

Package ‘SeqArray’

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LinkingTo gdsfmt

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Description Big data management of genome-wide variants using the
CoreArray C++ library: genotypic data and annotations are stored in
an array-oriented manner, offering efficient access of genetic
variants using the R programming language.

License GPL-3

VignetteBuilder knitr

URL <http://corearray.sourceforge.net/tutorials/SeqArray/>,
<http://github.com/zhengxwen/SeqArray>

BugReports <http://github.com/zhengxwen/SeqArray/issues>

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SeqArray-package

*Big Data Management of Genome-wide Sequencing Variants***Description**

Big-Data Management of Genome-Wide Sequencing Variants

Details

In the era of big data, thousands of gigabyte-size data sets are challenging scientists for data management, even on well-equipped hardware. Currently, next-generation sequencing techniques are being adopted to investigate common and rare variants, making the analyses of large-scale genotypic data challenging. For example, the 1000 Genomes Project has identified approximately 38 million single nucleotide polymorphisms (SNPs), 1.4 million short insertions and deletions, and more than 14,000 larger deletions from whole-genome sequencing technologies. In the near future, new technologies, like third-generation whole-genome sequencing, will be enabling data to be generated at an unprecedented scale. The Variant Call Format (VCF) was developed for the 1000 Genomes Project, which is a generic text format for storing DNA polymorphism data such as SNPs, insertions, deletions and structural variants, together with rich annotations. However, this format is less efficient for large-scale analyses since numeric data have to be parsed from a text VCF file before further analyses. The computational burden associated with sequencing variants is especially evident with large sample and variant sizes, and it requires efficient numerical implementation and data management.

Here I introduce a high-performance C/C++ computing library CoreArray (<http://corearray.sourceforge.net>) for big-data management of genome-wide variants. CoreArray was designed

for developing portable and scalable storage technologies for bioinformatics data, allowing parallel computing at the multicore and cluster levels. It provides the genomic data structure (GDS) file format for array-oriented data: this is a universal data format to store multiple data variables in a single file. A hierarchical data structure is used to store multiple extensible data variables in the GDS format, and all datasets are stored in a single file with chunked storage layout. Here, I focus on the application of CoreArray for statisticians working in the R environment, and developed an R/Bioconductor package SeqArray to address or reduce the computational burden associated with data management of sequencing variants. The kernels of SeqArray are written in C/C++ and highly optimized. Genotypic data and annotations are stored in an array-oriented manner, offering efficient access of genetic variants using the R language. There are five key functions in SeqArray, and most of data analyses could be done using these 6 functions:

Function	Description
seqVCF2GDS	Imports VCF files
seqSummary	Gets the summary of a sequencing GDS file (# of samples, # of variants, INFO/FORMAT variables, etc)
seqSetFilter	Sets a filter to sample or variant (define a subset of data)
seqGetData	Gets data from a sequencing GDS file (from a subset of data)
seqApply	Applies a user-defined function over array margins
seqParallel	Applies functions in parallel

The 1000 Genomes Project released 39 million genetic variants for 1092 individuals, and a 26G data file was created by SeqArray to store sequencing variants with phasing information, where 2 bits were used as an atomic data type. The file size can be further reduced to 1.3G by compression algorithms without sacrificing access efficiency, since it has a large proportion of rare variants.

SeqArray will be of great interest to scientists involved in data analyses of large-scale genomic sequencing data using R environment, particularly those with limited experience of low-level C programming and parallel computing.

Webpage: <http://corearray.sourceforge.net/>

Tutorial: <http://corearray.sourceforge.net/tutorials/SeqArray/>

Forums: <http://sourceforge.net/projects/corearray/forums>

Author(s)

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Examples

```
# the file of VCF
vcf.fn <- seqExampleFileName("vcf")
vcf.fn
# or vcf.fn <- "C:/YourFolder/Your_VCF_File.vcf"

# parse the header
seqVCF.Header(vcf.fn)

# get sample id
seqVCF.SampID(vcf.fn)
```

```

# convert
seqVCF2GDS(vcf.fn, "tmp.gds")
seqSummary("tmp.gds")

# list the structure of GDS variables
f <- seqOpen("tmp.gds")
f

seqClose(f)
unlink("tmp.gds")

#####

# the file of GDS
gds.fn <- seqExampleFileName("gds")
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

# display
(f <- seqOpen(gds.fn))

# get 'sample.id'
(samp.id <- seqGetData(f, "sample.id"))
# "NA06984" "NA06985" "NA06986" ...

# get 'variant.id'
head(variant.id <- seqGetData(f, "variant.id"))

# get 'chromosome'
table(seqGetData(f, "chromosome"))

# get 'allele'
head(seqGetData(f, "allele"))
# "T,C" "G,A" "G,A" ...

# set sample and variant filters
seqSetFilter(f, sample.id=samp.id[c(2,4,6,8,10)])
set.seed(100)
seqSetFilter(f, variant.id=sample(variant.id, 10))

# get genotypic data
seqGetData(f, "genotype")

# get annotation/info/DP
seqGetData(f, "annotation/info/DP")

# get annotation/info/AA, a variable-length dataset
seqGetData(f, "annotation/info/AA")
# $length      <- indicating the length of each variable-length data
# [1] 1 1 1 1 1 1 ...
# $data        <- the data according to $length

```

```

# [1] "T" "C" "T" "C" "G" "C" ...

# get annotation/format/DP, a variable-length dataset
seqGetData(f, "annotation/format/DP")
# $length          <- indicating the length of each variable-length data
# [1] 1 1 1 1 1 1 ...
# $data            <- the data according to $length
#   variant
# sample [,1] [,2] [,3] [,4] [,5] [,6] ...
# [1,] 25 25 22 3 4 17 ...

# read multiple variables variant by variant
seqApply(f, c(geno="genotype", phase="phase", qual="annotation/id"),
FUN=function(x) print(x), as.is="none")

# get the numbers of alleles per variant
seqApply(f, "allele",
FUN=function(x) length(unlist(strsplit(x,","))), as.is="integer")

#####

# remove the sample and variant filters
seqSetFilter(f)

# calculate the frequency of reference allele,
# a faster version could be obtained by C coding
af <- seqApply(f, "genotype", FUN=function(x) mean(x=="0", na.rm=TRUE), as.is="double")
length(af)
summary(af)

#####

# run in parallel

library(parallel)

# Use option cl.core to choose an appropriate cluster size or number of cores
cl <- makeCluster(getOption("cl.cores", 2))

# run in parallel
afreq <- seqParallel(cl, f, FUN = function(gdsfile) {
seqApply(gdsfile, "genotype", as.is="double",
FUN=function(x) mean(x=="0", na.rm=TRUE))
}, split = "by.variant")

length(afreq)
summary(afreq)

stopCluster(cl)

```

```
# close the GDS file
seqClose(f)
```

seqApply

Apply Functions Over Array Margins

Description

Returns a vector or list of values obtained by applying a function to margins of arrays or matrices

Usage

```
seqApply(gdsfile, var.name, FUN,
margin = c("by.variant"),
as.is = c("list", "integer", "double", "character", "none"),
var.index = c("none", "relative", "absolute"), ...)
```

Arguments

gdsfile	a SeqVarGDSClass object
var.name	the variable name(s), see details
FUN	the function to be applied
margin	giving the dimension which the function will be applied over. E.g., for a matrix 1 indicates rows, 2 indicates columns
as.is	returned value: a list, an integer vector, etc
var.index	if "none", call FUN(x, ...) without variable index; if "relative" or "absolute", add an argument to the user-defined function FUN like FUN(index, x, ...) where index is an index of variant starting from 1 if margin = "by.variant": "relative" for indexing in the selection defined by seqSetFilter , "absolute" for indexing with respect to all data
...	optional arguments to FUN

Details

The variable name should be "sample.id", "variant.id", "position", "chromosome", "allele", "annotation/id", "annotation/qual", "annotation/filter", "annotation/info/VARIABLE_NAME", or "annotation/format/VARIABLE_NAME".

The algorithm is highly optimized by blocking the computations to exploit the high-speed memory instead of disk.

Value

A vector or list of values.

Author(s)

Xiuwen Zheng

See Also[seqSetFilter](#), [seqGetData](#), [seqParallel](#)**Examples**

```

# the file of GDS
gds.fn <- seqExampleFileName("gds")
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

# display
(f <- seqOpen(gds.fn))

# get 'sample.id'
(samp.id <- seqGetData(f, "sample.id"))
# "NA06984" "NA06985" "NA06986" ...

# get 'variant.id'
head(variant.id <- seqGetData(f, "variant.id"))

# set sample and variant filters
set.seed(100)
seqSetFilter(f, sample.id=samp.id[c(2,4,6,8,10)],
variant.id=sample(variant.id, 10))

# read multiple variables variant by variant
seqApply(f, c(geno="genotype", phase="phase", qual="annotation/id"),
FUN=function(x) print(x), as.is="none")

# get the numbers of alleles per variant
seqApply(f, "allele",
FUN=function(x) length(unlist(strsplit(x,""))), as.is="integer")

#####
# with an index of variant

seqApply(f, c(geno="genotype", phase="phase", qual="annotation/id"),
FUN=function(index, x) { print(index); print(x); index },
as.is="integer", var.index="relative")
# it is as the same as
which(seqGetFilter(f)$variant.sel)

#####
# reset sample and variant filters
seqSetFilter(f)

```

```
# calculate the frequency of reference allele,
# a faster version could be obtained by C coding
af <- seqApply(f, "genotype", FUN=function(x) mean(x==0, na.rm=TRUE),
as.is="double")
length(af)
summary(af)

# close the GDS file
seqClose(f)
```

seqClose-methods *Close a SeqArray object*

Description

Close a SeqArray file or object

Usage

```
## S4 method for signature 'SeqVarGDSCClass'
seqClose(object)
```

Arguments

object a SeqArray object

Details

If object is

- [SeqVarGDSCClass](#), close the sequencing GDS file.

Value

None.

Author(s)

Xiuwen Zheng

See Also

[seqOpen](#)

seqCompress.Option *Compression Options for Importing VCF File(s)*

Description

Get compression options for importing VCF file(s)

Usage

```
seqCompress.Option(default="ZIP.MAX", ...)
```

Arguments

default	the default compression level
...	optional arguments

Value

Return a list with a class name "seqCompress.class".

Author(s)

Xiuwen Zheng

See Also

[seqVCF2GDS](#)

Examples

```
# the file of VCF
(vcf.fn <- seqExampleFileName("vcf"))
# or vcf.fn <- "C:/YourFolder/Your_VCF_File.vcf"

# convert
seqVCF2GDS(vcf.fn, "tmp1.gds", compress.option = seqCompress.Option())
(f1 <- seqOpen("tmp1.gds"))

# does not compress the genotypic data
seqVCF2GDS(vcf.fn, "tmp2.gds", compress.option = seqCompress.Option(genotype=""))
(f2 <- seqOpen("tmp2.gds"))

# close and remove
seqClose(f1)
seqClose(f2)
unlink(c("tmp1.gds", "tmp2.gds"))
```

`seqDelete`*Delete variables in a Sequencing GDS File*

Description

Delete variables in a sequencing GDS file

Usage

```
seqDelete(gdsfile, info.varname=NULL, format.varname=NULL)
```

Arguments

`gdsfile` a [SeqVarGDSClass](#) object
`info.varname` the variables in the INFO field, i.e., "annotation/info/VARIABLE_NAME"
`format.varname` the variables in the FORMAT field, i.e., "annotation/format/VARIABLE_NAME"

Value

None.

Author(s)

Xiuwen Zheng

See Also

[seqOpen](#), [seqClose](#)

Examples

```
# the file of VCF
vcf.fn <- seqExampleFileName("vcf")
# or vcf.fn <- "C:/YourFolder/Your_VCF_File.vcf"

# convert
seqVCF2GDS(vcf.fn, "tmp.gds")

# display
(f <- seqOpen("tmp.gds", FALSE))

seqDelete(f, info.varname=c("HM2", "AA"), format.varname="DP")

# close the GDS file
seqClose(f)

# clean up the fragments, reduce the file size
cleanup.gds("tmp.gds")
```

seqExampleFileName	<i>Example GDS file</i>
--------------------	-------------------------

Description

The example GDS file for sequencing variants

Usage

```
seqExampleFileName(type=c("gds", "vcf"))
```

Arguments

type either "gds" or "vcf"

Details

A GDS sequencing-variant file was created from a subset of VCF data of the 1000 Genomes Project.

Value

Return the file name of a VCF file shipped with the package if type = "vcf", or the file name of a GDS file if type = "gds".

Author(s)

Xiuwen Zheng

Examples

```
seqExampleFileName("gds")
```

```
seqExampleFileName("vcf")
```

seqGDS2VCF	<i>Convert to a VCF file</i>
------------	------------------------------

Description

Convert a GDS file to a VCF file

Usage

```
seqGDS2VCF(gdsfile, vcf.fn, info.var=NULL, fmt.var=NULL, verbose=TRUE)
```

Arguments

<code>gdsfile</code>	a SeqVarGDSClass object
<code>vcf.fn</code>	the file name, output a file of VCF format
<code>info.var</code>	a list of variable names in the INFO field, or NULL for using all variables; character(0) for no variable in the INFO field
<code>fmt.var</code>	a list of variable names in the FORMAT field, or NULL for using all variables; character(0) for no variable in the FORMAT field
<code>verbose</code>	if TRUE, show information

Details

[seqSetFilter](#) can be used to define a subset of data for the export.

GDS – Genomic Data Structures used for storing genetic array-oriented data, and the file format used in the [gdsfmt](#) package.

VCF – The Variant Call Format (VCF), which is a generic format for storing DNA polymorphism data such as SNPs, insertions, deletions and structural variants, together with rich annotations.

Value

Return the file name of VCF file with an absolute path.

Author(s)

Xiuwen Zheng

References

The variant call format and VCFtools. Danecek P, Auton A, Abecasis G, Albers CA, Banks E, DePristo MA, Handsaker RE, Lunter G, Marth GT, Sherry ST, McVean G, Durbin R; 1000 Genomes Project Analysis Group. *Bioinformatics*. 2011 Aug 1;27(15):2156-8. Epub 2011 Jun 7.

<http://corearray.sourceforge.net/>

See Also

[seqVCF2GDS](#)

Examples

```
# the file of GDS
gds.fn <- seqExampleFileName("gds")
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

# display
(f <- seqOpen(gds.fn))

# output the first 10 samples
samp.id <- seqGetData(f, "sample.id")
seqSetFilter(f, sample.id=samp.id[1:5])
```

```

# convert
seqGDS2VCF(f, "tmp.vcf.gz")

# no INFO and FORMAT
seqGDS2VCF(f, "tmp1.vcf.gz", info.var=character(0), fmt.var=character(0))

# output BN,GP,AA,DP,HM2 in INFO (the variables are in this order), no FORMAT
seqGDS2VCF(f, "tmp2.vcf.gz", info.var=c("BN","GP","AA","DP","HM2"), fmt.var=character(0))

# read
(txt <- readLines("tmp.vcf.gz", n=20))
(txt <- readLines("tmp1.vcf.gz", n=20))
(txt <- readLines("tmp2.vcf.gz", n=20))

#####
# Users could compare the new VCF file with the original VCF file
# call "diff" in Unix (a command line tool comparing files line by line)

# using all samples and variants
seqSetFilter(f)

# convert
seqGDS2VCF(f, "tmp.vcf.gz")

# file.copy(seqExampleFileName("vcf"), "old.vcf.gz", overwrite=TRUE)
# system("diff <(gunzip -c old.vcf.gz) <(gunzip -c tmp.vcf.gz)")

# 1a2,3
# > ##fileDate=20130309
# > ##source=SeqArray_RPackage_v1.0

# LOOK GOOD!

# delete temporary files
unlink(c("tmp.vcf.gz", "tmp1.vcf.gz", "tmp2.vcf.gz"))

# close the GDS file
seqClose(f)

```

Description

Get data from a sequencing GDS file

Usage

```
seqGetData(gdsfile, var.name)
```

Arguments

<code>gdsfile</code>	a SeqVarGDSClass object
<code>var.name</code>	the variable name, see details

Details

The variable name should be "sample.id", "variant.id", "position", "chromosome", "allele", "annotation/id", "annotation/qual", "annotation/filter", "annotation/info/VARIABLE_NAME", or "annotation/format/VARIABLE_NAME".

Value

Return vectors or lists.

Author(s)

Xiuwen Zheng

See Also

[seqSetFilter](#), [seqApply](#)

Examples

```
# the file of GDS
gds.fn <- seqExampleFileName("gds")
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

# display
(f <- seqOpen(gds.fn))

# get 'sample.id'
(samp.id <- seqGetData(f, "sample.id"))
# "NA06984" "NA06985" "NA06986" ...

# get 'variant.id'
head(variant.id <- seqGetData(f, "variant.id"))

# get 'chromosome'
table(seqGetData(f, "chromosome"))

# get 'allele'
head(seqGetData(f, "allele"))
```

```

# "T,C" "G,A" "G,A" ...

# set sample and variant filters
seqSetFilter(f, sample.id=samp.id[c(2,4,6,8,10)])
set.seed(100)
seqSetFilter(f, variant.id=sample(variant.id, 10))

# get genotypic data
seqGetData(f, "genotype")

# get annotation/info/DP
seqGetData(f, "annotation/info/DP")

# get annotation/info/AA, a variable-length dataset
seqGetData(f, "annotation/info/AA")
# $length          <- indicating the length of each variable-length data
# [1] 1 1 1 1 1 1 ...
# $data            <- the data according to $length
# [1] "T" "C" "T" "C" "G" "C" ...

# get annotation/format/DP, a variable-length dataset
seqGetData(f, "annotation/format/DP")
# $length          <- indicating the length of each variable-length data
# [1] 1 1 1 1 1 1 ...
# $data            <- the data according to $length
#   variant
# sample [,1] [,2] [,3] [,4] [,5] [,6] ...
# [1,]    25  25  22   3   4  17 ...

# close the GDS file
seqClose(f)

```

seqGetFilter

Get the Filter of Samples and Variants

Description

Get the filter of samples and variants

Usage

```
seqGetFilter(gdsfile)
```

Arguments

gdsfile a [SeqVarGDSClass](#) object

Details

It is strongly suggested to call `seqOpen` instead of `openfn.gds`, since `seqOpen` will initialize the internal data for `seqGetData`, `seqApply`, etc.

Value

Return a list:

`sample.sel` a logical vector for selected samples
`variant.sel` a logical vector for selected variants

Author(s)

Xiuwen Zheng

See Also

[seqGetData](#), [seqApply](#)

Examples

```
# the file of GDS
gds.fn <- seqExampleFileName("gds")
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

# display
(f <- seqOpen(gds.fn))

# get 'sample.id'
(samp.id <- seqGetData(f, "sample.id"))
# "NA06984" "NA06985" "NA06986" ...

# get 'variant.id'
head(variant.id <- seqGetData(f, "variant.id"))

# set sample and variant filters
seqSetFilter(f, sample.id=samp.id[c(2,4,6,8,10)])
set.seed(100)
seqSetFilter(f, variant.id=sample(variant.id, 10))

# get filter
z <- seqGetFilter(f)

# the number of selected samples
sum(z$sample.sel)
# the number of selected variants
sum(z$variant.sel)

# close the GDS file
seqClose(f)
```

seqInfoNewVar	<i>Add a variable to the INFO field</i>
---------------	---

Description

Add a new variable to the INFO field in the specified GDS file.

Usage

```
seqInfoNewVar(gdsfile, var.name, variant.id, val,
              description="", compress=c("ZIP.MAX", ""), no.data.index=TRUE)
```

Arguments

gdsfile	a SeqVarGDSClass object
var.name	the variable name(s), see details
variant.id	the variant ID(s), should have the same order as IDs in the GDS file
val	a vector or a matrix with the same order as <code>variant.id</code> ; if it is a matrix, the number of columns should be equal to the length of <code>variant.id</code> , see details
description	the variable description
compress	to specify the compression algorithm: "" for no compression; see <code>compress</code> in the function add.gdsn
no.data.index	applicable only if <code>val</code> is a numeric vector or a factor variable; if <code>no.data.index=TRUE</code> , non-exist values in <code>val</code> will be replaced by NA or NaN with respect to the variants not specified in <code>variant.id</code> , and no index data associated with this variable are created

Details

The variable name should be "sample.id", "variant.id", "position", "chromosome", "allele", "annotation/id", "annotation/qual", "annotation/filter", "annotation/info/VARIABLE_NAME", or "annotation/format/VARIABLE_NAME".

The argument `val` should be integers, numeric values, a logical variable, characters or factors. If `val` is a logical variable, one-bit storage mode will be used to store this variable, which corresponds to the variable defined with 'Type=Flag' in the VCF format.

Value

None.

Author(s)

Xiuwen Zheng

See Also[seqGetData](#)**Examples**

```

# the file of GDS
gds.fn <- seqExampleFileName("gds")

file.copy(gds.fn, "test.gds", overwrite=TRUE)

# display
(f <- seqOpen("test.gds", readonly=FALSE))

# get variant IDs
variant.id <- seqGetData(f, "variant.id")

####   add variables to the INFO field   ####

set.seed(100)

seqInfoNewVar(f, "int", variant.id[1:15], sample.int(256, 15, TRUE),
"integer variable")
seqGetData(f, "annotation/info/int")

seqInfoNewVar(f, "int.2", variant.id[1:15], sample.int(256, 15, TRUE),
"integer variable", no.data.index=FALSE)
seqGetData(f, "annotation/info/int.2")

seqInfoNewVar(f, "numeric", variant.id[15:30], rnorm(16),
"numeric variable")
seqGetData(f, "annotation/info/numeric")

seqInfoNewVar(f, "numeric.2", variant.id[15:30], rnorm(16),
"numeric variable", no.data.index=FALSE)
seqGetData(f, "annotation/info/numeric.2")

seqInfoNewVar(f, "flag", variant.id[4:9], rep(c(FALSE, TRUE), 3),
"flag variable")
# stored in `bit1`
seqGetData(f, "annotation/info/flag")

seqInfoNewVar(f, "factor", variant.id,
factor(c("ABC", "DDD", "CVX")[sample(1:3, length(variant.id), TRUE)]),
"string/factor variable")
# stored in `int32` with attributes
seqGetData(f, "annotation/info/factor")

```

```

seqInfoNewVar(f, "string", variant.id,
c("ABC", "DDD", "CVX")[sample(1:3, length(variant.id), TRUE)],
"string variable")
seqGetData(f, "annotation/info/string")

# show the file
f

# the corresponding VCF file
seqGDS2VCF(f, "test.vcf.gz")
txt <- strsplit(readLines("test.vcf.gz", n=40), "\t")[-c(1:21)]

# the INFO field:
sapply(txt, function(x) x[8])

# close the GDS file
seqClose(f)

# delete the temporary files
unlink("test.gds", force=TRUE)
unlink("test.vcf.gz", force=TRUE)

```

seqMerge

Merge Multiple Sequencing GDS Files

Description

Merge multiple sequencing GDS files

Usage

```
seqMerge(gds.fn, out.fn, compress.option = seqCompress.Option(),
verbose = TRUE)
```

Arguments

gds.fn	the file names of multiple GDS files
out.fn	the output file name
compress.option	specify the compression options, by default seqCompress.Option
verbose	if TRUE, show information

Details

The current implementation of seqMerge extracts and merges the genotypic data only without any annotation. Users can specify multiple VCF files in [seqVCF2GDS](#) to export a single GDS file.

Value

None.

Author(s)

Xiuwen Zheng

See Also

[seqVCF2GDS](#)

seqOpen

Open a Sequencing GDS File

Description

Open a Sequencing GDS file

Usage

```
seqOpen(gds.fn, readonly=TRUE)
```

Arguments

<code>gds.fn</code>	the file name
<code>readonly</code>	whether read-only or not

Details

It is strongly suggested to call `seqOpen` instead of `openfn.gds`, since `seqOpen` will perform internal checking for data integrality.

Value

Return an object of class `gds.class`.

Author(s)

Xiuwen Zheng

See Also

[seqGetData](#), [seqApply](#)

Examples

```

gds.fn <- seqExampleFileName("gds")
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

# open the GDS file
gdsfile <- seqOpen(gds.fn)

# display the contents of the GDS file in a hierarchical structure
gdsfile

# close the GDS file
seqClose(gdsfile)

```

seqParallel

Apply Functions in Parallel

Description

Apply a user-defined function in parallel

Usage

```

seqParallel(cl, gdsfile, FUN = function(gdsfile, ...) NULL,
split=c("by.variant", "by.sample", "none"), .combine=NULL, ...)

```

Arguments

cl	NULL or a cluster object, created by the package parallel or snow
gdsfile	a SeqVarGDSClass object
FUN	the function to be applied
split	split the dataset by variant or sample according to multiple processes, or "none" for no split
.combine	define a function for combining results from different processes; by default, 'c' is used; if .combine=="", return invisible()
...	optional arguments to FUN

Details

If `cl = NULL` or `length(cl) == 0`, the function simply calls `FUN(gdsfile, ...)`; otherwise, it splits jobs to different processes and calls `FUN(gdsfile, ...)` on each process, the optional arguments are passed to different processes.

Value

A vector or list of values.

Author(s)

Xiuwen Zheng

See Also

[seqSetFilter](#), [seqGetData](#) [seqApply](#)

Examples

```
library(parallel)

# Use option cl.core to choose an appropriate cluster size or number of cores
cl <- makeCluster(getOption("cl.cores", 2))

# the file of GDS
gds.fn <- seqExampleFileName("gds")
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

# display
(f <- seqOpen(gds.fn))

# the uniprocessor version
afreq1 <- seqParallel(NULL, f, FUN = function(gdsfile) {
  seqApply(gdsfile, "genotype", as.is="double",
  FUN=function(x) mean(x==0, na.rm=TRUE))
}, split = "by.variant")

length(afreq1)
summary(afreq1)

# run in parallel
afreq2 <- seqParallel(cl, f, FUN = function(gdsfile) {
  seqApply(gdsfile, "genotype", as.is="double",
  FUN=function(x) mean(x==0, na.rm=TRUE))
}, split = "by.variant")

length(afreq2)
summary(afreq2)

# check
all(afreq1 == afreq2)

#####
# check -- variant splits

seqParallel(cl, f, FUN = function(gdsfile) {
  v <- seqGetFilter(gdsfile)
  sum(v$variant.sel)
```

```

}, split = "by.variant")
# [1] 674 674

#####

stopCluster(cl)

# close the GDS file
seqClose(f)

```

seqSetFilter *Set a filter to sample or variant*

Description

Set a filter to sample and/or variant

Usage

```
seqSetFilter(gdsfile, sample.id=NULL, variant.id=NULL,
             samp.sel=NULL, variant.sel=NULL, action=c("set", "push", "push+set", "pop"),
             verbose=TRUE)
```

Arguments

gdsfile	a SeqVarGDSClass object
sample.id	IDs of selected samples
variant.id	IDs of selected variants
samp.sel	a logical vector indicating the selected samples
variant.sel	a logical vector indicating the selected variants
action	"set" – set the current filter via <code>sample.id</code> , <code>variant.id</code> , <code>samp.sel</code> or <code>variant.sel</code> ; "push" – push the current filter to the stack, and it could be recovered by "pop" later, no change on the current filter; "push+set" – push the current filter to the stack, and changes the current filter via <code>sample.id</code> , <code>variant.id</code> , <code>samp.sel</code> or <code>variant.sel</code> ; "pop" – pop up the last filter
verbose	if TRUE, show information

Value

None.

Author(s)

Xiuwen Zheng

See Also

[seqGetFilter](#), [seqGetData](#), [seqApply](#)

Examples

```
# the file of GDS
gds.fn <- seqExampleFileName("gds")
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

# display
(f <- seqOpen(gds.fn))

# get 'sample.id'
(samp.id <- seqGetData(f, "sample.id"))
# "NA06984" "NA06985" "NA06986" ...

# get 'variant.id'
head(variant.id <- seqGetData(f, "variant.id"))

# get 'chromosome'
table(seqGetData(f, "chromosome"))

# get 'allele'
head(seqGetData(f, "allele"))
# "T,C" "G,A" "G,A" ...

# set sample and variant filters
seqSetFilter(f, sample.id=samp.id[c(2,4,6,8,10)])
set.seed(100)
seqSetFilter(f, variant.id=sample(variant.id, 10))

# get genotypic data
seqGetData(f, "genotype")

# close the GDS file
seqClose(f)
```

seqSlidingWindow

Apply functions via a sliding window over variants

Description

Returns a vector or list of values obtained by applying a function to a sliding window over variants

Usage

```
seqSlidingWindow(gdsfile, var.name, win.size, shift=1, FUN,
  as.is = c("list", "integer", "double", "character", "none"),
  var.index = c("none", "relative", "absolute"), ...)
```


Arguments

<code>gdsfile</code>	a SeqVarGDSClass object
<code>var.name</code>	the variable name(s), see details
<code>win.size</code>	the size of sliding window
<code>shift</code>	the number of variants to shift the window at each step
<code>FUN</code>	the function to be applied
<code>as.is</code>	returned value: a list, an integer vector, etc
<code>var.index</code>	if "none", call <code>FUN(x, ...)</code> without variable index; if "relative" or "absolute", add an argument to the user-defined function <code>FUN</code> like <code>FUN(index, x, ...)</code> where <code>index</code> is an index of variant starting from 1: "relative" for indexing in the selection defined by seqSetFilter , "absolute" for indexing with respect to all data
<code>...</code>	optional arguments to <code>FUN</code>

Details

The variable name should be "sample.id", "variant.id", "position", "chromosome", "allele", "annotation/id", "annotation/qual", "annotation/filter", "annotation/info/VARIABLE_NAME", or "annotation/format/VARIABLE_NAME".

In the user-defined function `FUN(x, ...)` or `FUN(index, x, ...)`, `x` is a list with `win.size` elements, and each element includes values for the variable(s) `var.name`; `index` is the starting position of the sliding window.

The algorithm is highly optimized by blocking the computations to exploit the high-speed memory instead of disk.

Value

A vector or list of values.

Author(s)

Xiuwen Zheng

See Also

[seqSetFilter](#), [seqGetData](#), [seqApply](#)

Examples

```
# the file of GDS
gds.fn <- seqExampleFileName("gds")
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

# display
(f <- seqOpen(gds.fn))

# get 'sample.id'
```

```

(samp.id <- seqGetData(f, "sample.id"))
# "NA06984" "NA06985" "NA06986" ...

# get 'variant.id'
head(variant.id <- seqGetData(f, "variant.id"))

# set sample and variant filters
set.seed(100)
seqSetFilter(f, sample.id=samp.id[seq(2, 16, 2)],
variant.id=sample(variant.id, 10))

# apply a function via a sliding window over variants
seqSlidingWindow(f, c(qual="annotation/id"), win.size=3,
FUN = function(x) {
# x is a list with 'win.size' elements
print(x)
}, as.is="none")

# apply a function via a sliding window over variants
seqSlidingWindow(f, c(qual="annotation/id"), win.size=3,
FUN = function(x) {
cat(unlist(x), sep="\t"); cat("\n")
}, as.is="none")

# apply a function via a sliding window over variants
seqSlidingWindow(f, c(geno="genotype", phase="phase", qual="annotation/id"),
FUN = function(index, x) {
cat("Window ", index, ":\n", sep="")
print(x)
},
win.size=3, as.is="none", var.index="relative")

# apply a function via a sliding window over variants
seqSlidingWindow(f, "genotype", win.size=4,
FUN = function(index, x) {
z <- unlist(lapply(x, function(z) mean(z, na.rm=TRUE)))
cat("Window ", index, ", starting from Variant ", index,
"\n    ", format(round(z,3), nsmall=3, width=8), "\n", sep="")
},
as.is="none", var.index="relative")

# close the GDS file
seqClose(f)

```

Description

Get the summary of a sequencing GDS file

Usage

```
seqSummary(gdsfile, varname=NULL, check=c("check", "full.check", "none"),
  verbose=TRUE)
```

Arguments

gdsfile	a SeqVarGDSClass object
varname	if NULL, check the whole GDS file; or a character specifying variable name, and return a description of that variable. See details.
check	should be one of "check", "full.check", "none"
verbose	if TRUE, display information

Details

If `check = "check"`, this function performs regular checking: dimensions of variables, etc. If `check = "full.check"`, it performs more checking: unique sample id, unique variant id, whether genotypic data are in a valid range or not, etc.

Value

If `varname = NULL`, then return a list:

filename	the file name
sequence.variant.format	the sequencing format in GDS
num.of.sample	the number of samples
num.of.variant	the number of variants
info	the description of the INFO field: var.name, number, type and description
format	the description of the FORMAT field: var.name, number, type and description

If `varname = "genotype"` or `"phase"`, then return a list:

dim	dim[1] – ploidy, dim[2] – the number of samples, dim[3] – the number of variants
seldim	seldim[1] – the number of selected samples, seldim[2] – the number of selected variants

Author(s)

Xiuwen Zheng

See Also

[seqGetData](#), [seqApply](#)

Examples

```
(gds.fn <- seqExampleFileName("gds"))
# or gds.fn <- "C:/YourFolder/Your_GDS_File.gds"

seqSummary(gds.fn)

seqSummary(gds.fn, "genotype")

#####
# display
f <- seqOpen(gds.fn)

# get 'sample.id'
samp.id <- seqGetData(f, "sample.id")
# get 'variant.id'
variant.id <- seqGetData(f, "variant.id")

# set sample and variant filters
seqSetFilter(f, sample.id=samp.id[c(2,4,6,8,10)])
set.seed(100)
seqSetFilter(f, variant.id=sample(variant.id, 10))

seqSummary(f, "genotype")

# close a GDS file
seqClose(f)
```

seqTranspose

Transpose Data Array

Description

Transpose data array or matrix for possibly higher-speed access

Usage

```
seqTranspose(gdsfile, var.name, compress=NULL, verbose=TRUE)
```

Arguments

gdsfile	a SeqVarGDSClass object
var.name	the variable name with '/' as a separator
compress	the compression option used in add.gdsn ; or determine automatically if NULL
verbose	if TRUE, show information

Details

It is designed for possibly higher-speed access. More details will be provided in the future version.

Value

None.

Author(s)

Xiuwen Zheng

See Also

[seqGetData](#), [seqApply](#)

Examples

```
# the file name of VCF
(vcf.fn <- seqExampleFileName("vcf"))
# or vcf.fn <- "C:/YourFolder/Your_VCF_File.vcf"

# convert
seqVCF2GDS(vcf.fn, "tmp.gds")

# list the structure of GDS variables
f <- seqOpen("tmp.gds", FALSE)
f

seqTranspose(f, "genotype/data")
f

# the original array
index.gdsn(f, "genotype/data")
# the transposed array
index.gdsn(f, "genotype/~data")

# close
seqClose(f)
unlink("tmp.gds")
```

SeqVarGDSClass

SeqVarGDSClass

Description

A SeqVarGDSClass object provides access to a GDS file containing Variant Call Format (VCF) data. It extends [gds.class](#).

Details

A sequencing GDS file is created from a VCF file with [seqVCF2GDS](#). This file can be opened with [seqOpen](#) to create a SeqVarGDSClass object.

Accessors

In the following code snippets `x` is a `SeqVarGDSClass` object.

`granges(x)`: Returns the chromosome and position of variants as a `GRanges` object. Names correspond to the `variant.id`.

`ref(x)`: Returns the reference alleles as a `DNAStrngSet`.

`alt(x)`: Returns the alternate alleles as a `DNAStrngSetList`.

`filt(x)`: Returns the filter data.

`qual(x)`: Returns the quality scores.

Other data can be accessed with `seqGetData`.

Coercion methods

In the following code snippets `x` is a `SeqVarGDSClass` object.

`asVCF(x, info=NULL, geno=NULL)`: Coerces a `SeqVarGDSClass` object to a `VCF-class` object. Row names correspond to the `variant.id`. `info` and `geno` specify the 'INFO' and 'GENO' (FORMAT) fields to return, respectively. If not specified, all fields are returned; if 'NA' no fields are returned. Use `seqSetFilter` prior to calling `asVCF` to specify samples and variants to return.

Author(s)

Xiuwen Zheng, Stephanie Gogarten

See Also

[gds.class](#), [seqVCF2GDS](#), [seqOpen](#), [seqGetData](#), [seqSetFilter](#), [seqClose](#)

Examples

```
gds <- seqOpen(seqExampleFileName("gds"))
gds

## sample ID
head(seqGetData(gds, "sample.id"))

## variants
granges(gds)

## alleles as comma-separated character strings
head(seqGetData(gds, "allele"))

## alleles as DNAStrngSet or DNAStrngSetList
ref(gds)
alt(gds)

## genotype
geno <- seqGetData(gds, "genotype")
```

```

dim(geno)
## dimensions are: allele, sample, variant
geno[1,1:10,1:5]

## rsID
head(seqGetData(gds, "annotation/id"))

## alternate allele count
head(seqGetData(gds, "annotation/info/AC"))

## individual read depth
depth <- seqGetData(gds, "annotation/format/DP")
names(depth)
## VCF header defined DP as variable-length data
table(depth$length)
## all length 1, so depth$data should be a sample by variant matrix
dim(depth$data)
depth$data[1:10,1:5]

seqClose(gds)

```

seqVCF.Header

Parse the header of a VCF file

Description

Parse the header of a VCF file

Usage

```
seqVCF.Header(vcf.fn)
```

Arguments

vcf.fn the file name of VCF

Details

The ID description contains four columns: ID – variable name; Number – the number of elements, see the webpage of the 1000 Genomes Project; Type – data type; Description – a variable description.

Value

Return a list (with a class name "SeqVCFHeaderClass", S3 object):

fileformat	the file format
info	the ID description in the INFO field
filter	the ID description in the FILTER field

format	the ID description in the FORMAT field
alt	the ID description in the ALT field
contig	the description in the contig field
assembly	the link of assembly
header	the other header lines
num.ploidy	the number of ploidy, two for humans

Author(s)

Xiuwen Zheng

References

<http://www.1000genomes.org/wiki/Analysis/VariantCallFormat/vcf-variant-call-format-version-41>

The variant call format and VCFtools. Danecek P, Auton A, Abecasis G, Albers CA, Banks E, DePristo MA, Handsaker RE, Lunter G, Marth GT, Sherry ST, McVean G, Durbin R; 1000 Genomes Project Analysis Group. *Bioinformatics*. 2011 Aug 1;27(15):2156-8. Epub 2011 Jun 7.

See Also

[seqVCF.SampID](#), [seqVCF2GDS](#)

Examples

```
# the file name of VCF
(vcf.fn <- seqExampleFileName("vcf"))
# or vcf.fn <- "C:/YourFolder/Your_VCF_File.vcf"

# get sample id
seqVCF.Header(vcf.fn)
```

seqVCF.SampID

Get the sample IDs of a VCF file

Description

Return the sample IDs of a VCF file

Usage

```
seqVCF.SampID(vcf.fn)
```

Arguments

vcf.fn the file name of VCF

Author(s)

Xiuwen Zheng

References

<http://www.1000genomes.org/wiki/Analysis/VariantCallFormat/vcf-variant-call-format-version-41>

The variant call format and VCFtools. Danecek P, Auton A, Abecasis G, Albers CA, Banks E, DePristo MA, Handsaker RE, Lunter G, Marth GT, Sherry ST, McVean G, Durbin R; 1000 Genomes Project Analysis Group. *Bioinformatics*. 2011 Aug 1;27(15):2156-8. Epub 2011 Jun 7.

See Also

[seqVCF.Header](#), [seqVCF2GDS](#)

Examples

```
# the file name of VCF
(vcf.fn <- seqExampleFileName("vcf"))
# or vcf.fn <- "C:/YourFolder/Your_VCF_File.vcf"

# get sample id
seqVCF.SampID(vcf.fn)
```

seqVCF2GDS

Reformat VCF files

Description

Reformat Variant Call Format (VCF) files.

Usage

```
seqVCF2GDS(vcf.fn, out.fn, header = NULL, genotype.var.name = "GT",
  compress.option = seqCompress.Option(), info.import=NULL, fmt.import=NULL,
  ignore.chr.prefix="chr", raise.error=TRUE, verbose=TRUE)
```

Arguments

vcf.fn	the file name(s) of VCF format
out.fn	the file name of output GDS file
header	if NULL, header is set to be seqVCF.Header (vcf.fn)
genotype.var.name	the ID for genotypic data in the FORMAT column; "GT" by default, VCFv4.0
compress.option	specify the compression options, by default seqCompress.Option

<code>info.import</code>	characters, the variable name(s) in the INFO field for import; or NULL for all variables
<code>fmt.import</code>	characters, the variable name(s) in the FORMAT field for import; or NULL for all variables
<code>ignore.chr.prefix</code>	a vector of character, indicating the prefix of chromosome which should be ignored, like "chr"; it is not case-sensitive
<code>raise.error</code>	TRUE: throw an error if numeric conversion fails; FALSE: get missing value if numeric conversion fails
<code>verbose</code>	if TRUE, show information

Details

GDS – Genomic Data Structures used for storing genetic array-oriented data, and the file format used in the [gdsfmt](#) package.

VCF – The Variant Call Format (VCF), which is a generic format for storing DNA polymorphism data such as SNPs, insertions, deletions and structural variants, together with rich annotations.

If there are more than one files in `vcf.fn`, `seqVCF2GDS` will merge all dataset together if they contain the same samples. It is useful to merge genomic variants if VCF data are divided by chromosomes.

Value

Return the file name of GDS format with an absolute path.

Author(s)

Xiuwen Zheng

References

The variant call format and VCFtools. Danecek P, Auton A, Abecasis G, Albers CA, Banks E, DePristo MA, Handsaker RE, Lunter G, Marth GT, Sherry ST, McVean G, Durbin R; 1000 Genomes Project Analysis Group. *Bioinformatics*. 2011 Aug 1;27(15):2156-8. Epub 2011 Jun 7.

<http://corearray.sourceforge.net/>

See Also

[seqVCF.Header](#), [seqCompress.Option](#), [seqGDS2VCF](#)

Examples

```
# the file name of VCF
vcf.fn <- seqExampleFileName("vcf")
# or vcf.fn <- "C:/YourFolder/Your_VCF_File.vcf"

# convert
seqVCF2GDS(vcf.fn, "tmp.gds")
```

```
# display
(f <- seqOpen("tmp.gds"))
seqClose(f)
```

```
# convert without the INFO fields
seqVCF2GDS(vcf.fn, "tmp.gds", info.import=character(0))
```

```
# display
(f <- seqOpen("tmp.gds"))
seqClose(f)
```

```
# convert without the INFO fields
seqVCF2GDS(vcf.fn, "tmp.gds",
  info.import=character(0), fmt.import=character(0))
```

```
# display
(f <- seqOpen("tmp.gds"))
seqClose(f)
```

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