

# Package ‘minet’

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**Title** Mutual Information NETWORKs

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**Author** Patrick E. Meyer, Frederic Lafitte, Gianluca Bontempi

**Description**

This package implements various algorithms for inferring mutual information networks from data.

**Maintainer** Patrick E. Meyer <software@meyerp.com>

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**Depends** infotheo

**URL** <http://minet.meyerp.com>

**biocViews** Microarray, GraphsAndNetworks, NetworkAnalysis,NetworkInference

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aracne

*Algorithm for the Reconstruction of Accurate Cellular Networks*

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## Description

This function takes the mutual information matrix as input in order to return the inferred network according to the Aracne algorithm. This algorithm applies the data processing inequality to all triplets of nodes in order to remove the least significant edge in each triplet.

## Usage

```
aracne( mim, eps=0 )
```

## Arguments

mim	A square matrix whose $i,j$ th element is the mutual information between variables $X_i$ and $X_j$ - see <a href="#">build.mim</a> .
eps	Numeric value indicating the threshold used when removing an edge : for each triplet of nodes $(i,j,k)$ , the weakest edge, say $(ij)$ , is removed if its weight is below $\min\{(ik),(jk)\}-\text{eps}$ - see references.

## Details

The Aracne procedure starts by assigning to each pair of nodes a weight equal to their mutual information. Then, the weakest edge of each triplet is interpreted as an indirect interaction and is removed if the difference between the two lowest weights is above a threshold `eps`.

## Value

`aracne` returns a matrix which is the weighted adjacency matrix of the network. In order to display the network, load the package `Rgraphviz` and use the following command:  
`plot( as( returned.matrix , "graphNEL" ) )`

## References

Adam A. Margolin, Ilya Nemenman, Katia Basso, Chris Wiggins, Gustavo Stolovitzky, Riccardo Dalla Favera, and Andrea Califano. Aracne : An algorithm for the reconstruction of gene regulatory networks in a mammalian cellular context. BMC Bioinformatics, 2006.

Patrick E. Meyer, Frederic Lafitte and Gianluca Bontempi. `minet`: A R/Bioconductor Package for Inferring Large Transcriptional Networks Using Mutual Information. BMC Bioinformatics, Vol 9, 2008.

## See Also

[build.mim](#), [clr](#), [mrnet](#), [mrnetb](#)

## Examples

```
data(syn.data)
mim <- build.mim(syn.data,estimator="spearman")
net <- aracne(mim)
```

build.mim

*Build Mutual Information Matrix***Description**

build.mim takes the dataset as input and computes the mutual information between all pair of variables according to the mutual information estimator. The results are saved in the mutual information matrix (MIM), a square matrix whose (i,j) element is the mutual information between variables  $X_i$  and  $X_j$ .

**Usage**

```
build.mim(dataset, estimator = "spearman", disc = "none", nbins = sqrt(NROW(dataset)))
```

**Arguments**

dataset	data.frame containing gene expression data or any dataset where columns contain variables/features and rows contain outcomes/samples.
estimator	The name of the entropy estimator to be used. The package implements four estimators for discrete data: "mi.empirical", "mi.mm", "mi.shrink", "mi.sg" and three estimators based on correlation (can be used with continuous data): "pearson", "spearman", "kendall"(default:"spearman") - see details.
disc	The name of the discretization method to be used with one of the discrete estimators: "none", "equalfreq", "equalwidth" or "globalequalwidth" (default : "none") - see infotheo package.
nbins	Integer specifying the number of bins to be used for the discretization if disc is different from "none". By default the number of bins is set to $\sqrt{m}$ where m is the number of samples.

**Details**

- "mi.empirical" : This estimator computes the entropy of the empirical probability distribution.
- "mi.mm" : This is the Miller-Madow asymptotic bias corrected empirical estimator.
- "mi.shrink" : This is a shrinkage estimate of the entropy of a Dirichlet probability distribution.
- "mi.sg" : This is the Schurmann-Grassberger estimate of the entropy of a Dirichlet probability distribution.
- "pearson" : This computes mutual information for normally distributed variable.
- "spearman" : This computes mutual information for normally distributed variable using Spearman's correlation instead of Pearson's correlation.
- "kendall" : This computes mutual information for normally distributed variable using Kendall's correlation instead of Pearson's correlation.

**Value**

build.mim returns the mutual information matrix.

**Author(s)**

Patrick E. Meyer, Frederic Lafitte, Gianluca Bontempi

## References

Patrick E. Meyer, Frederic Lafitte, and Gianluca Bontempi. *minet: A R/Bioconductor Package for Inferring Large Transcriptional Networks Using Mutual Information*. BMC Bioinformatics, Vol 9, 2008.

J. Beirlant, E. J. Dudewica, L. Gyofi, and E. van der Meulen. *Nonparametric entropy estimation : An overview*. Journal of Statistics, 1997.

Jean Hausser. *Improving entropy estimation and the inference of genetic regulatory networks*. Master thesis of the National Institute of Applied Sciences of Lyon, 2006.

## See Also

[clr](#), [aracne](#), [mrnet](#), [mrnetb](#)

## Examples

```
data(syn.data)
mim <- build.mim(syn.data,estimator="spearman")
```

---

clr

*Context Likelihood or Relatedness Network*

---

## Description

clr takes the mutual information matrix as input in order to return the inferred network - see details.

## Usage

```
clr( mim )
```

## Arguments

**mim**                    A square matrix whose i,j th element is the mutual information between variables  $X_i$  and  $X_j$  - see [build.mim](#).

## Details

The CLR algorithm is an extension of relevance network. Instead of considering the mutual information  $I(X_i; X_j)$  between features  $X_i$  and  $X_j$ , it takes into account the score  $\sqrt{z_i^2 + z_j^2}$ , where

$$z_i = \max \left\{ 0, \frac{I(X_i; X_j) - \mu_i}{\sigma_i} \right\}$$

and  $\mu_i$  and  $\sigma_i$  are, respectively, the mean and the standard deviation of the empirical distribution of the mutual information values  $I(X_i; X_k)$ ,  $k=1, \dots, n$ .

## Value

clr returns a matrix which is the weighted adjacency matrix of the network. In order to display the network, load the package Rgraphviz and use the following command `plot( as( returned.matrix, "graphNEL" ) )`

## References

Jeremiah J. Faith, Boris Hayete, Joshua T. Thaden, Ilaria Mogno, Jamey Wierzbowski, Guillaume Cottarel, Simon Kasif, James J. Collins, and Timothy S. Gardner. Large-scale mapping and validation of escherichia coli transcriptional regulation from a compendium of expression profiles. PLoS Biology, 2007.

## See Also

[build.mim](#), [aracne](#), [mrnet](#), [mrnetb](#)

## Examples

```
data(syn.data)
mim <- build.mim(syn.data,estimator="spearman")
net <- clr(mim)
```

---

minet

*Mutual Information Network*

---

## Description

For a given dataset, minet infers the network in two steps. First, the mutual information between all pairs of variables in dataset is computed according to the estimator argument. Then the algorithm given by method considers the estimated mutual informations in order to build the network. This package is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

## Usage

```
minet(dataset, method="mrnet", estimator="spearman", disc="none", nbins=sqrt(NROW(dataset)))
```

## Arguments

dataset	data.frame where columns contain variables/features and rows contain outcomes/samples.
method	The name of the inference algorithm : "clr", "aracne", "mrnet" or "mrnetb" (default: "mrnet") - see references.
estimator	The name of an entropy estimator (or correlation) to be used for mutual information computation ("mi.empirical", "mi.mm", "mi.shrink", "mi.sg", "pearson", "spearman", "kendall"), (default: "spearman") . - see <a href="#">build.mim</a> .
disc	The name of the discretization method to be used, if required by the estimator : "none", "equalfreq", "equalwidth" or "globalequalwidth" (default : "none") - see infotheo package.
nbins	Integer specifying the number of bins to be used for the discretization if disc is set properly. By default the number of bins is set to $\sqrt{N}$ where N is the number of samples.

## Value

minet returns a matrix which is the weighted adjacency matrix of the network. The weights range from 0 to 1 and can be seen as a confidence measure on the presence of the arcs. In order to display the network, load the package Rgraphviz and use the following command:  
`plot( as(returned.matrix , "graphNEL") )`

**Author(s)**

Patrick E. Meyer, Frederic Lafitte, Gianluca Bontempi

**References**

Patrick E. Meyer, Frederic Lafitte, and Gianluca Bontempi. *minet: A R/Bioconductor Package for Inferring Large Transcriptional Networks Using Mutual Information*. BMC Bioinformatics, Vol 9, 2008.

**See Also**

[build.mim](#), [clr](#), [mrnet](#), [mrnetb](#), [aracne](#)

**Examples**

```
data(syn.data)
net1 <- minet( syn.data )
net2 <- minet( syn.data, estimator="pearson" )
net3 <- minet( syn.data, method="clr" )
```

---

 mrnet

---

*Maximum Relevance Minimum Redundancy*


---

**Description**

mrnet takes the mutual information matrix as input in order to infer the network using the maximum relevance/minimum redundancy feature selection method - see details.

**Usage**

```
mrnet(mim)
```

**Arguments**

mim                    A square matrix whose i,j th element is the mutual information between variables  $X_i$  and  $X_j$  - see [build.mim](#).

**Details**

The MRNET approach consists in repeating a MRMR feature selection procedure for each variable of the dataset. The MRMR method starts by selecting the variable  $X_i$  having the highest mutual information with the target  $Y$ . In the following steps, given a set  $\mathcal{S}$  of selected variables, the criterion updates  $\mathcal{S}$  by choosing the variable  $X_k$  that maximizes  $I(X_k; Y) - \frac{1}{|\mathcal{S}|} \sum_{X_i \in \mathcal{S}} I(X_k; X_i)$

The weight of each pair  $X_i, X_j$  will be the maximum score between the one computed when  $X_i$  is the target and the one computed when  $X_j$  is the target.

**Value**

mrnet returns a matrix which is the weighted adjacency matrix of the network. In order to display the network, load the package Rgraphviz and use the following command:  

```
plot( as( returned.matrix , "graphNEL" ) )
```

**Author(s)**

Patrick E. Meyer, Frederic Lafitte, Gianluca Bontempi

**References**

Patrick E. Meyer, Kevin Kontos, Frederic Lafitte and Gianluca Bontempi. Information-theoretic inference of large transcriptional regulatory networks. *EURASIP Journal on Bioinformatics and Systems Biology*, 2007.

Patrick E. Meyer, Frederic Lafitte and Gianluca Bontempi. minet: A R/Bioconductor Package for Inferring Large Transcriptional Networks Using Mutual Information. *BMC Bioinformatics*, Vol 9, 2008.

H. Peng, F.long and C.Ding. Feature selection based on mutual information: Criteria of max-dependency, max relevance and min redundancy. *IEEE transaction on Pattern Analysis and Machine Intelligence*, 2005.

**See Also**

[build.mim](#), [clr](#), [aracne](#), [mrnetb](#)

**Examples**

```
data(syn.data)
mim <- build.mim(syn.data, estimator="spearman")
net <- mrnetb(mim)
```

---

mrnetb

*Maximum Relevance Minimum Redundancy Backward*

---

**Description**

mrnetb takes the mutual information matrix as input in order to infer the network using the maximum relevance/minimum redundancy criterion combined with a backward elimination and a sequential replacement - see references. This method is a variant of mrnet.

**Usage**

```
mrnetb(mim)
```

**Arguments**

mim                    A square matrix whose  $i,j$  th element is the mutual information between variables  $X_i$  and  $X_j$  - see [build.mim](#).

**Value**

mrnetb returns a matrix which is the weighted adjacency matrix of the network. In order to display the network, load the package Rgraphviz and use the following command:  
`plot( as( returned.matrix , "graphNEL" ) )`

**Author(s)**

Patrick E. Meyer

## References

Patrick E. Meyer, Daniel Marbach, Sushmita Roy and Manolis Kellis. Information-Theoretic Inference of Gene Networks Using Backward Elimination. The 2010 International Conference on Bioinformatics and Computational Biology.

Patrick E. Meyer, Kevin Kontos, Frederic Lafitte and Gianluca Bontempi. Information-theoretic inference of large transcriptional regulatory networks. EURASIP Journal on Bioinformatics and Systems Biology, 2007.

## See Also

[build.mim](#), [clr](#), [mrnet](#), [aracne](#)

## Examples

```
data(syn.data)
mim <- build.mim(syn.data, estimator="spearman")
net <- mrnetb(mim)
```

---

syn.data

*Simulated Gene Expression Data*

---

## Description

Dataset containing 100 samples and 50 genes generated by the publicly available SynTReN generator using a yeast source network - see [syn.net](#)

## Usage

```
data(syn.data)
```

## Format

syn.data is a data frame containing 100 rows and 50 columns. Each row contains a microarray experiment and each column contains a gene.

## Source

SynTReN 1.1.3 with source network : yeast\\_nn.sif

## References

Tim Van den Bulcke, Koenraad Van Leemput, Bart Naudts, Piet van Remortel, Hongwu Ma, Alain Verschoren, Bart De Moor, and Kathleen Marchal. Syntren : a generator of synthetic gene expression dataset for design and analysis of structure learning algorithms. BMC Bioinformatics, 2006.

## Examples

```
data(syn.data)
data(syn.net)
mim <- build.mim(syn.data, estimator="spearman")
inferred.net <- mrnet(mim)
max(fscores(validate( inferred.net, syn.net )))
```



---

`syn.net`*SynTReN Source Network*

---

**Description**

This is the true underlying network used to generate the dataset loaded by `data(syn.data)` - see [syn.data](#).

**Usage**

```
data(syn.net)
```

**Format**

`syn.net` is a boolean adjacency matrix representing an undirected graph of 50 nodes.

**Source**

`syn.net` is the "yeast\\_nn.sif" source network from the SynTReN generator where all the variables/nodes not in `syn.data` were removed.

**References**

Tim Van den Bulcke, Koenraad Van Leemput, Bart Naudts, Piet van Remortel, Hongwu Ma, Alain Verschoren, Bart De Moor, and Kathleen Marchal. Syntren : a generator of synthetic gene expression dataset for design and analysis of structure learning algorithms. BMC Bioinformatics, 2006.

**Examples**

```
data(syn.data)
data(syn.net)
mim <- build.mim(syn.data,estimator="spearman")
inferred.net <- mrnet(mim)
max(fscores(validate( inferred.net, syn.net )))
```

---

`validate`*Inference Validation*

---

**Description**

`validate` compares the inferred network to the true underlying network for several threshold values and appends the resulting confusion matrices to the returned object.

**Usage**

```
validate(inet,tnet)
```

**Arguments**

inet	This is the inferred network, a data.frame or matrix obtained by one of the functions <a href="#">minet</a> , <a href="#">aracne</a> , <a href="#">clr</a> or <a href="#">mrnet</a> .
tnet	The true underlying network. This network must have the same size and variable names as inet.

**Details**

The first network `inet` is compared to the true underlying network, `tnet`, in order to compute a confusion (adjacency) matrix. All the confusion matrices, obtained with different threshold values, are appended to the returned object. In the end the `validate` function returns a data.frame containing `steps+1` confusion matrices.

**Value**

`validate` returns a data.frame with four columns named `thrsh`, `tp`, `fp`, `fn`. These values are computed for each of the steps thresholds. Thus each row of the returned object contains the confusion matrix for a different threshold.

**See Also**

[minet](#), [vis.res](#)

**Examples**

```
data(syn.data)
data(syn.net)
inf.net <- mrnet(build.mim(syn.data, estimator="spearman"))
table <- validate( inf.net, syn.net )
table <- validate( inf.net, syn.net )
```

---

vis.res

*Visualize Results*

---

**Description**

A group of functions to plot precision-recall and ROC curves and to compute f-scores from the data.frame returned by the [validate](#) function.

**Usage**

```
pr(table)
rates(table)
fscores(table, beta=1)
show.pr(table,device=-1,...)
show.roc(table,device=-1,...)
auc.roc(table)
auc.pr(table)
```

## Arguments

table	This is the data.frame returned by the validate function where columns contain TP,FP,TN,FN values (confusion matrix) as well as the threshold value used - see <a href="#">validate</a> .
beta	Numeric used as the weight of the recall in the f-score formula - see details. The default value of this argument is 1, meaning precision as important as recall.
device	The device to be used. This parameter allows the user to plot precision-recall and receiver operating characteristic curves for various inference algorithms on the same plotting window - see examples.
...	arguments passed to plot

## Details

A confusion matrix contains FP,TP,FN,FP values.

- "true positive rate"  $tpr = \frac{TP}{TN+TP}$
- "false positive rate"  $fpr = \frac{FP}{FN+FP}$
- "precision"  $p = \frac{TP}{FP+TP}$
- "recall"  $r = \frac{TP}{TP+FN}$
- "f-beta-score"  $F_\beta = (1 + \beta) \frac{pr}{r + \beta p}$

## Value

The function `show.roc` (`show.pr`) plots the ROC-curve (PR-curve) and returns the device associated with the plotting window.

The function `auc.roc` (`auc.pr`) computes the area under the ROC-curve (PR-curve) using the trapezoidal approximation.

The function `pr` returns a data.frame where `steps` is the number of thresholds used in the validation process. The first column contains precisions and the second recalls - see details.

The function `rates` also returns a data.frame where the first column contains true positive rates and the second column false positive rates - see details.

The function `fscores` returns fscores according to the confusion matrices contained in the 'table' argument - see details.

## References

Patrick E. Meyer, Frederic Lafitte, and Gianluca Bontempi. `minet`: A R/Bioconductor Package for Inferring Large Transcriptional Networks Using Mutual Information. BMC Bioinformatics, Vol 9, 2008.

## See Also

[validate](#), [plot](#)

**Examples**

```
data(syn.data)
data(syn.net)
# Inference
mr <- minet( syn.data, method="mrnet", estimator="spearman" )
ar <- minet( syn.data, method="aracne", estimator="spearman" )
clr <- minet( syn.data, method="clr", estimator="spearman" )
# Validation
mr.tbl <- validate(mr,syn.net)
ar.tbl <- validate(ar,syn.net)
clr.tbl <- validate(clr,syn.net)
# Plot PR-Curves
max(fscores(mr.tbl))
dev <- show.pr(mr.tbl, col="green", type="b")
dev <- show.pr(ar.tbl, device=dev, col="blue", type="b")
show.pr(clr.tbl, device=dev, col="red",type="b")
auc.pr(clr.tbl)
```

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