness = 0.8,
        ncores = 1)
    })
summary(res.stab[[1]])
res.stab[[1]]
plot(res.stab[[1]])

# Extract scores
ssp(res.stab[[1]])
gsp(res.stab[[1]])
head(scores(res.stab[[1]]))

# Compare concordance between stability selected GSPs and SSPs
conc.gsp <- concordance(lapply(res.stab, gsp))
conc.ssp <- concordance(lapply(res.stab, ssp))

pl.gsp <- plot(conc.gsp) + ggtitle("GSPs\n")
pl.ssp <- plot(conc.ssp) + ggtitle("SSPs\n")

if (require(grid)) {
  grid.newpage()
  pushViewport(viewport(layout = grid.layout(1, 2) ) )
  print(pl.gsp, vp = viewport(layout.pos.row = 1, layout.pos.col = 1))
  print(pl.ssp, vp = viewport(layout.pos.row = 1, layout.pos.col = 2))
} else {
  plot(pl.gsp)
  plot(pl.ssp)
}

```

---

annotate.gsp

*annotate.gsp*

---

## Description

Query Biomart HGNC symbols for the entrez identifiers of estimated GSPs. Currently, only implemented for species "hsapiens".

## Usage

```

## S4 method for signature 'Phenotypes'
annotate.gsp(object, organism = "hsapiens")

## S4 method for signature 'gespeR'
annotate.gsp(object, organism = "hsapiens")

```

## Arguments

|          |   |
|----------|---|
| object   | A <a href="#">gespeR</a> or <a href="#">Phenotypes</a> object |
| organism | String indicating the biomaRt organism                        |

**Value**

data.frame containing gene identifier, gene symbol and phenotypic score

**Author(s)**

Fabian Schmich

**See Also**

[gsp](#)  
[ssp](#)  
[scores](#)

**Examples**

```
data(stabilityfits)
gspA <- gsp(stabilityfits$A)
## Not run:
  annotate.gsp(gspA)

## End(Not run)
```

---

*as.data.frame,Phenotypes-method*  
*Convert Phenotypes object to a data.frame*

---

**Description**

Convert Phenotypes object to a data.frame

**Usage**

```
## S4 method for signature 'Phenotypes'
as.data.frame(x)
```

**Arguments**

x                    A [Phenotypes](#) object

**Value**

A data.frame

**Author(s)**

Fabian Schmich

**Examples**

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
  type = "SSP",
  col.id = 1,
  col.score = 2)
as.data.frame(phenos)
```

---

```
as.data.frame.concordance
```

*Coerce method*

---

**Description**

Coerce method

**Usage**

```
## S3 method for class 'concordance'
as.data.frame(x, ...)
```

**Arguments**

|     |                      |
|-----|----------------------|
| x   | concordance object   |
| ... | additional arguments |

**Value**

data.frame

**Author(s)**

Fabian Schmich

---

```
c,Phenotypes-method
```

*Concatenate Phenotypes objects*

---

**Description**

Concatenate Phenotypes objects

**Usage**

```
## S4 method for signature 'Phenotypes'
c(x, ..., recursive = FALSE)
```

**Arguments**

x                    A [Phenotypes](#) object  
 ...                  additional [Phenotypes](#) objects  
 recursive          recursive

**Value**

A concatenated [Phenotypes](#) object

**Author(s)**

Fabian Schmich

**Examples**

```
phenos.a <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
  type = "SSP",
  col.id = 1,
  col.score = 2)
phenos.b <- Phenotypes(system.file("extdata", "Phenotypes_screen_B.txt", package = "gespeR"),
  type = "SSP",
  col.id = 1,
  col.score = 2)
c(phenos.a, phenos.b)
```

---

concordance

*Evaluate the concordance between Phenotype objects*

---

**Description**

Measures include the correlation ( $\rho$ ) between pairs of phenotypes for the same gene, the rank biased overlap ([rbo](#)) of the top and bottom of ranked lists, and the Jaccard index (J) of selected genes.

**Usage**

```
concordance(..., min.overlap = 1, cor.method = "spearman", rbo.p = 0.98,
  rbo.k = NULL, rbo.mid = NULL, uneven.lengths = TRUE)
```

**Arguments**

...                    The phenotypes to be evaluated for concordance  
 min.overlap          The minimum number of overlapping genes required  
 cor.method          A character string indicating which correlation coefficient is to be computed  
 rbo.p                The weighting parameter for rank biased overlap ([rbo](#)) in [0, 1]. High p implies strong emphasis on top ranked elements

|                |  |
|----------------|--|
| rbo.k          | The evaluation depth for rank biased overlap extrapolation   |
| rbo.mid        | The mid point to split a ranked list, e.g. in order to split positive and negative scores choose mid=0 |
| uneven.lengths | Indicator if lists have uneven lengths   |

**Value**

A [concordance](#) object with the following elements:

|            |   |
|------------|---|
| pair.test  | Indicator of compared phenotypes  |
| cor        | The correlation between pairs of phenotypes for the same gene               |
| rbo.top    | The rank biased overlap of genes evaluated at the top of the ranked list    |
| rbo.bottom | The rank biased overlap of genes evaluated at the bottom of the ranked list |
| jaccard    | The Jaccard index of selected genes   |

**Author(s)**

Fabian Schmich

**See Also**

[Phenotypes](#)

[plot.concordance](#)

[rbo](#)

**Examples**

```
data(stabilityfits)
conc <- concordance(gsp(stabilityfits$A), gsp(stabilityfits$B),
  gsp(stabilityfits$C), gsp(stabilityfits$D))
plot(conc)
```

---

gespeR-class

*gespeR*

---

**Description**

Class that represents a gespeR model. It contains a SSP [Phenotypes](#) and [TargetRelations](#) representing a siRNA knockdown experiment. When the model is fitted, it additionally contains estimated GSP [Phenotypes](#).



**Usage**

```
gespeR(phenotypes, target.relations, ...)

## S4 method for signature 'Phenotypes,TargetRelations'
gespeR(phenotypes, target.relations,
       mode = c("cv", "stability"), alpha = 0.5, nbootstrap = 100,
       fraction = 0.67, threshold = 0.9, EV = 1, weakness = 0.8,
       ncores = 1, ...)

## S4 method for signature 'numeric,Matrix'
gespeR(phenotypes, target.relations, ...)
```

**Arguments**

|                  |   |
|------------------|---|
| phenotypes       | The siRNA-specific phenotypes                         |
| target.relations | The siRNA-to-gene target relations                    |
| ...              | Additional arguments                                  |
| mode             | The mode of covariate selection ("cv" or "stability") |
| alpha            | The <a href="#">glmnet</a> mixing parameter           |
| nbootstrap       | The number of bootstrap samples                       |
| fraction         | The fraction for each bootstrap sample                |
| threshold        | The selection threshold                               |
| EV               | The expected value of wrongly selected elements       |
| weakness         | The weakness parameter for randomised lasso           |
| ncores           | The number of cores for parallel computation          |

**Value**

A [gespeR](#) object

**Slots**

|                  |  |
|------------------|--|
| SSP              | The observed siRNA-specific phenotypes                           |
| GSP              | The deconvoluted gene-specific phenotypes                        |
| target.relations | The siRNA-to-gene target relations, e.g. predicted by TargetScan |
| is.fitted        | An indicator whether the <a href="#">gespeR</a> model was fitted |
| model            | The fitted regularized linear regression model                   |

**Author(s)**

Fabian Schmich

**See Also**

[gespeR-package](#)  
[plot.gespeR](#)  
[gsp](#)  
[ssp](#)  
[scores](#)  
[stability](#)  
[target.relations](#)

**Examples**

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
  type = "SSP",
  col.id = 1,
  col.score = 2)
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
res <- gespeR(phenotypes = phenos,
  target.relations = trels,
  mode = "stability",
  nbootstrap = 100,
  fraction = 0.67,
  threshold = 0.75,
  EV = 1,
  weakness = 0.8,
  ncores = 1)
gsp(res)
```

---

gsp

*Retrieve GSPs and SSPs from [gespeR](#) objects*

---

**Description**

Retrieve GSPs and SSPs from [gespeR](#) objects

**Usage**

```
gsp(object)

## S4 method for signature 'gespeR'
gsp(object)

ssp(object)

## S4 method for signature 'gespeR'
ssp(object)
```

**Arguments**

object            A [gespeR](#) object

**Value**

A [Phenotypes](#) object of GSPs and SSPs, respectively

**Author(s)**

Fabian Schmich

**See Also**

[annotate.gsp](#)  
[scores](#)

**Examples**

```
data(stabilityfits)
gsp(stabilityfits$A)
ssp(stabilityfits$B)
```

---

join

*join*

---

**Description**

Join a [TargetRelations](#) object and a [Phenotype](#) object

**Usage**

```
join(targets, phenotypes)
```

```
## S4 method for signature 'TargetRelations,Phenotypes'
join(targets, phenotypes)
```

**Arguments**

targets            A [TargetRelations](#) object.  
phenotypes        A [Phenotypes](#) object.

**Value**

List containing the matched targets and phenotypes

**Author(s)**

Fabian Schmich

**Examples**

```

phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
  type = "SSP",
  col.id = 1,
  col.score = 2)
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
phenos <- phenos[1:17]
stripped_down <- join(targets = trels, phenotypes = phenos)

```

---

lasso.rand

*Randomized Lasso*


---

**Description**

Based on Meinshausen and Buehlmann (2009)

**Usage**

```

lasso.rand(x, y, weakness = 1, subsample = 1:nrow(x), dfmax = (ncol(x) +
  1), lambda = NULL, standardize = FALSE, intercept = FALSE, ...)

```

**Arguments**

|             |   |
|-------------|---|
| x           | The design matrix                                   |
| y           | The response vector                                 |
| weakness    | The weakness parameter                              |
| subsample   | The data subsample (default: none)                  |
| dfmax       | The maximum number of degrees of freedom            |
| lambda      | The regularisation parameter                        |
| standardize | Indicator, whether to standardize the design matrix |
| intercept   | Indicator, whether to fit an intercept              |
| ...         | Additional arguments to <a href="#">glmnet</a>      |

**Value**

A [glmnet](#) object

**Author(s)**

Fabian Schmich

**Examples**

```

y <- rnorm(50)
x <- matrix(runif(50 * 20), ncol = 20)
lasso.rand(x = x, y = y)

```

---

loadValues                      *Methods for values of [TargetRelations](#) objects*

---

### Description

Load, unload or write to file the values of a [TargetRelations](#) object

### Usage

```
loadValues(object)

## S4 method for signature 'TargetRelations'
loadValues(object)

## S4 method for signature 'gespeR'
loadValues(object)

unloadValues(object, ...)

## S4 method for signature 'TargetRelations'
unloadValues(object, writeValues = TRUE,
             overwrite = FALSE, path = NULL)

## S4 method for signature 'gespeR'
unloadValues(object, writeValues = TRUE,
             overwrite = FALSE, path = NULL)

writeValues(object, ...)

## S4 method for signature 'TargetRelations'
writeValues(object, overwrite = FALSE)
```

### Arguments

|             |   |
|-------------|---|
| object      | A <a href="#">TargetRelations</a> object or <a href="#">gespeR</a> object |
| ...         | Additional arguments  |
| writeValues | Indicator, whether to write values  |
| overwrite   | Indicator, wheter to overwrite values if file exists at path              |
| path        | The path to write out values  |

### Value

A [TargetRelations](#) object or [gespeR](#) object

### Author(s)

Fabian Schmich

### Examples

```
data(stabilityfits)
## Not run:
loadValues(stabilityfits$A)

## End(Not run)
```

---

na.rem

*Remove NA/Inf values from phenotype vectors*

---

### Description

Remove NA/Inf values from phenotype vectors

### Usage

```
na.rem(object)

## S4 method for signature 'Phenotypes'
na.rem(object)
```

### Arguments

object           A [Phenotypes](#) object

### Value

A [Phenotypes](#) object without NA scores values

### Author(s)

Fabian Schmich

### Examples

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
  type = "SSP",
  col.id = 1,
  col.score = 2)
na.rem(phenos)
```

---

|        |             |
|--------|-------------|
| path<- | <i>path</i> |
|--------|-------------|

---

**Description**

Set the path of a [TargetRelations](#) object

**Usage**

```
path(object) <- value
```

```
## S4 replacement method for signature 'TargetRelations,character'  
path(object) <- value
```

**Arguments**

|        |  |
|--------|--|
| object | A <a href="#">TargetRelations</a> object |
| value  | A string defining the path               |

**Value**

A [TargetRelations](#) object with set path

**Author(s)**

Fabian Schmich

**Examples**

```
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gesper")))  
path(trels) <- "/dev/null"
```

---

|                  |                   |
|------------------|-------------------|
| Phenotypes-class | <i>Phenotypes</i> |
|------------------|-------------------|

---

**Description**

Class used to represent various types of phenotypes, e.g. from siRNA-specific (SSP) or estimated gene-specific phenotypes (GSP).

**Usage**

```
Phenotypes(phenotypes, ...)  
  
## S4 method for signature 'character'  
Phenotypes(phenotypes, type = c("SSP", "GSP"),  
  sep = "\t", col.id = 1, col.score = 2)  
  
## S4 method for signature 'cellHTS'  
Phenotypes(phenotypes, channel, sample)  
  
## S4 method for signature 'numeric'  
Phenotypes(phenotypes, ids = NULL, type = c("SSP",  
  "GSP"))
```

**Arguments**

|            |   |
|------------|---|
| phenotypes | The phenotypes as numeric vector, path to a .txt file with two columns (1: identifiers, 2: values), or a cellHTS object |
| ...        | Additional arguments  |
| type       | The type of phenotype (GSP, SSP)  |
| sep        | The separator string  |
| col.id     | Column number for the identifier  |
| col.score  | Column number for the phenotype score   |
| channel    | The cellHTS channel identifier  |
| sample     | The cellHTS sample index  |
| ids        | The phenotype identifiers   |

**Value**

A [Phenotypes](#) object

**Slots**

type The type of represented phenotypes (i.e., "SSP" or "GSP")  
ids The phenotype identifiers (i.e., siRNA or gene ids)  
values The phenotypic values

**Author(s)**

Fabian Schmich



**See Also**

[plot.Phenotypes](#)  
[join](#)  
[gsp](#)  
[ssp](#)  
[scores](#)  
[concordance](#)

**Examples**

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),  
  type = "SSP",  
  col.id = 1,  
  col.score = 2)
```

---

|                  |                         |
|------------------|-------------------------|
| plot.concordance | <i>Plot concordance</i> |
|------------------|-------------------------|

---

**Description**

Plots boxplots of concordance evaluated between multiple Phenotype objects. Measures include the correlation ( $\rho$ ) between pairs of phenotypes for the same gene, the rank biased overlap (rbo) of the top and bottom of ranked lists, and the Jaccard index (J) of selected genes.

**Usage**

```
## S3 method for class 'concordance'  
plot(x, ...)
```

**Arguments**

|     |   |
|-----|---|
| x   | The data of class <a href="#">concordance</a> |
| ... | Additional parameters for plot                |

**Value**

Boxplots of concordance measures

**Author(s)**

Fabian Schmich

---

|             |   |
|-------------|---|
| plot.gespeR | <i>Plot method for <a href="#">gespeR</a> objects</i> |
|-------------|---|

---

**Description**

Plot method for [gespeR](#) objects

**Usage**

```
## S3 method for class 'gespeR'  
plot(x, ...)
```

**Arguments**

|     |                                 |
|-----|---------------------------------|
| x   | A <a href="#">gespeR</a> object |
| ... | Additional paramters for plot   |

**Value**

Histogram of SSPs or GSPs

**Author(s)**

Fabian Schmich

---

|                 |  |
|-----------------|--|
| plot.Phenotypes | <i>Plot method for <a href="#">Phenotype</a> objects</i> |
|-----------------|--|

---

**Description**

Plot method for [Phenotype](#) objects

**Usage**

```
## S3 method for class 'Phenotypes'  
plot(x, ...)
```

**Arguments**

|     |                                     |
|-----|-------------------------------------|
| x   | A <a href="#">Phenotypes</a> object |
| ... | Additional arguments for plot       |

**Value**

Histogram of scores

**Author(s)**

Fabian Schmich

---

rbo*Rank biased overlap (Webber et al., 2010)*

---

**Description**

Evaluates the rank biased overlap (rbo) of two ranked lists based on formula based on (32) from "A Similarity Measure for Indefinite Rankings" (Webber et al.). Two ranked lists with high rbo are very similar, whereas low rbo indicates dissimilar lists. rbo ranges between 0 and 1. In this method the extrapolated version of rbo is implemented.

**Usage**

```
rbo(s, t, p, k = floor(max(length(s), length(t))/2), side = c("top",  
"bottom"), mid = NULL, uneven.lengths = TRUE)
```

**Arguments**

|                |  |
|----------------|--|
| s              | List 1   |
| t              | List 2   |
| p              | Weighting parameter in [0, 1]. High p implies strong emphasis on top ranked elements |
| k              | Evaluation depth for extrapolation   |
| side           | Evaluate similarity between the top or the bottom of the ranked lists                |
| mid            | Set the mid point to for example only consider positive or negative scores           |
| uneven.lengths | Indicator if lists have uneven lengths   |

**Value**

rank biased overlap (rbo)

**Author(s)**

Fabian Schmich

**See Also**[concordance](#)**Examples**

```
a <- rnorm(26)  
b <- rnorm(26)  
names(a) <- names(b) <- LETTERS  
rbo(a, b, p = 0.95)
```

---

|        |               |
|--------|---------------|
| scores | <i>scores</i> |
|--------|---------------|

---

## Description

Return a named vector of phenotype scores

## Usage

```
## S4 method for signature 'Phenotypes'  
scores(object)
```

```
## S4 method for signature 'gespeR'  
scores(object, type = c("GSP", "SSP"))
```

## Arguments

|        |   |
|--------|---|
| object | A <a href="#">gespeR</a> or <a href="#">Phenotypes</a> object |
| type   | The type of phenotype scores (GSP, SSP)                       |

## Value

A named vector of scores for each phenotype identifier

## Author(s)

Fabian Schmich

## See Also

[gespeR](#)

[Phenotypes](#)

## Examples

```
data(stabilityfits)  
scores(stabilityfits$A)
```

---

simData

*Example phenotype and target relations data A, B, C, and D*

---

### Description

The data set contains simulated data for four screens. Each screen consists of a phenotype vector and target relations between siRNAs and genes, i.e. which siRNA binds which genes (on- and off-targets).

### Examples

```
pheno.a <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package="gespeR"),
  type = "SSP", col.id = 1, col.score = 2)
targets.a <- TargetRelations(system.file("extdata", "TR_screen_A.rds", package="gespeR"))
```

---

stability

*stability*

---

### Description

Retrieve a [Phenotypes](#) object with stability values from a [gespeR](#) object.

### Usage

```
stability(object)

## S4 method for signature 'gespeR'
stability(object)
```

### Arguments

object            A [gespeR](#) object

### Value

A [Phenotypes](#) object of SSPs

### Author(s)

Fabian Schmich

**Examples**

```

phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
  type = "SSP",
  col.id = 1,
  col.score = 2)
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
res <- gespeR(phenotypes = phenos,
  target.relations = trels,
  mode = "stability",
  nbootstrap = 100,
  fraction = 0.67,
  threshold = 0.75,
  EV = 1,
  weakness = 0.8,
  ncores = 1)
stab <- stability(res)
ans <- merge(as.data.frame(gsp(res)), as.data.frame(stability(res)), by = "ID")
colnames(ans)[2:3] <- c("Phenotype", "Stability")
ans[order(ans$Stability, decreasing = TRUE),]

```

---

stability.selection    *Stability Selection*

---

**Description**

Based on Meinshausen and Buehlmann (2009)

**Usage**

```

stability.selection(x, y, fraction = 0.5, threshold = 0.75, EV = 1,
  nbootstrap = 100, weakness = 1, intercept = FALSE, ncores = 1, ...)

```

**Arguments**

|            |  |
|------------|--|
| x          | The design matrix                                  |
| y          | The response vector                                |
| fraction   | The fraction for each bootstrap sample             |
| threshold  | The selection threshold                            |
| EV         | The expected value of wrongly selected elements    |
| nbootstrap | The number of bootstrap samples                    |
| weakness   | The weakness parameter for randomised lasso        |
| intercept  | Indicator, whether to fit an intercept             |
| ncores     | The number of cores for parallel computation       |
| ...        | Additional arguments to <a href="#">lasso.rand</a> |

**Value**

A list containing selected covariates with frequencies, and the fitted model

**Author(s)**

Fabian Schmich

---

|               |  |
|---------------|--|
| stabilityfits | <i>Example fits for phenotypes from example screening data A, B, C and D</i> |
|---------------|--|

---

**Description**

The data set contains four fitted gespeR models using stability selection

**Examples**

```
data(stabilityfits)
```

---

|                  |                         |
|------------------|-------------------------|
| target.relations | <i>target.relations</i> |
|------------------|-------------------------|

---

**Description**

Retrieve siRNA-to-gene target relations from a [gespeR](#) object.

**Usage**

```
target.relations(object)

## S4 method for signature 'gespeR'
target.relations(object)
```

**Arguments**

object            A [gespeR](#) object

**Value**

A [TargetRelations](#) object

**Author(s)**

Fabian Schmich

**Examples**

```
data(stabilityfits)
target.relations(stabilityfits$A)
```

---

TargetRelations-class *TargetRelations*

---

### Description

Class used to represent siRNA-to-gene on- and off-target relations for a knockdown library and a set of genes.

### Usage

```
TargetRelations(targets)

## S4 method for signature 'character'
TargetRelations(targets)

## S4 method for signature 'Matrix'
TargetRelations(targets)
```

### Arguments

targets            Path to a .rds target relations matrix file or [Matrix](#) object

### Value

A [TargetRelations](#) object

### Slots

siRNAs The siRNA identifiers  
genes The gene identifiers (Entrez)  
path The path to and .rds [TargetRelations](#) file  
is.loaded An indicator if target relations values are loaded  
values The quantitative target relation values between siRNAs and genes

### Author(s)

Fabian Schmich

### See Also

[join](#)  
[loadValues](#)  
[unloadValues](#)  
[writeValues](#)  
[values](#)  
[path<-](#)



**Examples**

```
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
```

---

|        |               |
|--------|---------------|
| values | <i>values</i> |
|--------|---------------|

---

**Description**

Retrieve the numeric siRNA-to-gene target relations from a [TargetRelations](#) object

**Usage**

```
values(object)

## S4 method for signature 'TargetRelations'
values(object)
```

**Arguments**

object            A [TargetRelations](#) object

**Value**

A [Matrix](#) object

**Author(s)**

Fabian Schmich

**Examples**

```
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
values(trels)[1:5, 1:5]
```

---

[,Phenotypes,ANY,ANY,ANY-method  
*Subsetting for Phenotype objects.*

---

**Description**

Subsetting for Phenotype objects.

**Usage**

```
## S4 method for signature 'Phenotypes,ANY,ANY,ANY'
x[i, j, ..., drop = TRUE]
```

**Arguments**

|      |                                     |
|------|-------------------------------------|
| x    | A <a href="#">Phenotypes</a> object |
| i    | The subsetting indices              |
| j    | Subsetting indices (not used)       |
| ...  | Additional parameters               |
| drop | Drop Redundant Extent Information   |

**Value**

A [Phenotypes](#) object

**Author(s)**

Fabian Schmich

---

[,TargetRelations,ANY,ANY,ANY-method

*Subsetting for TargetRelations objects.*

---

**Description**

Subsetting for TargetRelations objects.

**Usage**

```
## S4 method for signature 'TargetRelations,ANY,ANY,ANY'
x[i, j, ..., drop = TRUE]
```

**Arguments**

|      |  |
|------|--|
| x    | A <a href="#">TargetRelations</a> object |
| i    | The row subsetting indices (siRNAs)      |
| j    | The column subsetting indices (genes)    |
| ...  | Additional parameters                    |
| drop | Drop Redundant Extent Information        |

**Value**

A [TargetRelations](#) object

**Author(s)**

Fabian Schmich

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